

COMPARATIVE CYTOTOXIC ACTIVITY OF CARBOPLATIN AND β -CRYPTOXANTHIN IN FREE AND LIPOSOMAL FORMS AGAINST BREAST CANCER CELL LINE

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The study of the effectiveness of the synthetic and natural anticarcinogenic compounds in liposomal form is urgent for their possible use in therapy. In this work, the alkylating agent carboplatin and the representative of carotenoids β -cryptoxanthin were used. The aim of the research was to study the toxicity of these compounds in free and liposomal forms against breast cancer MCF-7 cell line. According to DSC and FTIR data, when carboplatin or β -cryptoxanthin were added to liposomal bilayers, a single peak was observed indicating their mutual mixing. Integration of β -cryptoxanthin into bilayer was found to be more proper for the creation of PE acyl chains ordered and cooperative state. It was found that MCF-7 cells sensitivity was much higher to the free β -cryptoxanthin than to the free carboplatin with IC_{50} 42 and 235 $\mu\text{g/ml}$, respectively. The IC_{50} values for β -cryptoxanthin loaded into liposomes and for free carboplatin were similar. At the same time, no cytotoxic effect of carboplatin-loaded liposomes was observed. The data obtained allow proposing a possible antitumor treatment regimen where carboplatin is replaced by free β -cryptoxanthin or its liposomal form to increase the effectiveness of breast cancer therapy.

Key words: β -cryptoxanthin, carboplatin, liposomes, DSC, FTIR, MCF-7 cancer cell line, cytotoxicity.

Cancer is characterized by the uncontrolled growth and proliferation of abnormal cells that can invade or spread to other parts of the body. If the spread is not managed, then it can result in death. Treatments include surgery, radiation, chemotherapy, hormone therapy, immune and targeted therapy [1]

Breast cancer is the second leading carcinogenic disease as the most common non-cutaneous malignancy among women. Options for treating breast cancer are limited and linked to toxicity. Emerging nanotechnologies have exhibited the potential to treat or target breast cancer. Over the years, different lipid nanoparticles for breast cancer therapy have been created, namely liposomes, solid lipid nanoparticles, nanostructured lipid carriers, and lipid polymer hybrid nanoparticles [2, 3].

All hydrophilic and lipophilic drugs can be encapsulated and protected from degradation by liposomes, the vesicles of phospholipid bilayers. The liposomal similarity to the membrane bilayer core makes liposomes a very useful tool for examining the importance of drug-membrane interactions with anticancer.

By prolonging biological half-life or reducing toxicity, liposomes have been used to enhance the therapeutic index of new or established drugs [4, 5]. Liposomal clinical applications are a large area of research where cancer therapy is the area of greatest impact [6, 7].

Carboplatin is one of the platinum anti-cancer compounds and its parent compound is cisplatin with a carboxy-cyclobutane moiety instead of the chloride atoms which makes it more stable and perhaps less

toxic than cisplatin. Carboplatin and cisplatin act as alkylating agents causing cross-linkage between and within DNA strands, resulting in inhibition of DNA, RNA, and protein synthesis and triggering programmed cell death, often in rapidly dividing cells. It causes the development of free radicals or reactive oxygen species. Free radicals cause cellular damage in many ways due to their high chemical reactivity. DNA damage, which can lead to a number of pathological conditions, including cancer, is among the most damaging effects of free radicals. [8]. Cisplatin and carboplatin are effective chemotherapy drugs commonly used to treat solid tumors such as cancer of the testicles, ovaries, cervix, lungs, head and neck, esophagus, and bladder [9]. Cisplatin and carboplatin have been investigated in metastatic breast cancer as monotherapies and in combination with other chemotherapeutic agents. [10].

Carotenoids are among the bioactive substances that have a potential impact on cancer risk and progression and have been the focus of numerous investigations [11, 12]. Carotenoids are naturally occurring organic compounds in plants and photosynthetic organisms such as algae [13, 14]. Present reports suggest they have identified > 600 carotenoids. However, as a result of selective digestive tract uptake, only 14 carotenoids with their metabolites were found in human plasma and peripheral tissues [15].

Several studies have shown that diets rich in vegetables and fruit can reduce the risk of multiple chronic diseases, including cancer, cardiovascular diseases, and diabetes [16]. In addition to β -cryptoxanthin, various carotenoids show more potent activity to suppress the carcinogenic process. Evidence shows that β -cryptoxanthin has a potent antioxidant ability *in vitro* and can defend human cells from oxidative damage, contributing to inflammation suppression, even serving as a scavenger of free radicals, and preventing biomolecules such as lipids, proteins, and nucleic acids from oxidative damage. [17-19]. Carotenoids have numerous

pathways to exhibit anticancer and cancer prevention mechanisms, including antioxidant and direct antiproliferative effects (induction of apoptosis) and inhibition of oncogene expression and angiogenesis [20].

To our knowledge, no previous studies on the interaction of β -cryptoxanthin or carboplatin with phospholipids were performed from the perspective of the thermotropic phase behavior of phospholipids and to detect the changes of acyl chain conformations and characteristic PO_2^- bands in the polar heads of phospholipids. To date, the anti-proliferative activity of β -cryptoxanthin in free and nanoliposomal forms towards MCF-7 (Breast cancer cell line) has not yet been studied.

The objective of this study was to determine whether free or integrated β -cryptoxanthin in liposomes could inhibit the proliferation of human MCF-7 cell breast cancer by detecting the possible effects of these compounds on the cell death of human MCF-7 cell line carcinoma compared to free carboplatin and its conjugation with liposomes. The formulated β -cryptoxanthin or carboplatin/liposomal conjugate was characterized by analytical techniques to evaluate the size, size distribution, thermotropic and conformation changes of the formulated β -cryptoxanthin or carboplatin/liposomal conjugate along with *in vitro* possible cytotoxicity against MCF-7 (breast cancer cell line).

