

■ РЕФЕРАТИ СТАТЕЙ, ОПУБЛІКОВАНИХ В «CYTOLOGY AND GENETICS», № 1, 2025 р.

CO-OVEREXPRESSION OF GENES *RFE1*, *GND1*, AND *RIB6* ENHANCES RIBOFLAVIN PRODUCTION IN YEAST *CANDIDA FAMATA*

W. LIU ^{1,2}, A. TSYRULNYK ¹, K. DMYTRUK ¹,
D. FEDOROVYCH ¹, Y. KANG ², A. SIBIRNY ^{1,3,*}

¹ Department of Molecular Genetics and Biotechnology, Institute of Cell Biology, NAS of Ukraine, Drahomanov St., 14/16, Lviv 79005, Ukraine

² Key Laboratory of Environmental Pollution Monitoring and Disease Control, Ministry of Education of Guizhou & School of Basic Medical Science & Institution of One Health Research, Guizhou Medical University, Guiyang, China.

³ Institute of Biology and Biotechnology, University of Rzeszow, Zelwerowicza 4, Rzeszow 35-601, Poland

* Correspondence: Sibirny A., e-mail: sibirny@cellbiol.lviv.ua

Riboflavin (vitamin B₂) is an essential compound for the nutrition of living organisms, serving as a precursor of coenzymes flavin mononucleotide and flavin adenine dinucleotide, which are involved in numerous enzymatic reactions mainly in oxidative metabolism. The yeast Candida famata is a natural riboflavin-producing species, which are able to oversynthesis riboflavin under conditions of iron starvation. The aim of this study was to construct recombinant C. famata strains with increased riboflavin production by the simultaneous overexpression of the three genes: RFE1, GND1, and RIB6, which encode riboflavin excretase, 6-phosphogluconate dehydrogenase and 3,4-dihydroxy-2-butanone-4-phosphate synthase, respectively. The expression of various combinations of two genes, as well as the co-expression of all three genes, resulted in increased riboflavin production in C. famata VKM Y-9 in different media. The strains engineered for the co-overexpression of all three genes exhibited up to a 3.3-fold increase in riboflavin production by the fifth day of cultivation in cheese whey, compared to the parental strain.

Key words: *Candida famata*, riboflavin, vitamin B₂, cheese whey, yeast.

КОЕКСПРЕСІЯ ГЕНІВ *RFE1*, *GND1* ТА *RIB6*
ПІДВИЩУЄ ПРОДУКЦІЮ РИБОФЛАВІНУ
У ДРІЖДЖІВ *CANDIDA FAMATA*

Рибофлавін (вітамін В₂) є невід'ємним компонентом живих організмів, слугуючи попередником коферментів флавінмононуклеотиду та флавінаденіндинуклеотиду, що беруть участь у численних ферментативних реакціях, переважно в окисному метаболізмі. Дріжджі *Candida famata* здатні до надсинтезу рибофлавіну за умов дефіциту заліза. Метою даної роботи було конструювання рекомбінантних штамів *C. famata* з підвищеною продукцією рибофлавіну шляхом одночасної надекспресії трьох генів: *RFE1*, *GND1* та *RIB6*, що кодують рибофлавін-екскретазу, 6-фосфоглюконатдегідрогеназу та 3,4-дігидрокси-2-бутанон-4-фосфатсинтазу, відповідно. Експресія комбінацій двох генів, а також коекспресія трьох генів приводила до зростання продукції рибофлавіну у *C. famata* VKM Y-9 при культивуванні на різних середовищах. Штами, з посиленою коекспресією трьох генів, характеризувалися збільшенням продукції рибофлавіну в 3,3 раза порівняно з батьківським штамом на п'ятий день культивування в середовищі з молочною сироваткою.

Ключові слова: *Candida famata*, рибофлавін, вітамін В₂, молочна сироватка, дріжджі.

