

GAMMA RAYS INDUCED CYTOMIC TIC VARIATIONS IN POLLEN MOTHER CELLS OF CUMIN (*CUMINUM CYMINUM*) L.

G. KUMAR, M. BHARDWAJ *

Plant Genetics Laboratory, Department of Botany,
University of Allahabad, India

*E-mail-mohinibhardwaj2811@gmail.com

Cytomixis is an enigmatic cytological phenomenon found between proximately situated PMCs reported in Cuminum cyminum L. The transmigration of chromatin material between two adjacent meiocytes was observed at various exposure rate of gamma radiation viz., 100, 125, 150 and 175Gy in GC-4 variety of cumin. The microsporogenesis study of gamma treated sets reveals intercellular migration through various channels or direct fusion. In the present study, the cytomixis was more frequently noticed at meiosis I as compared to later stage i.e., meiosis II. The rate of recurrence of cytomixis shows gradual increment along with the increasing dose rate of gamma rays. As consequences of cytomixis various aberrant post meiotic products were recorded at later phase of cell cycle. Moreover, in addition to cytomixis other chromosomal abnormalities were also recorded which leads to reduction in pollen fertility. Syncytes were observed at low frequency but have high evolutionary significance as they produce gametes with higher ploidy level. The production of gametes with unbalanced chromosomes can be further exploited in breeding techniques as they are potent variation inducer.

Key words: *Cytomixis, cytoplasmic connection, Cuminum cyminum, Gamma rays, Hyperploid, Hypoploid, Syncytes.*

ГАММА-ПРОМЕНИ ІНДУКУЮТЬ
ЗМІНИ ЦИТОМІКСИСУ
В МАТЕРИНСЬКИХ КЛІТИНАХ ПИЛКУ ЗІРИ
(*CUMINUM CYMINUM*) L.

Цитоміксис – це загадкове цитологічне явище, яке спостерігається між близько розташованими материнськими клітинами пилку (МКП) *Cuminum cyminum* L. Взаємне переміщення складу хроматину між двома прилеглими мейоцитами спостерігали при різних рівнях гамма-опромінення *viz.* 100, 125, 150 і 175 Гр у сорту зіри GC-4. Дослідження мікроспорогенезу наборів, опромінених гамма-променями, виявило міжклітинну міграцію різними каналами або безпосереднє злиття. У цій роботі цитоміксис набагато частіше спостерігали як мейоз I порівняно з пізнішим етапом, тобто мейозом II. Рівень повторюваності цитоміксису демонструє поступове зростання з одночасним збільшенням дози

гамма-опромінення. Внаслідок цитоміксису було зафіксовано різні аномальні продукти постмейозу на пізнішій стадії клітинного циклу. Більше того, окрім цитоміксису, спостерігалися також інші хромосомні аномалії, які призвели до зниження родючості пилку. Синцитії спостерігали при низькій частоті, однак вони мали високу еволюційну значимість, оскільки вони виробляють гамети з високим рівнем плоідності. Утворення гамет з незбалансованих хромосом можна використовувати надалі у технологіях селекції, оскільки вони є потужним індуктором змін.

Ключові слова: цитоміксис, поєднання цитоплазми, *Cuminum cyminum*, гамма-промені, гіперплоідний, гіпоплоідний, синцитій.