Material and Methods

Chemicals. β -cryptoxanthin was purified from natural sources and verified for purity by TLC and HPLC. The structure of β -cryptoxanthin was confirmed from spectral data and elemental analysis. The molecular weight of β -cryptoxanthin is 552.85. Carboplatin with a molecular weight of 371.25, was purchased from Asta (Germany). The molecular structure of β -cryptoxanthin and Carboplatin are shown in Fig. 1. Ethanol was of analytical grade and was bought from DaeJung

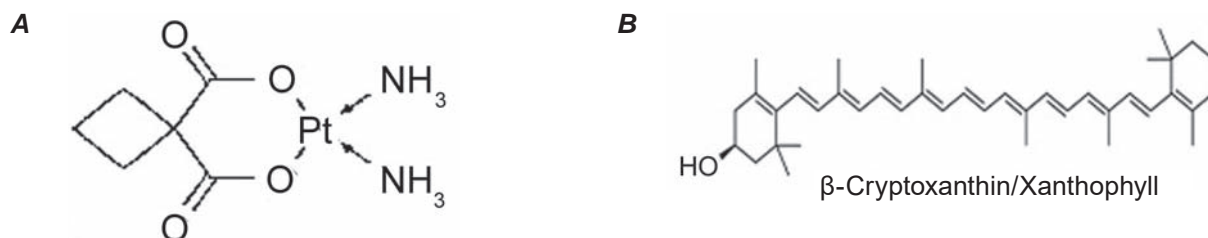


Fig. 1. The chemical structure of carboplatin (A) and β -cryptoxanthin (B)

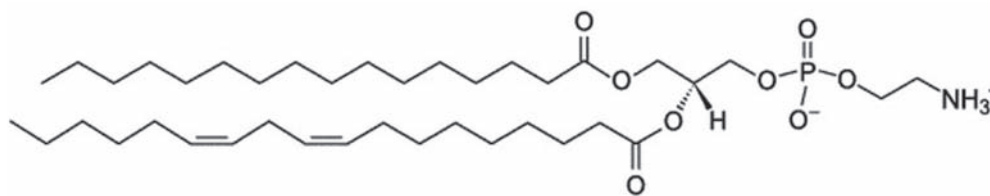


Fig. 2. Schematic chemical structure of *L*- α -phosphatidylethanolamine

Chemicals (Seohaean-ro, Gyeonggi-do, Korea). *L*- α -Phosphatidylethanolamine (PE) from sheep brain with a molecular weight of 691.515 of purity $\geq 98\%$ bought from Sigma (ST. Louis, Mo, USA) is presented in (Figure 2), Bought from CDH, New Delhi, India, Tris base in powder form, the molecular weight of 121.1. Human tumor cell line MCF 7 was obtained frozen in liquid nitrogen (-180°C) from the American Type Culture Collection. The tumor cell lines were maintained in the National Cancer Institute, Cairo, Egypt, by serial sub-culturing. Dimethylsulphoxide (DMSO), RPMI-1640 medium, Sodium bicarbonate, Trypan blue: An isotonic solution of 0.05% trypan blue in normal saline, Fetal bovine serum (FBS), Penicillin/Streptomycin, Trypsin, Acetic acid, Sulphorhodamine-B (SRB), 0.4% SRB dissolved in 1% acetic acid, Trichloroacetic acid (TCA), 100% isopropanol were obtained from (Sigma Chemical Co., St. Louis, Mo, USA). Solutions were prepared in distilled ultra-pure water. All other reagents and solvents used in this work were of research-grade.

Liposome preparation. At molar ratio 7:2 of *L*- α -Phosphatidylethanolamine derived from sheep brain to carboplatin (LipoCarbo) or β -cryptoxanthin (LipoCrypto) using the method of [21], neutral liposomal multilamellar vesicles (MLVs) were prepared.

In a round bottom flask of capacity 50 ml, 20 mg of *L*- α -Phosphatidylethanolamine and 3.06 mg carboplatin (LipoCarbo) or 4.57 mg β -cryptoxanthin (LipoCrypto) of the drug powder were mixed. Then 20 ml of ethanol (EtOH) was added, and the flask was shaken until all lipids dissolved in the EtOH. For a few minutes, the solution was well shaken then intense vortexing took place to ensure full solvation. The organic solvent was phased out using a rotary evaporator under vacuum provided by a circulating water aspiration vacuum pump in a warm water bath (50°C) at 60 rpm to create a uniform thin film of lipid on the inner wall of the flask. With Tris buffer (pH 7.4 in 37°C) in a water bath at 50°C . The lipid film was hydrated in a water bath at 50°C for 15 min at 60 rpm to form multila-

mellar vesicles (MLV) of liposomes. Following the same method as described above using only aliquots of *L*- α -Phosphatidylethanolamine (Cephalin), control empty liposomes were prepared.

Size distribution and Zeta potential measurements. In Tris buffer (pH 7.4) at 25°C using a particle sizing system (Nanotracs Wave II, Microtracs, USA) for dynamic light scattering, the mean particle size, size distribution, and zeta potential of freshly prepared empty liposomes, LipoCarbo and LipoCrypto were calculated. The results were an average of three individual measurements.

DSC measurements. Using indium-calibrated differential scanning calorimetry (DSC) (model DSC-50, Shimadzu, Japan), the thermal properties of different lyophilized liposomal formulations were studied. On 5-mg samples sealed in standard aluminum pans the analyses are performed. The thermogram of each sample covers the $25\text{--}200^{\circ}\text{C}$ temperature range at a scanning rate of $3^{\circ}\text{C}/\text{min}$.

