

## TAP2 EFFECT ON MIN-PIG STROMAL VASCULAR FRACTION CELL GENE EXPRESSION

L. WANG, D. LIU\*, H. MA, D. ZHANG,  
X. HE, W. WANG, B. FU, Z. LI, Z GUO\*

Postdoctoral Workstation of Heilongjiang Academy of Agricultural Sciences, Institute of Animal Husbandry, Key Laboratory of Combining Farming and Animal Husbandry, Ministry of Agriculture and Rural Affairs, No. 368 Xuefu Road, Harbin 150086, P. R. China  
E-mail: liudi1963@163.com, gzhh00@163.com

*We have hypothesized that the TAP2 gene is associated with lipid metabolism. Here, 10 Min-pig tissues were collected to detect the expression of TAP2 in different tissues. We obtained dorsal subcutaneous structural vascular fraction (SVF) cells from the Min-pig's back adipose tissue and induced SVF cells into mature adipocytes. By overexpression and interference, the effect of TAP2 on fat deposition in Min-pig SVF cells was studied. Recombinant human insulin, dexamethasone, indomethacin, 3-isobutyl-1-methylxanthine, triiodothyronine, and rosiglitazone could successfully induce SVF cells into mature adipocytes, and the induction efficiency was above 50 %. The tissue expression profile showed that TAP2 was expressed in different tissues, and the highest expression was found in back fat, spleen, and back muscle. Overexpression of the TAP2 gene in SVF cells could significantly increase the expression of adipose differentiation-related genes. The expression of TAP2 in SVF cells was reduced to 0.6 times after transfection of the TAP2 gene interference fragment. The adipogenic transcription factor gene C/EBP $\alpha$ , fatty acid synthase gene FSA, and adipocyte directional differentiation factor ADD1 were downregulated, while the expression of lipolysis-related gene LPL was inhibited. In conclusion, TAP2 expression can promote the deposition of subcutaneous fat on Min-pig's back adipose tissue.*

**Key words:** gene interference, gene overexpression, Min-pig, SVF cells, TAP2.

### ВПЛИВ TAP2 НА ЕКСПРЕСІЮ ГЕНІВ КЛІТИН СТРОМАЛЬНОЇ СУДИННОЇ ФРАКЦІЇ У СВИНЕЙ ПОРОДИ МІН

Ми припустили, що ген TAP2 пов'язаний з метаболізмом ліпідів. Для виявлення експресії TAP2 різних тканин використали 10 зразків тканин, отриманих від свиней породи Мін. Ми отримали клітини дорсальної підшкірної стромальної судинної фракції (ССФ) із хребтових жирових тканин сви-

ней породи Мін та індукували клітини ССФ у зрілі жирові клітини. Вплив TAP2 на відкладення жиру в ССФ клітинах свиней породи Мін вивчали за надмірною експресією та інтерференцією. Рекombінантний інсулін людини, дексаметазон, індометацин, 3-ізобутил-1-метилксантин, трийодтиронін і розиглітазон були здатні успішно індукувати ССФ клітини в зрілі жирові клітини; ефективність індукції становила більше 50 %. Профіль експресії тканин продемонстрував, що експресія TAP2 проходила в різних тканинах, найвищий рівень експресії було зафіксовано в хребтовому жиру, селезінці та спинних м'язах. Надмірна експресія гену TAP2 у ССФ клітинах може значно підвищити рівень експресії генів жирових клітин, пов'язаних із диференціацією. Експресія TAP2 у ССФ клітинах знизилась у 0,6 рази після трансфекції фрагменту інтерференції гену TAP2. Було відмічено негативне регулювання адипогенного гена фактора транскрипції C/EBP $\alpha$ , гена синтази жирних кислот FSA та адипоцитного фактора диференціації спрямованої дії ADD1, а також інгібування експресії гена LPL, пов'язаного з ліполізом. Отже, експресія TAP2 може сприяти відкладенню підшкірного жиру в хребтових жирових тканинах свиней породи Мін.