REFERENCES

- Bellucci M, Roscini C, Mariani A. (2003) Cytomixis in the Pollen Mother Cells of *Medicago sativa* L J Hereditas 94:512–516. doi: <https://doi.org/10.1093/jhered/esg096>
- Bhat TA, Parveen S, Khan AH. (2006) MMS-induced cytomixis in pollen mother cells of broad bean (*Vicia faba* L.) Turk J Bot 30:273–279.
- Bione NCP, Pagliarini MS, de Tolero JFF. (2000) Meiotic Behavior of Several Brazilian Soybean Varieties Genet Mol Biol 23:623–631. doi: <https://doi.org/10.1590/S1415-4757200000300022>.
- Boldrini KR, Pagliarini MS, Valle CB. (2006) Cell fusion and cytomixis during microsporogenesis in *Brachiaria humidicola* (Poaceae) S Afr 72:478–481. doi: <https://doi.org/10.1016/j.sajb.2005.11.004>
- Datta AK, Mukherjee M, Iqbal M. (2005) Persistent cytomixis in *Ocimum basilicum* L. (Lamiaceae) and *Withania somnifera* (L.) Dun (Solanaceae) Cytologia 70:309–313. doi: <https://doi.org/10.1508/cytologia.70.309>
- Dwivedi H, Kumar G. (2018) Induced syncyte formation via cytomixis in *Trachyspermum ammi* (L.) Sprague (Apiaceae) Caryologia 71(4):420–427. doi: <https://doi.org/10.108/00087114.2018.1499480>
- Ghaffari SM. (2006) Occurrence of diploid and polyploidy microspores in *Sorghum bicolor* (Poaceae) is the result of cytomixis Afr J Biotechnol 5:1450–1453
- Guan JZ, Wang JJ, Cheng ZH, Li ZY. (2012) Cytomixis and meiotic abnormalities during microsporogenesis are responsible for male sterility and chromosome variations in *Houttuynia cordata* Genet Mol Res 11:121–130. doi: <http://dx.doi.org/10.4238/2012pp>.
- Guo GQ, Zheng GC. (2004) Hypothesis for Functions of Intercellular Bridges in Male Germ Cell Development and Its Cellular Mechanisms J Theor Biol 139–146. doi: <https://doi.org/10.1016/j.jtbi.2004.03.010>
- Haroun SA, Al Shehr AM, Al Wadie HM. (2004)