FTIR Spectroscopy. On a Jasco FT/IR-4100 spectrometer (Tokyo, Japan), FTIR spectra of lyophilized samples of empty *L*- α -Phosphatidylethanolamine (Cephalin) liposomes and those carboplatin (LipoCarbo) liposomes or β -cryptoxanthin (LipoCrypto) deposited in KBr discs are registered. Scanning was performed at room temperature, at a speed of 2 mm/s and a resolution of 4 cm^{-1} in the range $400\text{--}4,000\text{ cm}^{-1}$.

In-Vitro cytotoxicity assay. Breast cancer cell line (MCF-7) was grown in a humidified incubator below 5% CO_2 and 95% air at 37°C in RPMI 1640 media supplemented with penicillin (100 IU/ml), streptomycin ($100\mu\text{g}/\text{ml}$), and decomplexed fetal bovine serum (10%, v/v). Drug treatments were carried out on 96 well culture plates (growth assays). For triplicate experiments, drugs were added to the culture medium at a calculated final concentration. By SRB (Sulfo-RhodamineB-stain) assay, the extent of the cell viability and proliferation (cell numbers) were measured.

Cytotoxic activity of free carboplatin, free β -cryptoxanthin, carboplatin (LipoCarbo), or (LipoCrypto) β -cryptoxanthin-loaded liposomes was tested individually using the method of [22]. In order to permit adhesion of the cell to the wall of the plate MCF-7 cells were plated in 96-multi well plate (10^4 cells/well) for 24 h before treatment with the applied drugs. Different concentrations of free carboplatin, free β -cryptoxanthin, carboplatin (LipoCarbo) or (LipoCrypto) β -cryptoxanthin-loaded liposomes under test (50, 100, 200, 400, 800, 1000, 1200, 1400, and 1500 $\mu\text{g/ml}$) were applied to the cell monolayer triplicate wells were prepared for each dose in the laminar flow cabinet for more fertilization. Free carboplatin, LipoCarbo, and LipoCrypto were co-cultivated with monolayer cells separately in an atmosphere of 5% CO_2 for 48 h. Cells with Sulfo-Rhodamine-B stain were fixed, washed, and stained after 48 h. The excess stain was washed with acetic acid and the attached stain was recovered with Tris-EDTA buffer. In an ELISA reader (ELISA-TECAN-SUNRISE, Germany), color intensity was evaluated. The relation between cell viability percentage (surviving fraction) and drug concentration is plotted to find the cell viability curve for MCF-7 cancer cell line after the treatment of plain Carboplatin, free β -cryptoxanthin, LipoCarbo, and LipoCrypto.

The percentage of cell survival was calculated as follows:

$$\text{Survival fraction} = \frac{\text{O.D. (treated cells)}}{\text{O.D. (control cells)}}$$

The IC_{50} values (drug concentrations needed to inhibit cell growth by 50%). The experiment was repeated 3 times for each cell line.

Statistical analysis. All results are recorded as the mean \pm SD. Statistical analysis was performed by one-way variance analysis (ANOVA), where a commercially available software package (SPSS-17 for windows, SPSS Inc., Chicago, Illinois, USA), was used and the significance level was considered at $P < 0.05$.

Results and Discussion

Dynamic light scattering (DLS) is a technique that is used in particle size measurement. Nano drug delivery systems improve the bioavailability of the entrapped drug according to their size which matches cell dimensions.

Table 1 shows the particle size for each formulation of carboplatin or β -cryptoxanthin -loaded liposomes as compared to empty liposomes. As

Table 1. Particle size distribution measured by dynamic light scattering (DLS) for various liposomal formulations

Liposomal formulation	Average size vesicles, nm
Empty liposomes	177.10 ± 63.25
LipoCarbo	1128.0 ± 380.5
LipoCrypto	153.80 ± 11.17

can be observed from Table 1, the average particle size of the empty liposomal sample was concentrated around 177.10 ± 63.25 nm mean size diameter. The incorporation of carboplatin (LipoCarbo) into liposomes resulted significantly ($P < 0.05$) in an increase in the calculated mean size diameter of blank liposomes from 177.10 ± 63.25 nm to 1128.0 ± 380.5 nm. These findings show that carboplatin can be physically associated with carboplatin on the surface of liposomes and the carboplatin molecule tends to be hidden in the bilayer of lipids, which can clarify why the size is increased.

Upon the encapsulation of β -cryptoxanthin into liposomes (LipoCrypto), the mean vesicle sizes was decreased to be in the range of 153.80 ± 11.17 nm. These results indicate that the inclusion of β -cryptoxanthin into liposomes decreased the spacing between the adjacent bilayers resulting in the formation of liposomes smaller in size compared with the control ones. The reduction of particle size may be due to stronger β -cryptoxanthin interactions via hydrogen bonding with the lipid bilayer of liposomes. Within a smaller size range, liposomes favor the accumulation of drugs into certain target tissues. Besides, they also have good stability, thus displaying predictable drug-release rates.

The possible stability of the colloidal system is indicated by the magnitude of the zeta potential. If the zeta potential increases, there will be an increased repulsion between particles, leading to a more stable dispersion of colloids. If all suspended particles have a strong zeta potential that is negative or positive, they seem to repel each other, and the particles are not likely to join together. [23].

