

SURVEYING SELECTION SIGNATURES IN MURRAH BUFFALO USING GENOME-WIDE SNP DATA

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The population of in-milk buffaloes climbed by 4.3 % in 2012–2019, as compared to the decline of 28.9 % in 2007–2012, suggesting a trend reversal towards buffalo rearing due to consumer preferences and policy interventions. Moreover, Murrah buffaloes have quadrupled in the same period from 11.7 million to 47.06 million headcounts, constituting about 42.8 % of the total buffaloes. Analysis of selection signatures reveals important details about how the genomic environments of contemporary livestock have been altered by both natural and artificial selective pressures. In the present study to delineate signals of positive selection, a total of 246 million (98.08 %) clean ddRAD-seq reads were mapped to Bubalus bubalis reference genome assembly. CLR approach was used to detect selected regions and a total of 289 selection signatures were found in Murrah genome. The outlier fraction containing 289 sites, bracketed by a 10 kb window, 5 kb up- and down- stream were declared as the putative selection signatures. A total of 106 genes and 22 loci were traced in 179 selective sweep regions in Murrah buffalo. The information mined in this study will aid in future polymorphism studies of economic traits in buffaloes.

Key words: selection; CLR; murrah; ddRAD-seq; genome.

ДОСЛІДЖЕННЯ ХАРАКТЕРНИХ СЕЛЕКЦІЙНИХ ОЗНАК БУЙВОЛІВ ПОРОДИ МУРРА З ВИКОРИСТАННЯМ ДАНИХ SNP ПО ВСЬОМУ ГЕНОМУ

У 2012–2019 рр. популяція дійних буйволиць зросла на 4,3 % порівняно зі спадом на 28,9 % у 2007–2012, що дозволяє припустити зміну тенденції

щодо розведення буйволів внаслідок преференцій споживачів та втручань у політику. Крім того, поголів'я буйволів породи Мурра за той же період зросло в чотири рази – з 11,7 млн до 47,06 млн голів, що становить близько 42,8 % від загального поголів'я буйволів. Аналіз селекційних ознак розкриває важливі деталі того, як геномне середовище сучасної худоби було змінено під природним і штучним селективним тиском. У цьому дослідженні для визначення сигналів позитивного добору було зіставлено 246 мільйонів (98,08 %) чистих ddRAD-секвенів на референтну збірку генома *Bubalus bubalis*. Для виявлення відібраних ділянок було використано підхід CLR, і в цілому в геномі Мурра було знайдено 289 селекційних ознак. Фракція викидів, що містить 289 ділянок, розділених вікном розміром 10 кб, по 5 кб вгору і вниз, була визначена як передбачувані селекційні ознаки. Загалом було відстежено 106 генів і 22 локуси в 179 регіонах селективного розгортання буйволів Мурра. Інформація, отримана в цьому дослідженні, буде корисною для майбутніх досліджень поліморфізму економічно цінних ознак буйволів.

Ключові слова: селекція, CLR, Мурра, ddRAD-секвени, геном.

REFERENCES

- Albooshoke, S.N., and Bakhtiarizadeh, M.R., Divergent gene expression through PI3K/akt signalling pathway cause different models of hypertrophy growth in chicken, *Italian J. Anim. Sci.*, 2019, vol. 18, no. 1, pp. 1161–1173. <https://doi.org/10.1080/1828051X.2019.1634498>
- Alomer, R.M., da Silva, E.M., Chen, J., Piekarz, K.M., McDonald, K., Sansam, C.G., et al., Escol and Esco2 regulate distinct cohesin functions during cell cycle progression, *Proc. Nat. Acad. Scie.*, 2017, vol. 114, no. 37, pp. 9906–9911. <https://doi.org/10.1073/pnas.1708291114>
- Andrews, S., FastQC: a quality control tool for high throughput sequence data, 2010
- Azmal, S.A., Bhuiyan, A.A., Omar, A.I., Ma, S., Sun, C., Han, Z., et al., Novel polymorphisms in RAPGEF6 gene associated with egg-laying rate in Chinese Jing Hong chicken using genome-wide SNP scan, *Genes*, 2019, vol. 10 no. 5, pp. 384. <https://doi.org/10.3390/genes10050384>