**Ключові слова:** інтерференція гена, надмірна експресія гена, свині породи Мін, клітини ССФ, TAP2.

### REFERENCES

- Al-Ghadban S, Diaz ZT, Singer HJ et al (2020) Increase in Leptin and PPAR-gamma Gene Expression in Lipedema Adipocytes Differentiated *in vitro* from Adipose-Derived Stem Cells Cells 9(2). <https://doi.org/10.3390/cells9020430>
- Albuquerque A, Ovilo C, Nunez Y et al (2021) Transcriptomic Profiling of Skeletal Muscle Reveals Candidate Genes Influencing Muscle Growth and Associated Lipid Composition in Portuguese Local Pig Breeds Animals (Basel) 11(5). <https://doi.org/10.3390/ani11051423>
- Alharthi AM, Banaganapalli B, Hassan SM et al (2022) Complex Inheritance of Rare Missense Variants in PAK2, TAP2, and PLCL1 Genes in a Consanguineous Arab Family With Multiple Autoimmune Diseases Including Celiac Disease. Front Pediatr 10:895298. <https://doi.org/10.3389/fped.2022.895298>
- Barr VA, Balagopalan L, Barda-Saad M et al (2006) T-cell antigen receptor-induced signaling complexes: internalization via a cholesterol-dependent endocytic pathway. Traffic 7(9):1143–1162. <https://doi.org/10.1111/j.1600-0854.2006.00464.x>
- Belleville-Rolland T, Leuci A, Mansour A et al (2021) Role of membrane lipid rafts in MRP4 (ABCC4)-dependent regulation of the cAMP pathway in blo-

- od platelets. *Thromb Haemost.* <https://doi.org/10.1055/a-1481-2663>
- Catani MV, Tullio V, Maccarrone M et al (2023) DNA-Protein-Interaction (DPI)-ELISA Assay for PPAR-gamma Receptor Binding. *Methods Mol Biol* 2576:133–143. [https://doi.org/10.1007/978-1-0716-2728-0\\_10](https://doi.org/10.1007/978-1-0716-2728-0_10)
- Chen J, Zhang H, Gao H et al (2019) Effects of Dietary Supplementation of Alpha-Ketoglutarate in a Low-Protein Diet on Fatty Acid Composition and Lipid Metabolism Related Gene Expression in Muscles of Growing Pigs. *Animals (Basel)* 9(10). <https://doi.org/10.3390/ani9100838>
- Chen Z, Torrens JI, Anand A et al (2005) Krox20 stimulates adipogenesis via C/EBPbeta-dependent and -independent mechanisms. *Cell Metab* 1(2):93–106. <https://doi.org/10.1016/j.cmet.2004.12.009>
- Cimmino I, Oriente F, D'Esposito V et al (2019) Low-dose Bisphenol-A regulates inflammatory cytokines through GPR30 in mammary adipose cells. *J Mol Endocrinol* 63(4):273–283. <https://doi.org/10.1530/JME-18-0265>
- Darazam IA, Hakamifard A, Momenilandi M et al (2023) Delayed Diagnosis of Chronic Necrotizing Granulomatous Skin Lesions due to TAP2 Deficiency. *J Clin Immunol* 43(1):217–228. <https://doi.org/10.1007/s10875-022-01374-7>
- El-Assaad A, Dawy Z, and Nemer G (2015) Electrostatic study of Alanine mutational effects on transcription: application to GATA-3:DNA interaction complex. *Annu Int Conf IEEE Eng Med Biol Soc* 2015:4005–4008. <https://doi.org/10.1109/EMBC.2015.7319272>
- Farmer SR (2006) Transcriptional control of adipocyte formation. *Cell Metab* 4(4):263–273. <https://doi.org/10.1016/j.cmet.2006.07.001>
- Feng Y, An Z, Chen H et al (2020) Ulva prolifera Extract Alleviates Intestinal Oxidative Stress via Nrf2 Signaling in Weaned Piglets Challenged With Hydrogen Peroxide. *Front Immunol* 11:599735. <https://doi.org/10.3389/fimmu.2020.599735>