Following the findings of others, empty liposomes showed negative zeta potential (-31.05 ± 3.06 mV) [24-27]. β -cryptoxanthin-loaded liposomes (LipoCrypto) had higher negative zeta potential (-34.51 ± 3.33 mV) than blank liposomes and carboplatin (LipoCarbo) (-21.60 ± 2.99 mV) due

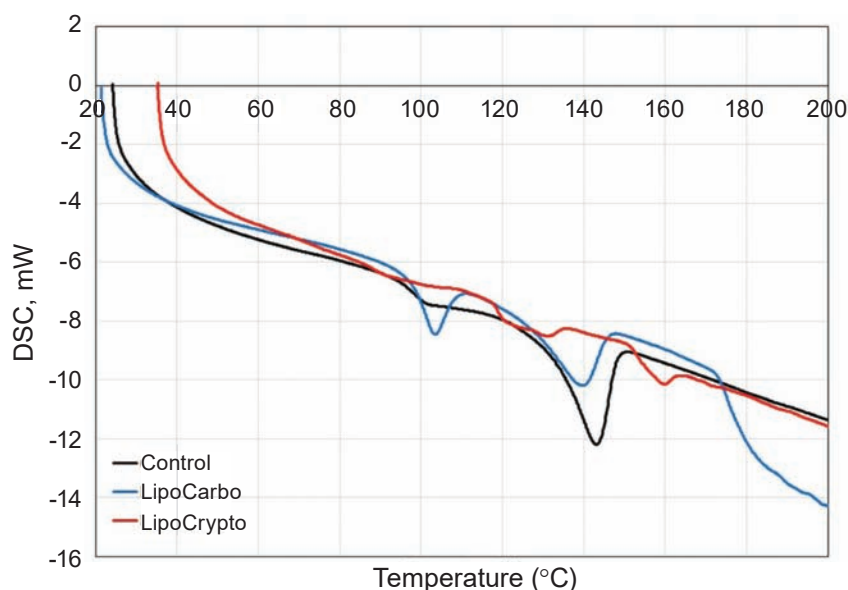


Fig. 3. DSC diagrams of liposomes are made of pure PE and liposomes doped with Carboplatin or β -cryptoxanthin

to the integration into the liposomal membranes. Within liposomal membranes, the incorporation of β -cryptoxanthin (LipoCrypto) appears to increase the density of negative charge and hence made the zeta potential negative.

The concept statement of DSC is that phospholipid vesicles undergo a reversible phase transition, under the effect of increasing temperature, from a 'gel' state to a 'liquid crystal' state. The pre-transition temperature (T_p) at which a transition from the gel phase to the rippled phase takes place is primarily related to the phospholipid polar region. Subsequently, the melting of the bilayer from the rippled phase to the liquid phase occurs at the main transition temperature (T_m). Changes in the lipid structure severely affect all the above phases.

When submitted for DSC analysis, pure L- α -Phosphatidylethanolamine (PE) vesicles upon dehydration when submitted to DSC analysis, showed a significant endothermic peak at 143°C (Fig. 3), in agreement with [28]. The pre-transition temperature (T_p) was around 100°C for pure PE liposomes.

The introduction of carboplatin into PE liposomes showed a slight shift to a lower temperature at 140°C compared to the main endothermic peak (T_m) of empty PE that exists at 143°C, indicating that carboplatin had a significant effect on PE bilayers' acyl chains, creating a conformational disorder within the phospholipid acyl chains and decreasing the transition cooperatively.

The lowered temperature of the main PE transition process suggested that carboplatin incorporation is more favored for the creation of a disordered and loose state of acyl chains. The pre-transition temperature (T_p) peak for carboplatin liposomes has been moved to higher temperatures from 100°C to 103°C, suggesting that it prefers a change from a tilted to a rippled chain gel phase.

Once β -cryptoxanthin was incorporated into PE liposomes, it resulted in distinct broadening and shift to higher temperature 160°C compared to the main characteristic endothermic peak (T_m) of pure PE that exists at 143°C, its intensity was markedly depressed.

The increased temperature of the main PE transition process indicated that the integration of β -cryptoxanthin is more proper for the creation of an ordered and cooperative state of acyl chains. The increase in PE phase transition, subsequently the reduction of the fluidity of the lipid bilayer membrane as a consequence of drug trapping. It has been found that the pre-transition temperature for β -cryptoxanthin liposomes was changed from 100°C to 127°C, which has confirmed that it prefers a transformation from a tilted to a rippled chain gel phase. Using DSC, it was observed that the PE and carboplatin or β -cryptoxanthin mixtures displayed a single peak, which indicates that they are miscible.

Such changes observed in the current DSC work can be confirmed by FTIR, which was used to

detect any vesicle structural alterations in the liposomal membrane structure by analyzing the wavenumber of different vibrational modes.

FTIR spectra of empty lyophilized PE liposomes compared with carboplatin or β -cryptoxanthin liposomal samples in the region of 4000–400 cm^{-1} as shown in (Fig. 4). Two peaks are related to the symmetric and antisymmetric stretching vibrations of the CH_2 in the acyl chain around 2850 and 2920 cm^{-1} , respectively were apparent. The peak noticed near 1470 cm^{-1} is due to the CH_2 bending vibration and that at 1734 cm^{-1} is due to the carbonyl stretching vibration $\text{C}=\text{O}$. Two peaks corresponding to the symmetric and antisymmetric PO_2^- stretching vibrations approximately at 1090 and 1220 cm^{-1} , respectively were observed. These findings were in agreement with the data reported in the literature [29].

The detailed spectral analyses are performed in three distinct wavenumber regions, namely 3500–2500 cm^{-1} (Fig. 5), 1800–1500 cm^{-1} (Fig. 6), and 1800–800 cm^{-1} (Fig. 7).

Encapsulation of carboplatin or β -cryptoxanthin into the PE liposomes caused a shift in the wavenumber of the symmetric CH_2 stretching bands in the acyl chain that appeared in Fig. 5, indicating that carboplatin or β -cryptoxanthin create a conformational disorder within the acyl chains of phospholipids.