- Bahbahani, H., Clifford, H., Wragg, D., Mbole-Kariuki, M. N., Van Tassell, C., Sonstegard, T., et al., Signatures of positive selection in East African Shorthorn Zebu: A genome-wide single nucleotide polymorphism analysis, *Scientific Reports*, 2015, vol. 5, no. 1, pp. 11729. <https://doi.org/10.1038/srep11729>
- Bahbahani, H., Salim, B., Almathen, F., Al Enezi, F., Mwacharo, J.M., and Hanotte, O., Signatures of positive selection in African Butana and Kenana dairy zebu cattle. *PLoS one*, 2018, vol. 13, no. 1, pp. e0190446. <https://doi.org/10.1371/journal.pone.0190446>
- Banerjee, S., Batabyal, K., Joardar, S.N., Isore, D.P., Dey, S., Samanta, I., et al., Detection and characterization of pathogenic *Pseudomonas aeruginosa* from bovine subclinical mastitis in West Bengal, India, *Veterinary world*, 2017, vol. 10, no. 7, pp. 738. <https://doi.org/10.14202%2Fvetworld.2017.738-742>
- Barazandeh, A., Mohammadabadi, M., Ghaderi-Zefrehei, M., Rafeie, F., and Imumorin, I.G., Whole genome comparative analysis of CpG islands in camelid and other mammalian genomes, *Mammalian Biol.*, 2019, vol. 98, pp. 73–79. <https://doi.org/10.1016/j.mambio.2019.07.007>
- Basic Animal Husbandry Statistics, BAHs, Department of Animal Husbandry, Dairying & Fisheries (DAHD), 2022, GOI. <https://dahd.nic.in/schemes/programmes/animal-husbandry-statistics>
- Blay, C., Haffray, P., Bugeon, J., D'ambrosio, J., Dechamp, N., Collewet, G., et al., Genetic parameters and genome-wide association studies of quality traits characterised using imaging technologies in Rainbow trout, *Oncorhynchus mykiss*, *Frontiers in Genetics*, 2021, vol. 12, pp. 639223. <https://doi.org/10.3389/fgene.2021.639223>
- Bolen, H., Genetic and genomic factors influencing gestational length in beef cattle, Ph.D. Thesis (University of Saskatchewan, 2022). <https://hdl.handle.net/10388/14184>
- Bonhomme, M., Chevalet, C., Servin, B., Boitard, S., Abdallah, J., Blott, S., and SanCristobal, M., Detecting selection in population trees: the Lewontin and Krakauer test extended, *Genetics*, 2010, vol. 186, no. 1, pp. 241–262. <https://doi.org/10.1534/genetics.110.117275>
- Boughdaira, I.B., The centrin-binding protein Sfi1: functions in fission yeast and human, Ph.D. thesis (Université Paris-Saclay, 2017)
- Bouvard, D., Pouwels, J., De Franceschi, N., and Ivaska, J., Integrin inactivators: balancing cellular functions in vitro and in vivo, *Nature Reviews Molecular Cell Biology*, 2013, vol. 14, no. 7, pp. 430–442. <https://doi.org/10.1038/nrm3599>
- Braverman, J.M., Hudson, R.R., Kaplan, N.L., Langley, C.H., and Stephan, W., The hitchhiking effect on the site frequency spectrum of DNA polymorphisms, *Genetics*, 1995, vol. 140, pp. 783–796. <https://doi.org/10.1093/genetics/140.2.783>
- Cadigan, K.M., and Waterman, M.L., TCF/LEFs and Wntsignaling in the nucleus, *Cold Spring Harbor perspectives in biology*, 2012, vol. 4, no. 11, a007906. <https://doi.org/10.1101/cshperspect.a007906>
- Campanile, G., Neglia, G., Gasparrini, B., Galiero, G., Prandi, A., Di Palo, R., et al., Embryonic mortality in buffaloes synchronized and mated by AI during the seasonal decline in reproductive function, *Theriogenology*, 2005, vol. 63, no. 8, pp. 2334–2340. <https://doi.org/10.1016/j.theriogenology.2004.10.012>
- Cánovas, A., Rincyn, G., Islas-Trejo, A., Jimenez-Flores, R., Laubscher, A., and Medrano, J.F., RNA sequencing to study gene expression and single nucleotide polymorphism variation associated with citrate content in cow milk, *J. Dairy Sci.*, 2013, vol. 96, no. 4, pp. 2637–2648. <https://doi.org/10.3168/jds.2012-6213>
- Capuco, A.V., and Choudhary, R.K., Symposium review: Determinants of milk production: Understanding population dynamics in the bovine mammary epithelium, *J. Dairy Sci.*, 2020, vol. 103, no. 3, pp. 2928–2940. <https://doi.org/10.3168/jds.2019-17241>
- Catchen, J.M., Amores, A., Hohenlohe, P., Cresko, W., and Postlethwait, J.H., Stacks: building and genotyping loci de novo from short-read sequences, *G3: Genes|Genomes|Genetics*, 2011, vol. 1, no. 3, pp. 171–182. <https://doi.org/10.1534/g3.111.000240>
- Cerri, R.L.A., Thompson, I.M., Kim, I.H., Ealy, A.D., Hansen, P.J., Staples, C.R., et al., Effects of lactation and pregnancy on gene expression of endometrium of Holstein cows at day 17 of the estrous cycle or pregnancy, *J. Dairy Sci.*, 2012, vol. 95, no. 10, pp. 5657–5675. <https://doi.org/10.3168/jds.2011-5114>
- Chao, J., Li, Z., Sun, Y., Aluko, O.O., Wu, X., Wang, Q., and Liu, G., MG2C: A user-friendly online tool for drawing genetic maps, *Molecular Horticulture*, 2021, vol. 1, no. 1, pp. 1–4. <https://doi.org/10.1186/s43897-021-00020-x>
- Choi, S.K., Kim, M.J., and You, J.S., SMARCB1 acts as a quiescent gatekeeper for cell cycle and immune response in human cells, *Inter. J. Mol. Sci.*, 2020, vol. 21, no. 11, pp. 3969. <https://doi.org/10.3390/ijms21113969>
- Choi, Y., Ballow, D.J., Xin, Y., and Rajkovic, A., Lim homeobox gene, *lhx8*, is essential for mouse oocyte differentiation and survival, *Biol. Reprod.*, 2008, vol. 79, no. 3, pp. 442–449. <https://doi.org/10.1095/biolreprod.108.069393>
- Dai, B., Ren, L.Q., Han, X.Y., and Liu, D.J., Bioinformatics analysis reveals 6 key biomarkers associated with non-small-cell lung cancer, *J. Inter. Med. Res.*, 2020, vol. 48, no. 3, 0300060519887637. <https://doi.org/10.1177/0300060519887637>