- Cytomixis in the microsporogenesis of *Vicia faba* L. (Fabaceae) *Cytologia* 69:7–11. doi: <https://doi.org/10.1508/cytologia.69.7>
- Kornicke M. (1901) Über Ortsveränderung von Zellkarnern SB, Niederhein Ges Natur-U Heilkunde 14–25p
- Kravets EA. (2009) Cellular and tissue mechanisms of recovery processes in *Hordeum distichum* L. under irradiation *Cytol Genet* 43:9–17. doi: <https://doi.org/10.3103/S0095452709010022>
- Kravets EA. (2012) Nature, significance, and cytological consequences of cytomixis *Cytol Genet* 46:188–195. doi: <https://doi.org/10.3103/S0095452712030061>
- Kravets EA. (2018) Cytomixis as a primary form of sexual process *Adv Cytol Pathol* 3:88–91. doi: [10.15406/acp.2018.03.00059](https://doi.org/10.15406/acp.2018.03.00059)
- Kumar G, Singh S. (2020) Induced cytotoxic cross-talk behaviour among micro-meioocytes of *Cyamopsis tetragonoloba* (L.) Taub. (cluster bean): Reasons and repercussions *Caryologia Inter J Cytol Cytosystem Cytogen* 73(2):111–119. doi: <https://doi.org/10.13128/caryologia-544>
- Kumar P, Singhal VK. (2008) Cytology of *Caltha palustris* L. (Ranunculaceae) from Cold Regions of Western Himalayas *Cytologia* 73:137–143. doi: <https://doi.org/10.1508/cytologia.73.137>
- Kumar P, Singhal VK. (2011) Male meiosis, morphometric analysis and distribution pattern of 2x and 4x cytotypes of *Ranunculus hirtellus* Royle, 1834 (Ranunculaceae) from the cold regions of northwest Himalayas (India) *Comp Cytogenet* 5:143–161. doi: <https://doi.org/10.3897/CompCytogen.v5i3.1359>
- Kunakh VA. (2011) Ontogeneticheskaya plastichnost' genoma kak osnova adaptivnosti rastenii. Zhebrakovskie Chteniya. III, Minsk: Pravo i ekonomika,.
- Larrosa FH, Maune JF, Erazzu LE, Camadro EL. (2012) Meiotic abnormalities underlying pollen sterility in wild potato hybrids and spontaneous populations *Pl Biol* 14:223–233. doi: <https://doi.org/10.1111/j.1438-8677.2011.00470.x>
- Lattoo SK, Khan S, Bamotra S, Dhar AK. (2006) Cytomixis Impairs Meiosis and Influences Reproductive Success in *Chlorophytum comosum* (Thunb.) Jacq. An Additional Strategy and Possible Implications *J Biosci* 31:629–637. doi: <https://doi.org/10.1007/BF02708415>
- Liu H, Guo GQ, He YK, Lu YP, Zheng GC. (2007) Visualization on Intercellular Movement of Chromatin in Intact Living Anthers of Transgenic Tobacco Expressing Histone 2BCFP Fusion Protein *Caryologia* 60:1–20. doi: doi.org/10.1080/00087114.2007.10589542
- Lone FA, Wafai BA. (2009) Chromosome conspectus and cytogenetic appraisal of some commercial cultivars of cherry (*Prunus avium* L., *P. cerasus* L.) and plum (*P. domestica* L.) using a partially modified staining technique *Phytomorphology* 59:29–34.
- Malallah GA, Talaat AA. (2003) Cytomixis and its possible evolutionary role in a Kuwaiti population of *Diplotaxis harra* (Brassicaceae) *Bot J Linn Soc* 143:169–175. doi: <https://doi.org/10.1046/j.1095-8339.2003.00218.x>
- Malallah GA. (2011) Cytomixis and Its Possible Evolutionary Role in a Kuwaiti Population of *Diplotaxis harra* (Brassicaceae) *Comp Cytogenet* 5:143–161.
- Mamun EA, Alfred S, Cantrill LC, Overall RL, Sutton BG. (2006) Effects of chilling on male gametophyte development in rice *Cell Biol Int* 30:583–591. doi: <https://doi.org/10.1016/j.cellbi.2006.03.004>
- Mursalimov SR, Deineko EV. (2012) An ultrastructural study of microsporogenesis in tobacco line SR1 *Biologia* 67:369–376. doi: <https://doi.org/10.2478/s11756-012-0005-1>
- Mursalimov S, Sidorchuk Y, Deineko E. (2013) The role of spherosome-like vesicles in formation of cytotoxic channels between tobacco microsporocytes *Biol Plantarum* 57:291–297. doi: <https://doi.org/10.1007/s10535-012-0276-y>
- Mursalimov SR, Deineko EV. (2015) How cytomixis can form unreduced gametes in tobacco *Plant Syst Evol* 301:1293–1297. doi: <https://doi.org/10.1007/s00606-014-1150-5>
- Nirmala C, Kaul MLH. (1994) Male Sterility in Pea 6 *Gene Action Duplicity Cytologia* 59:195–201.
- Pagliarini MS. (2000) Meiotic behaviour of economically important plant species: the relationship between fertility and male sterility *Genet Mol Biol* 23:997–1002. doi: <https://doi.org/10.1590/S1415-47572000000400045>
- Papihin RV, Yandovka LF. (2014) Cytomixis During Microsporogenesis in *Cerasus fruticosa* Pall *Biomed Pharmacol J* 7:473–479. doi: <http://dx.doi.org/10.13005/bpj/514>
- Peng ZHS, Yang J, Zheng GCH. (2003) Cytomixis in pollen mother cells of new synthetic hexaploid amphidiploid (*Aegilops tauschii*, *Triticum turgidum*) *Cytologia* 68:335–340. doi: <https://doi.org/10.1508/cytologia.68.335>
- Popova AF, Ivanenko GF, Ustinova AYU, Zaslavsky VA. (2008) Localization of callose in microspores and pollen grains in *Stium latifolium* L. plants in different water regimes *Cytol Genet* 42:363–368. doi: <https://doi.org/10.3103/S0095452708060017>
- Ramanna MS, Jacobsen E. (2003) Relevance of sexual polyploidization for crop improvement – a review *Euphytica* 133:3–18. doi: <https://doi.org/10.1023/A:1025600824483>
- Ramsey J, Schemske DW. (2002) Neopolyploidy in flo-