The peak at 2844.489 cm^{-1} for the pure PE is moved towards higher wavenumber at 2847.381 cm^{-1}

for carboplatin liposomes. This could imply an increase in the number of gauche conformers that indicates an increase in bilayer disorder [30]. The signal intensity became more intensive for carboplatin loaded liposomes. Once β -cryptoxanthin is integrated into PE, the peak for pure PE at 2844.489 cm^{-1} is moved to a higher wavenumber at 2845.453 cm^{-1} , which suggests an increase in membrane fluidity and thus destabilization of the system in the gel phase (Fig. 5). The shifts to higher wavenumbers correspond to an increase in the number of gauche conformers [31]. The wavenumber of the CH_2 stretching bands is significantly changed, showing that carboplatin or β -cryptoxanthin increased the number of gauche conformers and this suggests an increase in the conformational disorder (trans-gauche isomerization) of the bilayer.

The $\text{C}=\text{O}$ stretching band was analyzed for the interaction of carboplatin or β -cryptoxanthin with the backbone of glycerol near the head group of phospholipids in the interfacial region. The wavenumber variation of this band is shown in (Fig. 6). As can be seen from (Fig. 6), for the liposomal sample containing carboplatin or β -cryptoxanthin, the wavenumber value of the $\text{C}=\text{O}$ group at 1629.555 cm^{-1} was moved to higher wavenumbers at 1630.519 cm^{-1} for the liposomal sample containing carboplatin or β -cryptoxanthin without any evidence of hydrogen bonding formation. The absorption bands of ester $\text{C}=\text{O}$ are sensitive to changes in the polarity of their local environments and are affected by hydrogen

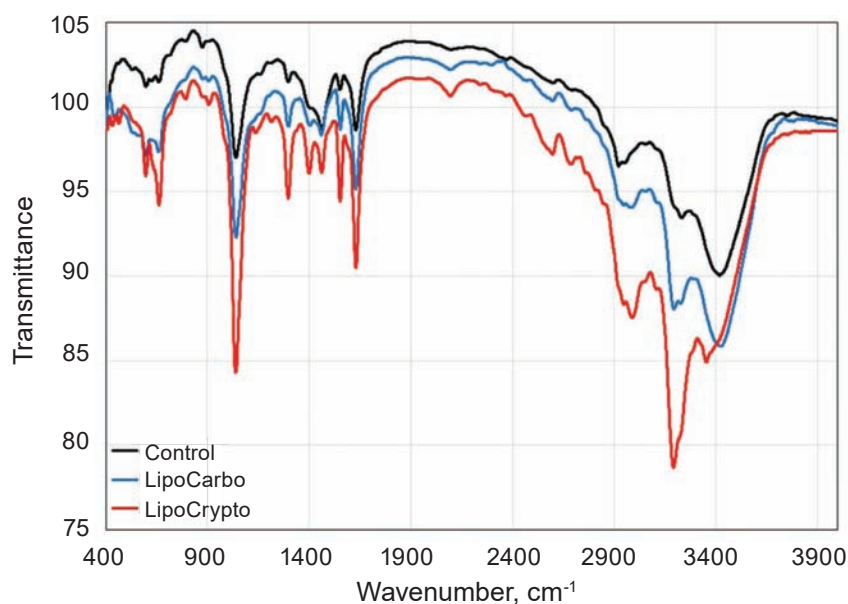


Fig. 4. The full FTIR spectra of empty PE and PE/carboplatin or β -cryptoxanthin liposomal samples

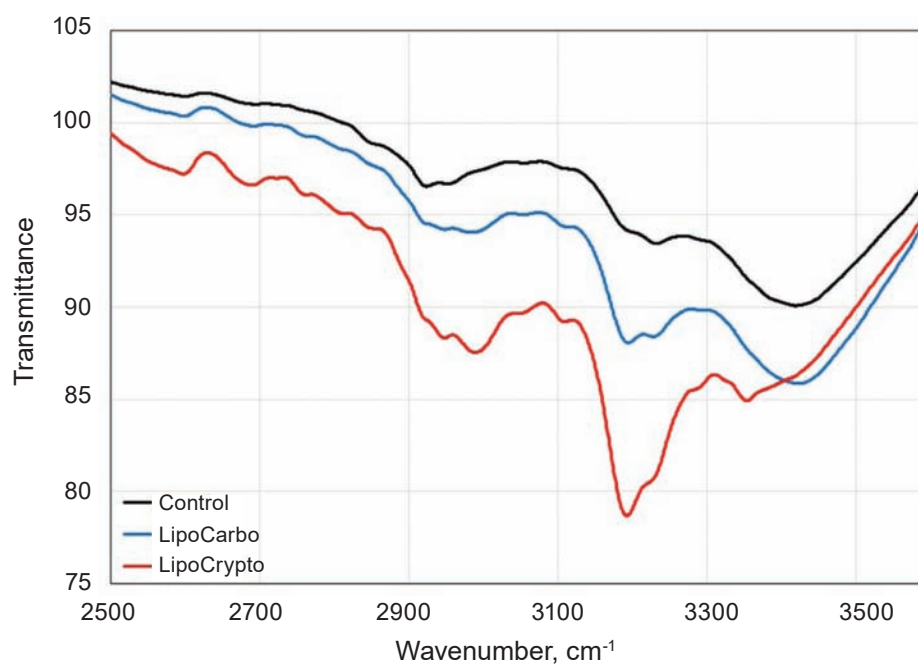


Fig. 5. The magnified part ($3500\text{--}2500\text{ cm}^{-1}$) of FTIR spectra of empty PE and PE/carboplatin or β -cryptoxanthin liposomal samples