- Danecek, P., Bonfield, J.K., Liddle, J., Marshall, J., Ohan, V., Pollard, M.O., et al., Twelve years of SAMtools and BCFtools, *Gigascience*, 2021, vol. 10, no. 2, giab008. <https://doi.org/10.1093/gigascience/giab008>
- Davoodi, S., Cooke, R.F., Fernandes, A.C.D.C., Cappellozza, B.I., Vasconcelos, J.L.M., and Cerri, R.L.A., Expression of estrus modifies the gene expression profile in reproductive tissues on day 19 of gestation in beef cows, *Theriogenology*, 2016, vol. 85, no. 4, pp. 645–655. <https://doi.org/10.1016/j.theriogenology.2015.10.002>
- Deng, T.X., Ma, X.Y., Lu, X.R., Duan, A.Q., Shokrollahi, B., and Shang, J. H., Signatures of selection reveal candidate genes involved in production traits in Chinese crossbred buffaloes, *J. Dairy Sci.*, 2022, vol. 105, no. 2, pp. 1327–1337. <https://doi.org/10.3168/jds.2021-21102>
- Devarajan, A., Bourquard, N., Grijalva, V.R., Gao, F., Ganapathy, E., Verma, J., and Reddy, S.T., Role of PON2 in innate immune response in an acute infection model, *Mol. Genet. Metabol.*, 2013, vol. 110, no. 3, pp. 362–370. <https://doi.org/10.1016/j.ymgme.2013.07.003>
- Ding, G., Ma, T., Zhang, K., Chen, G., Shen, J., Zhang, S., et al., A pan-cancer analysis of the role of WDFY2 in human tumors, *Biotechnol. Genet. Engineer. Rev.*, 2023, pp. 1–16
- doNascimento, A.V., Scanning of Selection Signatures in Dairy Buffalo *Ph.D. thesis (Bubalus bubalis)* (Universidade Estadual Paulista (Unesp). Faculdade de Ciências Agrárias e Veterinárias. Jaboticabal, 2020). <http://hdl.handle.net/11449/193499>
- Fan, W., Liu, X., Zhang, J., Qin, L., Du, J., Li, X., et al., TRIM67 suppresses TNF α -triggered NF- κ B activation by competitively binding beta-TrCP to I κ B α , *Front. Immunol.*, 2022, vol. 13, 793147. <https://doi.org/10.3389/fimmu.2022.793147>
- Fariello, M.I., Boitard, S., Naya, H., SanCristobal, M., and Servin, B., Detecting signatures of selection through haplotype differentiation among hierarchically structured populations, *Genetics*, 2013, vol. 193, no. 3, pp. 929–941. <https://doi.org/10.1534/genetics.112.147231>
- Fay, J.C., and Wu, C.I., Hitchhiking under positive Darwinian selection, *Genetics*, 2000, vol. 155, no. 3, pp. 1405–1413. <https://doi.org/10.1093/genetics/155.3.1405>
- Friedrich, J., and Wiener, P., Selection signatures for high-altitude adaptation in ruminants, *Anim. Genet.*, 2020, vol. 51, no. 2, pp. 157–165. <https://doi.org/10.1111/age.12900>
- Gao, Y., Jiang, J., Yang, S., Hou, Y., Liu, G.E., Zhang, S., et al., CNV discovery for milk composition traits in dairy cattle using whole genome resequencing, *BMC genomics*, 2017, vol. 18, pp. 1–12. <https://doi.org/10.1186/s12864-017-3636-3>
- Ge, S.X., Jung, D., and Yao, R., ShinyGO: a graphical gene-set enrichment tool for animals and plants, *Bioinformatics*, 2020, vol. 36, no. 8, pp. 2628–2629. <https://doi.org/10.1093/bioinformatics/btz931>
- George, L., Alex, R., Sukhija, N., Jaglan, K., Vohra, V., Kumar, R., and Verma, A., Genetic improvement of economic traits in Murrah buffalo using significant SNPs from genome-wide association study, *Tropical Animal Health and Production*, 2023, vol. 55, no. 3, pp. 199. <https://doi.org/10.1007/s11250-023-03606-3>
- Gouveia, J.J.D.S., Silva, M.V.G.B.D., Paiva, S.R., and Oliveira, S.M.P.D., Identification of selection signatures in livestock species, *Genet. Mol. Biol.*, 2014, vol. 37, pp. 330–342. <https://doi.org/10.1590/S1415-47572014000300004>
- Grădinaru, A.C., Ciopeică, C., and Munteanu, A., Congenital supernumerary teats in cows: a review associated with personal observations, *Anim. Biol. Anim. Husband.*, 2021, vol. 13, no. 2
- Grossi, D.A., Abo-Ismael, M.K., Koeck, A., Miller, S.P., Stothard, P., Plastow, G., et al., Genome-wide Association Analyses for Mastitis in Canadian Holsteins, In *Proceedings of the 10th World Congress on Genetics Applied to Livestock Production, ASAS, Vancouver, BC, Canada*, 2014, pp. 17–22
- Hall, E.G., Characterization of SPECC1L Function in Palatogenesis, Ph. D. thesis (University of Kansas, 2019)
- Han, H., Randhawa, I.A., MacHugh, D.E., McGivney, B.A., Katz, L.M., Dugarjaviin, M., and Hill, E.W., Selection signatures for local and regional adaptation in Chinese Mongolian horse breeds reveal candidate genes for hoof health, *BMC Genomics*, 2023, vol. 24, no. 1, pp. 1–11. <https://doi.org/10.1186/s12864-023-09116-8>
- Hao, Q., Samten, B., Ji, H.L., Zhao, Z.J., and Tang, H., Tyrosine phosphatase PTP-MEG2 negatively regulates vascular endothelial growth factor receptor signaling and function in endothelial cells, *Amer. J. Physiol.-Cell Physiol.*, 2012, vol. 303, no. 5, pp. C548–C553. <https://doi.org/10.1152/ajpcell.00415.2011>
- Haydar, S., Grigorescu, F., Vintilă, M., Cogne, Y., Lautier, C., Tutuncu, Y., et al., Fine-scale haplotype mapping of MUT, AACS, SLC6A15 and PRKCA genes indicates association with insulin resistance of metabolic syndrome and relationship with branched chain amino acid metabolism or regulation, *PloS one*, 2019, vol. 14, no. 3, e0214122. <https://doi.org/10.1371/journal.pone.0214122>
- He, W., Fang, X., Lu, X., Liu, Y., Li, G., Zhao, Z., et al., Function Identification of Bovine ACSF3 Gene and Its Association With Lipid Metabolism Traits in Beef Cattle, *Front. Veterin. Sci.*, 2022, vol. 8, 766765. <https://doi.org/10.3389/fvets.2021.766765>