REFERENCES

- Abbas CA, Sibirny AA (2011) Genetic control of biosynthesis and transport of riboflavin and flavin nucleotides and construction of robust biotechnological producers. *Microbiol Mol Biol R* 75:321–360. <https://doi.org/10.1128/MMBR.00030-10>
- Cavka A, Jönsson LJ (2014) Comparison of the growth of filamentous fungi and yeasts in lignocellulose-derived media. *Biocatal Agr Biotech* 3:197–204. <https://doi.org/10.1016/j.bcab.2014.04.003>
- Ding Q, Ye C (2023) Microbial cell factories based on filamentous bacteria, yeasts, and fungi. *Microb Cell Fact* 22:20. <https://doi.org/10.1186/s12934-023-02025-1>
- Dmytruk K, Lyzak O, Yatsyshyn V, Kluz M, Sibirny V, Puchalski C, Sibirny A (2014) Construction and fed-batch cultivation of *Candida famata* with enhanced riboflavin production. *J Biotechnol* 172:11–17. <https://doi.org/10.1016/j.jbiotec.2013.12.005>
- Dmytruk KV, Abbas CA, Voronovsky AY, Kshanovska BV, Sybirna KA, Sybirny AA (2004) Cloning of structural genes involved in riboflavin synthesis of the yeast *Candida famata*. *Ukr Biokhim Zh* (1999) 76:78–87