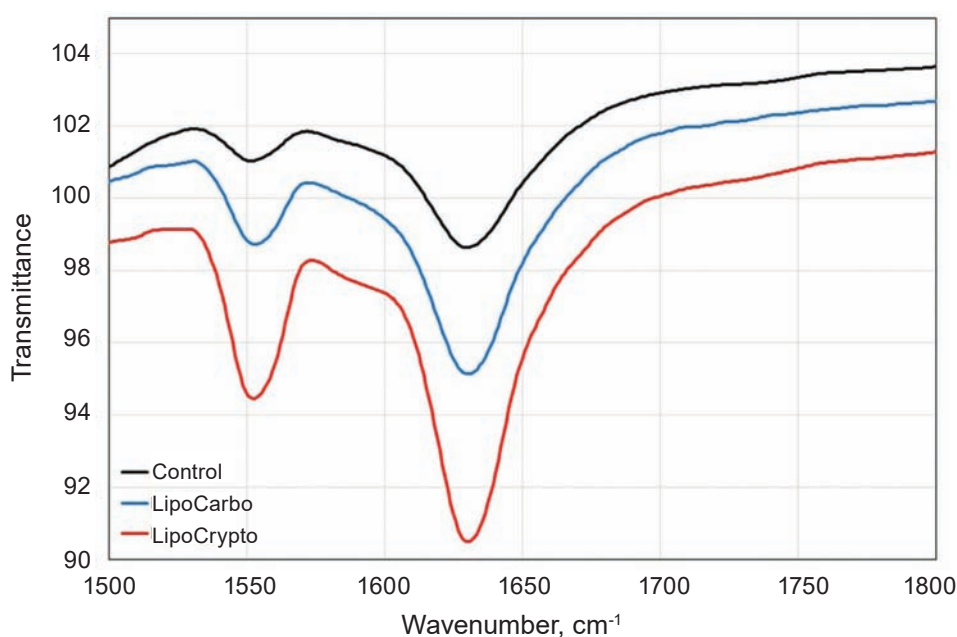


Fig. 6. The magnified part ($1800\text{--}1500\text{ cm}^{-1}$) of FTIR spectra of empty PE and PE/carboplatin or β -cryptoxanthin liposomal samples

bonding and other interactions. Any variations in the spectra in this region may, therefore, be due to an interaction between carboplatin or β -cryptoxanthin and the membrane's polar/apolar interfacial region.

Utilizing the PO_2^- antisymmetric stretching band, which is located at 1216.863 cm^{-1} , the interaction between carboplatin or β -cryptoxanthin

and the head group of PE liposomes was investigated. Fig. 7 shows the PO_2^- antisymmetric stretching band for PE liposomes formulations in the absence and presence of carboplatin or β -cryptoxanthin. As can be seen from Fig. 7, the wavenumber was shifted to lower values after the addition of carboplatin (1211.077 cm^{-1}) or β -cryptoxanthin (1213.97 cm^{-1})

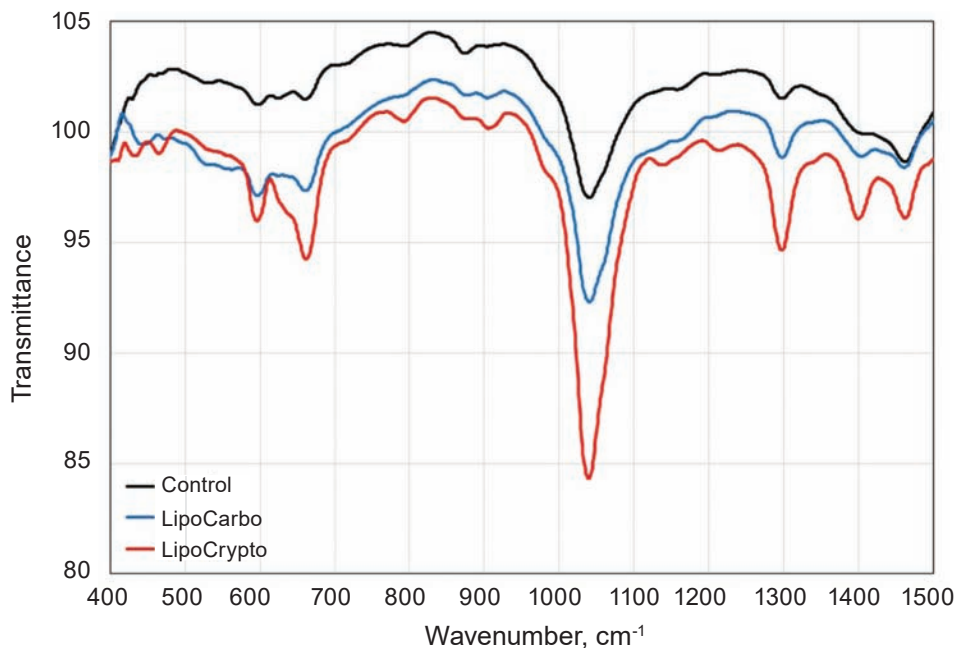


Fig. 7. The magnified part ($1800\text{--}1500\text{ cm}^{-1}$) of FTIR spectra of empty PE and PE/carboplatin or β -cryptoxanthin liposomal samples

into PE liposomes. This implied the presence of hydrogen bonding between the liposome head group and carboplatin or β -cryptoxanthin. The decrease in the wavenumber value indicates a strengthening of existing hydrogen bonds or even a formation of new hydrogen bonding between the components [30].

The CH_2 scissoring vibration mode which is located at 1461.778 cm^{-1} is influenced by the inclusion of β -cryptoxanthin into PE liposomal preparation. Following the encapsulation of β -cryptoxanthin into PE liposomes, the wavenumber was shifted towards higher values at 1462.742 cm^{-1} . This could presume that the β -cryptoxanthin molecules act as small spacers of the polar head group, resulting in a slight disorder in the hydrocarbon chains. The wavenumber was shifted to a lower value at 1460.814 cm^{-1} after the encapsulation of carboplatin into PE liposomes.