- Hosseinzadeh, S., and Hasanpur, K., Gene expression networks and functionally enriched pathways involved in the response of domestic chicken to acute heat stress, *Front. Genet.*, 2023. vol. 14, 1102136. <https://doi.org/10.3389/fgene.2023.1102136>
- Huang, L., Hua, Z., Xiao, H., Cheng, Y., Xu, K., Gao, Q., et al., CRISPR/Cas9-mediated ApoE^{-/-} and LDLR^{-/-} double gene knockout in pigs elevates serum LDL-C and TC levels, *Oncotarget.*, 2017, no. 8, no. 23, 37751. <https://doi.org/10.18632/oncotarget.17154>
- Huang, Q., Ma, C., Chen, L., Luo, D., Chen, R., and Liang, F., Mechanistic insights into the interaction between transcription factors and epigenetic modifications and the contribution to the development of obesity, *Front. Endocrinol.*, 2018, vol. 9, pp. 370. <https://doi.org/10.3389/fendo.2018.00370>
- Hung, T., Pratt, G.A., Sundararaman, B., Townsend, M.J., Chaivorapol, C., Bhangale, T., et al., The Ro60 autoantigen binds endogenous retroelements and regulates inflammatory gene expression, *Science*, 2015, vol. 350, no. 6259, pp. 455–459. <https://doi.org/10.1126/science.aac7442>
- Huynh, H., Bottini, N., Williams, S., Cherepanov, V., Musumeci, L., Saito, K., Bruckner, S., Vachon, E., Wang, X., Kruger, J., Chow, C.-W., Pellecchia, M., Monosov, E., Greer, P.A., Trimble, W., Downey, G.P., and Mustelin, T., Control of vesicle fusion by a tyrosine phosphatase, *Nat. Cell Biol.*, 2004, vol. 6, no. 9, pp. 831–839. <https://doi.org/10.1038/ncb1164>
- Jaglan, K., Ravikumar, D., Sukhija, N., George, L., Alex, R., Vohra, V., and Verma, A., Genomic clues of association between clinical mastitis and SNPs identified by ddRAD sequencing in Murrah buffaloes, *Anim. Biotechnol.*, 2023, pp. 1–9. <https://doi.org/10.1080/10495398.2023.2165937>
- Jovanovic, V.M., Sarfert, M., Reyna-Blanco, C.S., Indrischek, H., Valdivia, D.I., Shelest, E., and Nowick, K., Positive selection in gene regulatory factors suggests adaptive pleiotropic changes during human evolution, *Front. Genet.*, 2021, vol. 12, 662239. <https://doi.org/10.3389/fgene.2021.662239>
- Julien, S., Bobowski, M., Steenackers, A., Le Bourhis, X., and Delannoy, P., How do gangliosides regulate RTKs signaling?, *Cells*, 2013, vol. 2, no. 4, pp. 751–767. <https://doi.org/10.3390/cells2040751>
- Kandasamy, S., Jain, A., Kumar, R., Agarwal, S.K., Joshi, P., and Mitra, A., Molecular characterization and expression profile of uterine serpin (SERPINA14) during different reproductive phases in water buffalo (*Bubalus bubalis*), *Anim. Reprod. Sci.*, 2010, vol. 122, no. 1–2, pp. 133–141. <https://doi.org/10.1016/j.ani-reprosci.2010.08.005>
- Khan, M.Z., Khan, A., Xiao, J., Ma, Y., Ma, J., Gao, J., and Cao, Z., Role of the JAK-STAT pathway in bovine mastitis and milk production, *Animals*, 2020, vol. 10, no. 11, pp. 2107. <https://doi.org/10.3390/ani10112107>