- wering plants *Ann Rev Ecol Syst* 33:589–639. doi: <https://doi.org/10.1146/annurev.ecolsys.33.010802.150437>
- Ranjbar M, Karamian R, Nouri S. (2011) Impact of cytotoxicity on meiosis in *Astragalus cyclophyllus* Beck (Fabaceae) from Iran *Caryologia* 64:256–264. doi: <https://doi.org/10.1080/00087114.2011.10589791>
- Reis AC, Sousa SM, Vale AA, Pierre PMO, Franco AL, Campos JMS, Viera RF, Viccini LF. (2014) *Lippia alba* (Verbenaceae): a new tropical autopolyploid complex? *Amer J Bot* 101:1002–1012. doi: <https://doi.org/10.3732/ajb.1400149>
- Saraswathy Amma CK, Namboodiri AN, Panikkar AON, Sethuraj MR. (1990) Radiation induced male sterility in *Hevea brasiliensis* (Willd. ex Adr. De Juss.) Muell Arg *Cytologia* 55:547–551. doi: <https://doi.org/10.1508/cytologia.55.547>
- Shkutina FM, Kozlovskaya VF. (1974) Cytotoxicity in Meiosis of Some Hybrid Forms of Cereals of the Triticeae Subtribe *Genetics* 10:5–12.
- Sidorchuk YuV, Deineko EV, Shumnyi VK. (2007) Characteristics of the cytotoxicity in the pollen mother cells of transgenic tobacco plants (*Nicotiana tabacum* L.) with mutant phenotype *Tsitologiya* 49:870–875
- Singhal VK, Gill BS, Dhaliwal RS. (2007) Status of chromosomal diversity in the hardwood tree species of Punjab State *Cytol Genet* 8:67–83.
- Singhal VK, Kumar P. (2008) Impact of Cytotoxicity on Meiosis, Pollen Viability and Pollen Size in Wild Populations of Himalayan Poppy (*Meconopsis aculeate* Royle) *J Biosci* 33:371–380.
- Singhal VK, Kaur S, Kumar P. (2010) Aberrant Male Meiosis, Pollen Sterility and Variable Sized Pollen Grains in *Clematis Montana* Buch. Ham. ex DC. From Dalhousie Hills, Himachal Pradesh *Cytologia* 75:31–36. doi: <https://doi.org/10.1508/cytologia.75.31>
- Singhal VK, Gill BS, Dhaliwal RS. (2007) Status of chromosomal diversity in the hardwood tree species of Punjab State *Cytol Genet* 8:67–83.
- Soltis DE, Pires JC, Kovarik A, Tate J, Mavrodiev E. (2004) Recent and recurrent polyploidy in *Tragopogon* (Asteraceae): cytogenetic, genomic and genetic comparisons *Biol J Linn Soc* 82:485–501. doi: <https://doi.org/10.1111/j.1095-8312.2004.00335.x>
- Wang SYu, Yu ChH, Li S, Wang ChY, Zheng GC. (2004) Ultrastructural Aspects and Possible Origin of Cytoplasmic Channels Providing Intercellular Connection in Vegetative Tissues of Anthers *Russ J Plant Physiol* 51:97–106. doi: <https://doi.org/10.1023/B:RUPP.0000011308.61339.fe>
- Yu CH, Guo GQ, Nie XW, Zheng GC. (2004) Cytochemical localization of pectinase activity in pollen mother cells of tobacco during meiotic prophase and its relation to the formation of secondary plasmodesmata and cytoplasmic channels, *Acta Bot Sin* 46:143–145
- Zhou SQ. (2003) Viewing the difference between the diploid and the polyploid in the light of the upland cotton aneuploid *Hereditas* 138:65–72. doi: <https://doi.org/10.1034/j.1601-5223.2003.01689.x>

Received October 13, 2020

Received March 02, 2021

Accepted May 18, 2022