After encapsulation of carboplatin or β -cryptoxanthin into PE liposomes, the $\text{N}(\text{CH}_3)_3^+$ symmetric deformation band which is located at 1400.067 cm^{-1} was also modified (Fig. 7). This may be attributed to the presence of new intermolecular hydrogen bond formation between carboplatin or β -cryptoxanthin and $\text{N}(\text{CH}_3)_3^+$.

A shift to lower wavenumber 904.451 cm^{-1} was observed for the $\text{CN}(\text{CH}_3)_3^+$ symmetric stretching band of choline at 905.4152 cm^{-1} by the inclusion of carboplatin/PE liposomal sample. In the polar

head of PE, the existence of $\text{N}(\text{CH}_3)_3^+$ group might prevent the carboplatin NH_3^+ group from getting close to the polar head group because of the electrostatic repulsive force between the $\text{PE-N}(\text{CH}_3)_3^+$ group and the carboplatin NH_3^+ group; thus, weakening the interactions between phospholipid $\text{N}(\text{CH}_3)_3^+$ group and carboplatin NH_3^+ group. Table 2 shows the chemical shifts observed for LipoCarbo or LipoCrypto after the incorporation into L- α -Phosphatidylethanolamine liposomes.

Using cell viability (*in vitro* cytotoxicity SRB) assay, the potential cytotoxic activity of the drug delivery system has been investigated at different drug concentrations of free carboplatin or carboplatin combined with liposomes and free β -cryptoxanthin or encapsulated into liposomes against breast carcinoma (MCF-7) cell line [22]. At zero concentration of each drug untreated cells served as controls. The assay was terminated at 48 h and measurements of cell viability were performed. The MCF-7 cancer cell line was incubated separately with different concentrations of drugs in the same sequence (50, 100, 200, 400, 800, 1000, 1200, 1400, and 1500 $\mu\text{g/ml}$), for 48 h, (Fig. 8).

The highest cytotoxicity against MCF-7 cell line was observed for free β -cryptoxanthin or doped with liposomes (LipoCrypto) treated with the same series of different concentrations which applicable for the other drugs. At the highest free β -cryptoxanthin

Table 2. The chemical shifts observed for LipoCarbo or LipoCrypto after the incorporation into L- α -phosphatidylethanolamine liposomes

Peak assignment	Wavenumber, cm^{-1}	Wavenumber, cm^{-1}		
		Control Liposomes	LipoCarbo	LipoCrypto
Symmetric stretching vibration of the CH_2 in the acyl chain	(2800–2855)	2844.489	2847.381	2845.453
Carbonyl stretching vibrations (C=O)	(1724–1740)	1629.555	1630.519	1630.519
CH_2 scissoring vibration	(1457-1466)	1461.778	1460.814	1462.742
Antisymmetric PO_2 stretching vibration	(1215-1228)	1216.863	1211.077	1213.97
+N- CH_3 symmetric deformation	(1396-1405)	1400.067	1403.924	1399.1028
Symmetric stretching of Choline CN-(CH_3) ₃	(904-930)	905.4152	904.451	905.4152

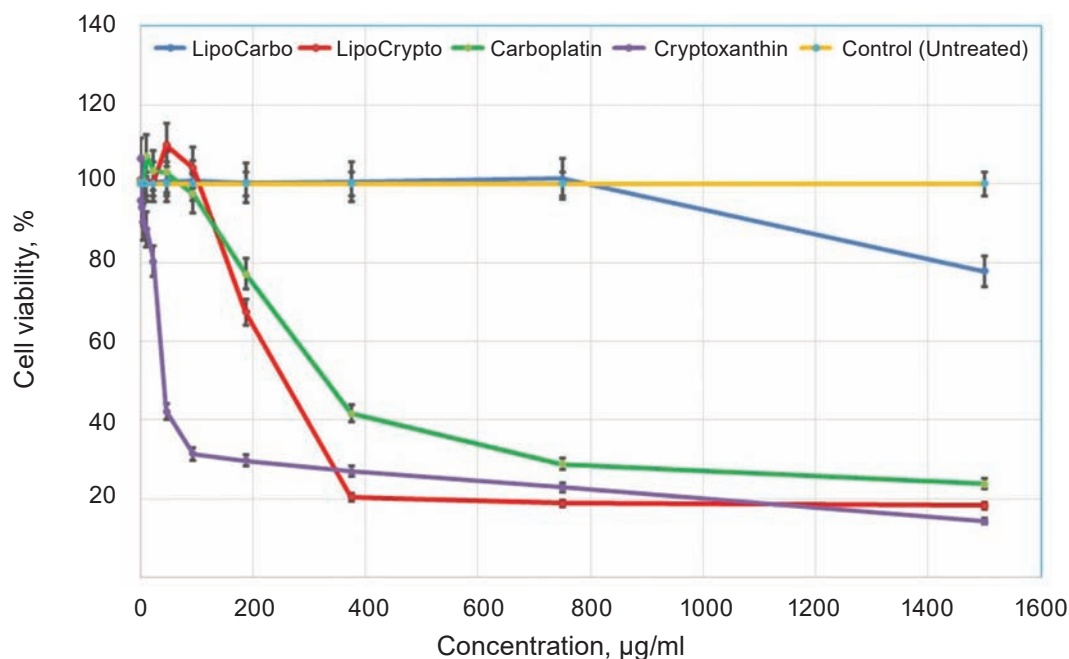


Fig. 8. Cytotoxicity of free carboplatin, free β -cryptoxanthin, and carboplatin or β -cryptoxanthin bound liposomes as opposed to untreated control breast carcinoma (MCF-7) cell line; incubated for 48 h with different concentrations (50, 100, 200, 400, 800, 1000, 1200, 1400 and 1500 $\mu\text{g}/\text{ml}$). The cell viability was determined using the SRB assay. The data represent the mean \pm SD of triplicate experiments

or integrated into liposomes concentration (1000 $\mu\text{g}/\text{ml}$), MCF-7 treated cells showed cell viability of approximately 20% ($P < 0.05$), 48 hours post-incubation. While the cell viability was approximately 30% ($P < 0.05$) for free carboplatin treated cells and approximately 90% for carboplatin-loaded liposomes at the same concentration (1000 $\mu\text{g}/\text{ml}$). For carbo-

platin loaded liposomes (LipoCarpo) treated cells, the increase in cell viability compared to the free carboplatin could be attributed to the sustained release of carboplatin from liposomes.