© ІНСТИТУТ КЛІТИННОЇ БІОЛОГІЇ ТА ГЕНЕТИЧНОЇ
ІНЖЕНЕРІЇ НАН УКРАЇНИ, 2025

- Dmytruk KV, Ruchala J, Fedorovych DV, Ostapiv RD, Sibirny AA (2020) Modulation of the purine pathway for riboflavin production in flavinogenic recombinant strain of the yeast *Candida famata*. *Biotechnol J* 15:1900468. <https://doi.org/10.1002/biot.201900468>
- Dmytruk KV, Sibirny AA (2012) *Candida famata* (*Candida flarerii*). *Yeast* 29:453–458. <https://doi.org/10.1002/yea.2929>
- Dmytruk KV, Voronovsky AY, Sibirny AA (2006) Insertion mutagenesis of the yeast *Candida famata* (*Debaryomyces hansenii*) by random integration of linear DNA fragments. *Curr Genet* 50:183–191. <https://doi.org/10.1007/s00294-006-0083-0>
- Dmytruk KV, Yatsyshyn VY, Sybirna NO, Fedorovych DV, Sibirny AA (2011) Metabolic engineering and classic selection of the yeast *Candida famata* (*Candida flarerii*) for construction of strains with enhanced riboflavin production. *Metab Eng* 13:82–88. <https://doi.org/10.1016/j.ymben.2010.10.005>
- Dzanaeva LS, Wojdyła D, Fedorovych DV, Ruchala J, Dmytruk KV, Sibirny AA (2024) Riboflavin overproduction on lignocellulose hydrolysate by the engineered yeast *Candida famata*. *FEMS Yeast Res* 24:foae020. <https://doi.org/10.1093/femsyr/foae020>
- Fischer M, Römisch W, Illarionov B, Eisenreich W, Bacher A (2005) Structures and reaction mechanisms of riboflavin synthases of eubacterial and archaeal origin. *Biochem Soc T* 33:780–784. <https://doi.org/10.1042/BST0330780>
- Kato T, Park EY (2012) Riboflavin production by *Ashbya gossypii*. *Biotechnol. Lett.* 34:611–618. <https://doi.org/10.1007/s10529-011-0833-z>
- Liu S, Hu W, Wang Z, Chen T (2020) Production of riboflavin and related cofactors by biotechnological processes. *Microb Cell Fact* 19:31. <https://doi.org/10.1186/s12934-020-01302-7>
- Petrovska Y, Lyzak O, Ruchala J, Dmytruk K, Sibirny A (2022) Co-overexpression of *RIB1* and *RIB6* increases riboflavin production in the yeast *Candida famata*. *Fermentation* 8:141. <https://doi.org/10.3390/fermentation8040141>
- Riboflavin market size, share 2023–2031, n.d. URL <https://straitsresearch.com/report/riboflavin-market> (accessed 23.09.2024)
- Ruchala J, Andreieva YA, Tsyrlunyk AO, Sobchuk SM, Najdecka A, Wen L, Kang Y, Dmytruk OV, Dmytruk KV, Fedorovych DV, Sibirny AA (2022) Cheese whey supports high riboflavin synthesis by the engineered strains of the flavinogenic yeast *Candida famata*. *Microb Cell Fact* 21:161. <https://doi.org/10.1186/s12934-022-01888-0>
- Sambrook J, Fritsch EF, Maniatis T, Russell DW, Green MR (1989) *Molecular cloning: a laboratory manual*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY
- Schwechheimer SK, Park EY, Revuelta JL, Becker J, Wittmann C (2016) Biotechnology of riboflavin. *Appl Microbiol Biotechnol* 100:2107–2119. <https://doi.org/10.1007/s00253-015-7256-z>
- Sibirny AA (2023) Metabolic engineering of non-conventional yeasts for construction of the advanced producers of biofuels and high-value chemicals. *BBA Adv* 3:100071. <https://doi.org/10.1016/j.bbadv.2022.100071>
- Stahmann K-P, Revuelta JL, Seulberger H (2000) Three biotechnical processes using *Ashbya gossypii*, *Candida famata*, or *Bacillus subtilis* compete with chemical riboflavin production. *Appl Microbiol Biotechnol* 53:509–516. <https://doi.org/10.1007/s002530051649>
- Tsyrlunyk AO, Andreieva YA, Ruchala J, Fayura LR, Dmytruk KV, Fedorovych DV, Sibirny AA (2020) Expression of yeast homolog of the mammal *BCRP* gene coding for riboflavin efflux protein activates vitamin B₂ production in the flavinogenic yeast *Candida famata*. *Yeast* 37:467–473. <https://doi.org/10.1002/yea.3470>
- Tsyrlunyk AO, Fedorovych DV, Sobchuk SM, Dmytruk KV, Sibirny AA (2021) Lactose inducible expression of transcription factor gene *SEF1* increases riboflavin production in the yeast *Candida famata*. *Mikrobiol Zh* 83:3–10. <https://doi.org/10.15407/mikrobiolzh83.05.003>
- Vincenzi A, Maciel MJ, Burlani EL, Oliveira EC, Volpato G, Lehn DN, Souza CFV (2014) Ethanol bio-production from Ricotta cheese whey by several strains of the yeast *Kluyveromyces*. *Am J Food Technol* 9:281–291. <https://doi.org/10.3923/ajft.2014.281.291>
- Voronovsky A, Abbas C, Fayura L, Kshanovska B, Dmytruk K, Sybirna K, Sibirny A (2002) Development of a transformation system for the flavinogenic yeast *Candida famata*. *FEMS Yeast Res* 2:381–388. [https://doi.org/10.1016/S1567-1356\(02\)00112-5](https://doi.org/10.1016/S1567-1356(02)00112-5)
- Voronovsky AY, Abbas CA, Dmytruk KV, Ishchuk OP, Kshanovska BV, Sybirna KA, Gaillardin C, Sibirny AA (2004) *Candida famata* (*Debaryomyces hansenii*) DNA sequences containing genes involved in riboflavin synthesis. *Yeast* 21:1307–1316. <https://doi.org/10.1002/yea.1182>
- Wang Y, Liu L, Jin Z, Zhang D (2021) Microbial cell factories for green production of vitamins. *Front Bioeng Biotechnol* 9:661562. <https://doi.org/10.3389/fbioe.2021.661562>
- You J, Yang C, Pan X, Hu M, Du Y, Osire T, Yang T, Rao Z (2021) Metabolic engineering of *Bacillus subtilis* for enhancing riboflavin production by alleviating dissolved oxygen limitation. *Biores Technol* 333:125228. <https://doi.org/10.1016/j.biortech.2021.125228>

Received October 18, 2024
Received November 05, 2024
Accepted January 18, 2025