- Kim, Y., and Stephan, W., Detecting a local signature of genetic hitchhiking along a recombining chromosome, *Genetics*, 2002, vol. 160, no. 2, pp. 765–777. <https://doi.org/10.1093/genetics/160.2.765>
- Klein, S.L., Yin, T., Swalve, H.H., and Kunig, S., Single-step genomic best linear unbiased predictor genetic parameter estimations and genome-wide associations for milk fatty acid profiles, interval from calving to first insemination, and ketosis in Holstein cattle, *J. Dairy Sci.*, 2021, vol. 104, no. 10, pp. 10921–10933. <https://doi.org/10.3168/jds.2021-20416>
- Kuhlwilm, M., and Boeckx, C., A catalog of single nucleotide changes distinguishing modern humans from archaic hominins, *Scientific Reports*, 2019, vol. 9, no. 1, pp. 8463. <https://doi.org/10.1038/s41598-019-44877-x>
- Kumar, A., Rajendran, V., Sethumadhavan, R., and Purohit, R., CEP proteins: the knights of centrosome dynasty, *Protoplasma*, 2013, vol. 250, pp. 965–983. <https://doi.org/10.1007/s00709-013-0488-9>
- Kumar, D.R., Devadasan, M.J., Surya, T., Vineeth, M.R., Choudhary, A., Sivalingam, J., et al., Genomic diversity and selection sweeps identified in Indian swamp buffaloes reveals its uniqueness with riverine buffaloes, *Genomics*, 2020, vol. 112, no. 3, pp. 2385–2392. <https://doi.org/10.1016/j.ygeno.2020.01.010>
- Langmead, B., and Salzberg, S.L., Fast gapped-read alignment with Bowtie 2, *Nature methods*, 2012, vol. 9, no. 4, pp. 357–359. <https://doi.org/10.1038/nmeth.1923>
- Li, H., A statistical framework for SNP calling, mutation discovery, association mapping and population genetic parameter estimation from sequencing data, *Bioinformatics*, 2011, vol. 27, no. 21, pp. 2987–2993. <https://doi.org/10.1093/bioinformatics/btr509>
- Li, X., Yang, S., Dong, K., Tang, Z., Li, K., Fan, B., et al., Identification of positive selection signatures in pigs by comparing linkage disequilibrium variances, *Anim. Genet.*, 2017, vol. 48, no. 5, pp. 600–605. <https://doi.org/10.1111/age.12574>
- Lin, S., Xian, M., Ren, T., Mo, G., Zhang, L., and Zhang, X., Mining of chicken muscle growth genes and the function of important candidate gene RPL3L in muscle development, *Front. Physiol.*, 2022, pp. 2295. <https://doi.org/10.3389/fphys.2022.1033075>
- Lindsay, B.G., Composite likelihood methods, *Contemporary Mathematics*, 1988, vol. 80, no. 1, pp. 221–239
- Liu, X., Li, Z., Yan, Y., Li, Y., Wu, H., Pei, J., et al., Selection and introgression facilitated the adaptation of Chinese native endangered cattle in extreme environments, *Evolution. Applicat.*, 2021, vol. 14, no. 3, pp. 860–873. <https://doi.org/10.1111/eva.13168>
- Ludwig, D., Carter, J., Smith, J.R., Borsani, G., Barla-