At higher concentrations, cytotoxic activity between different formulations of drugs shows the order of free β -cryptoxanthin or its liposomal

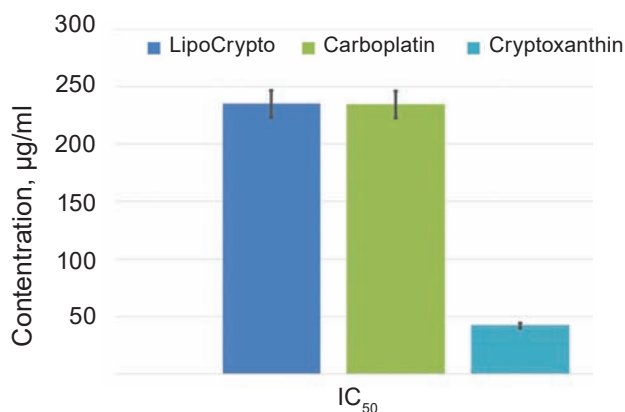


Fig. 9. IC₅₀ chart with significant values for free carboplatin, free β -cryptoxanthin or integrated into liposomes against breast carcinoma (MCF-7) cell line by using SRB assay, 48 h post-treatment

form > carboplatin > carboplatin-loaded liposomes according to Fig. 8.

At a lower concentration of approximately 42.17 $\mu\text{g/ml}$, MCF-7 treated cells with free β -cryptoxanthin showed cell viability 42% ($P < 0.05$) compared to its liposomal form of about 60 %, while 80% and 100% of the cell remained viable for carboplatin and its liposomal form (LipoCarbo), respectively Fig. 8.

In a cytotoxic assay with MCF-7 treated cells, the IC₅₀ value for free β -cryptoxanthin was 23.43 $\mu\text{g/ml}$ ($P < 0.05$), while carboplatin was counted 235 $\mu\text{g/ml}$ similarly in value with IC₅₀ of MCF-7 treated cells with β -cryptoxanthin-loaded liposomes. It can be noticed that the IC₅₀ of carboplatin-loaded liposomes (Lipocarbo) was not applicable in cytotoxic assay with MCF-7 Fig. 9. Natural medicine may be the perfect alternative for cancer treatment, eliminating the multiple physical side effects that chemotherapy can cause.

Conclusion. Our study shows that a combination of β -cryptoxanthin with liposomes shows synergistic growth inhibitory activity on breast carcinoma (MCF-7) cell line via induction of cytotoxicity. The current data suggest a new treatment regimen in which carboplatin is substituted with free β -cryptoxanthin or its liposomal form to enhance its anticancer activity against the MCF-7 cancer cell line. This finding provides a molecular basis for the development of natural compounds as novel anticancer agents and will allow lowering the dose of cytotoxic agents, which will in turn lead to more specific and less toxic therapies for mammary can-

cer in humans. Liposomal carotenoids encapsulation increases their activity and decreases the concentration needed to give their desired actions.

Conflict of interest. Authors have completed the Unified Conflicts of Interest form at http://ukr-biochemjournal.org/wp-content/uploads/2018/12/coi_disclosure.pdf and declare no conflict of interest.

ПОРІВНЯЛЬНА ЦИТОТОКСИЧНА ДІЯ КАРБОПЛАТИНУ ТА β -КРИПТОКСАНТИНУ У ВІЛЬНІЙ ТА ЛІПОСОМНІЙ ФОРМАХ НА ЛІНІЮ КЛІТИН МОЛОЧНОЇ ЗАЛОЗИ

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Дослідження ефективності антиканцерогенних сполук як синтетичного, так і природного походження у ліпосомній формі є актуальним для можливого їх застосування у терапії. У цій роботі використано алкілувальний агент карбоплатин та представник каротиноїдів β -криптоксантин. Метою дослідження було оцінити токсичну дію цих сполук у вільній та ліпосомній формах на клітини лінії MCF-7 раку молочної залози. За даними DSC та FTIR після додавання карбоплатину або β -криптоксантину до ліпосомних бішарів виявлено один пік, що вказує на їх змішування. У разі включення до ліпосом β -криптоксантину спостерігався більш упорядкований та кооперативний стан ацильних ланцюгів фосфатидилетаноламіну. Виявлено, що чутливість клітин MCF-7 до вільного β -криптоксантину була набагато вищою, ніж до вільного карбоплатину з IC₅₀ 42 та 235 мкг/мл відповідно. Значення IC₅₀ для завантаженого у ліпосоми β -криптоксантину та для вільного карбоплатину були подібними, у той самий час не було виявлено цитотоксичного ефекту ліпосом, навантажених карбоплатином. Одержані дані дозволяють запропонувати заміну карбоплатину на β -криптоксантин у вільній або ліпосомальній формі як можливу схему для підвищення ефективності терапії раку молочної залози.

Ключові слова: β -криптоксантин, карбоплатин, ліпосоми, DSC, FTIR, клітини лінії MCF-7, цитотоксичність.

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