- Flori L, Rogel-Gaillard C, Cochet M et al (2008) Transcriptomic analysis of the dialogue between Pseudorabies virus and porcine epithelial cells during infection. *BMC Genomics* 9:123. <https://doi.org/10.1186/1471-2164-9-123>
- Gajate C, and Mollinedo F (2021) Lipid Raft Isolation by Sucrose Gradient Centrifugation and Visualization of Raft-Located Proteins by Fluorescence Microscopy: The Use of Combined Techniques to Assess Fas/CD95 Location in Rafts During Apoptosis Triggering. *Methods Mol Biol* 2187:147–186. [https://doi.org/10.1007/978-1-0716-0814-2\\_9](https://doi.org/10.1007/978-1-0716-0814-2_9)
- Garcia-Borges CN, Phanavanh B, and Crew MD (2006) Characterization of porcine TAP genes: alternative splicing of TAP1. *Immunogenetics* 58(5–6):374–382. <https://doi.org/10.1007/s00251-006-0103-8>
- Han SL, Liu Y, Limbu SM et al (2021) The reduction of lipid-sourced energy production caused by ATGL inhibition cannot be compensated by activation of HSL, autophagy, and utilization of other nutrients in fish. *Fish Physiol Biochem* 47(1):173–188. <https://doi.org/10.1007/s10695-020-00904-7>
- Huang W, Zhang X, Li A et al (2018) Genome-Wide Analysis of mRNAs and lncRNAs of Intramuscular Fat Related to Lipid Metabolism in Two Pig Breeds. *Cell Physiol Biochem* 50(6):2406–2422. <https://doi.org/10.1159/000495101>
- Jiang MJ, Li L, Huang WF et al (2021) Rubus chingii var. suavissimus alleviates high-fat diet-induced lipid metabolism disorder via modulation of the PPARs/SREBP pathway in Syrian golden hamsters. *J Nat Med.* <https://doi.org/10.1007/s11418-021-01536-8>
- Lee SH, Lee SH, Park HB et al (2023) Identification of key adipogenic transcription factors for the pork belly parameters via the association weight matrix. *Meat Sci* 195:109015. <https://doi.org/10.1016/j.meatsci.2022.109015>
- Li D, Zhang F, Zhang X et al (2016) Distinct functions of PPARgamma isoforms in regulating adipocyte plasticity. *Biochem Biophys Res Commun* 481(1–2):132–138. <https://doi.org/10.1016/j.bbrc.2016.10.152>
- Lin J, Cao C, Tao C et al (2017) Cold adaptation in pigs depends on UCP3 in beige adipocytes. *J Mol Cell Biol* 9(5):364–375. <https://doi.org/10.1093/jmcb/mjx018>
- Liu L, Cao P, Zhang L et al (2021) Comparisons of adipogenesis- and lipid metabolism-related gene expression levels in muscle, adipose tissue and liver from Wagyu-cross and Holstein steers. *PLoS One* 16(2):e0247559. <https://doi.org/10.1371/journal.pone.0247559>
- Liu Y, Yang X, Jing X et al (2017) Transcriptomics Analysis on Excellent Meat Quality Traits of Skeletal Muscles of the Chinese Indigenous Min Pig Compared with the Large White Breed. *Int J Mol Sci* 19(1). <https://doi.org/10.3390/ijms19010021>
- Ma W, Lehner PJ, Cresswell P et al (1997) Interferon-gamma rapidly increases peptide transporter (TAP) subunit expression and peptide transport capacity in endothelial cells. *J Biol Chem* 272(26):16585–16590. <https://doi.org/10.1074/jbc.272.26.16585>
- Nagy N, Vanky F, and Klein E (1998) Tumor surveillance: expression of the transporter associated with antigen processing (TAP-1) in *ex vivo* human tumor samples and its elevation by *in vitro* treatment with IFN-gamma and TNF-alpha. *Immunol Lett* 64(2–3):153–160. [https://doi.org/10.1016/s0165-2478\(98\)00104-7](https://doi.org/10.1016/s0165-2478(98)00104-7)

- Nian HY, Zhang RX, Ding SS et al (2023) Emotional responses of piglets under long-term exposure to negative and positive auditory stimuli. *Domest Anim Endocrinol* 82:106771. <https://doi.org/10.1016/j.domaniend.2022.106771>
- Pan Y, Jing J, Qiao L et al (2018) MiRNA-seq reveals that miR-124-3p inhibits adipogenic differentiation of the stromal vascular fraction in sheep via targeting C/EBPalpha. *Domest Anim Endocrinol* 65:17–23. <https://doi.org/10.1016/j.domaniend.2018.05.002>
- Pang T, Tang Q, Wei J et al (2022) Construction of a Novel Immune-Related prognostic-predicting model of gastric cancer. *Gene* 147032. <https://doi.org/10.1016/j.gene.2022.147032>
- Park WY, Park J, Lee S et al (2022) PEX13 is required for thermogenesis of white adipose tissue in cold-exposed mice. *Biochim Biophys Acta Mol Cell Biol Lipids* 1867(1):159046. <https://doi.org/10.1016/j.bbailip.2021.159046>
- Raghavan M, Del Cid N, Rizvi SM et al (2008) MHC class I assembly: out and about. *Trends Immunol* 29(9):436–443. <https://doi.org/10.1016/j.it.2008.06.004>
- Rosen ED, Hsu CH, Wang X et al (2002) C/EBPalpha induces adipogenesis through PPARgamma: a unified pathway. *Genes Dev* 16(1):22–26. <https://doi.org/10.1101/gad.948702>
- Roth D, Benz J, Grether U et al (2023) Homogeneous Time-Resolved Fluorescence Resonance Energy Transfer (TR-FRET) Cofactor Recruitment Assay for PPARalpha and PPARgamma. *Methods Mol Biol* 2576:155–169. [https://doi.org/10.1007/978-1-0716-2728-0\\_12](https://doi.org/10.1007/978-1-0716-2728-0_12)
- Salazar-Onfray F, Charo J, Petersson M et al (1997) Down-regulation of the expression and function of the transporter associated with antigen processing in murine tumor cell lines expressing IL-10. *J Immunol* 159(7):3195–202
- Sidhom EH, Kim C, Kost-Alimova M et al (2021) Targeting a Braf/Mapk pathway rescues podocyte lipid peroxidation in CoQ-deficiency kidney disease. *J Clin Invest* 131(5). <https://doi.org/10.1172/JCI141380>
- Sun N, Liu D, Chen H et al (2012) Localization, expression change in PRRSV infection and association analysis of the porcine TAP1 gene. *Int J Biol Sci* 8(1):49–58. <https://doi.org/10.7150/ijbs.8.49>
- Suzuki K, Yanagi M, Mori-Aoki A et al (2002) Transfection of single-stranded hepatitis A virus RNA activates MHC class I pathway. *Clin Exp Immunol* 127(2):234–242. <https://doi.org/10.1046/j.1365-2249.2002.01767.x>
- Tanaka T, Yoshida N, Kishimoto T et al (1997) Defective adipocyte differentiation in mice lacking the C/EBPbeta and/or C/EBPdelta gene. *EMBO J* 16(24):7432–7443. <https://doi.org/10.1093/emboj/16.24.7432>
- Tang S, Xie J, Wu W et al (2020) High ammonia exposure regulates lipid metabolism in the pig skeletal muscle via mTOR pathway. *Sci Total Environ* 740:139917. <https://doi.org/10.1016/j.scitotenv.2020.139917>
- Vaske DA, Ruohonen-Lehto MK, Larson RG et al (1994) Rapid communication: restriction fragment length polymorphisms at the porcine transporter associated with antigen processing 1 (TAP1) locus. *J Anim Sci* 72(1):255. <https://doi.org/10.2527/1994.721255x>
- Wang J, Hua L, Chen J et al (2017) Identification and characterization of long non-coding RNAs in subcutaneous adipose tissue from castrated and intact full-sib pair Huainan male pigs. *BMC Genomics* 18(1):542. <https://doi.org/10.1186/s12864-017-3907-z>
- Wang Q, Song Z, Yang J et al (2022) Transcriptomic analysis of the innate immune signatures of a SARS-CoV-2 protein subunit vaccine ZF2001 and an mRNA vaccine RRV. *Emerg Microbes Infect* 11(1):1145–1153. <https://doi.org/10.1080/22221751.2022.2059404>
- Xue P, Hou Y, Zuo Z et al (2020) Long isoforms of NRF1 negatively regulate adipogenesis via suppression of PPARgamma expression. *Redox Biol* 30:101414. <https://doi.org/10.1016/j.redox.2019.101414>
- Zhang M, Shao Y, Gao B et al (2020) Erchen Decoction Mitigates Lipid Metabolism Disorder by the Regulation of PPARgamma and LPL Gene in a High-Fat Diet C57BL/6 Mice Model. *Evid Based Complement Alternat Med* 2020:9102475. <https://doi.org/10.1155/2020/9102475>
- Zhang Y, Wu H, He R et al (2021) Dickkopf-2 knockdown protects against classic macrophage polarization and lipid loading by activation of Wnt/beta-catenin signaling. *J Cardiol*. <https://doi.org/10.1016/j.jicc.2021.04.010>
- Zhu K, Wang J, Zhu J et al (1999) p53 induces TAP1 and enhances the transport of MHC class I peptides. *Oncogene* 18(54):7740–7747. <https://doi.org/10.1038/sj.onc.1203235>
- Zou C, Li S, Deng L et al (2017) Transcriptome Analysis Reveals Long Intergenic Noncoding RNAs Contributed to Growth and Meat Quality Differences between Yorkshire and Wannanhua Pig. *Genes (Basel)* 8(8). <https://doi.org/10.3390/genes8080203>
- Zou C, Li L, Cheng X et al (2018) Identification and Functional Analysis of Long Intergenic Non-coding RNAs Underlying Intramuscular Fat Content in Pigs. *Front Genet* 9:102. <https://doi.org/10.3389/fgene.2018.00102>
- Zubiria MG, Giordano AP, Gambaro SE et al (2020) Dexamethasone primes adipocyte precursor cells for differentiation by enhancing adipogenic competency. *Life Sci* 261:118363. <https://doi.org/10.1016/j.lfs.2020.118363>

Received November 30, 2022

Received February 03, 2023

Accepted January 18, 2024