- ti, S., and Hafizi, S., Functional characterisation of human cells harbouring a novel t (2p; 7p) translocation involving TNS3 and EXOC6B genes, *BMC Med. Genet.*, 2013, vol. 14, pp. 1–10. <https://doi.org/10.1186/1471-2350-14-65>
- Ma, J., Gao, X., Li, J., Gao, H., Wang, Z., Zhang, L., et al., Assessing the genetic background and selection signatures of Huaxi cattle using high-density SNP array, *Animals*, 2021, vol. 11, no. 12, pp. 3469. <https://doi.org/10.3390/ani11123469>
- Malik, A.A., Sharma, R., Ahlawat, S., Deb, R., Negi, M. S., and Tripathi, S.B., Analysis of genetic relatedness among Indian cattle (*Bos indicus*) using genotyping-by-sequencing markers, *Anim. Genet.*, 2018, vol. 49, no. 3, pp. 242–245. <https://doi.org/10.1111/age.12650>
- Mishra, D.C., Sikka, P., Yadav, S., Bhati, J., Paul, S.S., Jerome, A., et al., Identification and characterization of trait-specific SNPs using ddRAD sequencing in water buffalo, *Genomics*, 2020, vol. 112, no. 5, pp. 3571–3578. <https://doi.org/10.1016/j.ygeno.2020.04.012>
- Munoz, M., Rodríguez, M.C., Alves, E., Folch, J.M., Ibáñez-Escriche, N., Siliy, L., and Fernández, A.I., Genome-wide analysis of porcine backfat and intramuscular fat fatty acid composition using high-density genotyping and expression data, *BMC Genomics*, 2013, vol. 14, pp. 1–15. <https://doi.org/10.1186/1471-2164-14-845>
- Nakahara, K., Ohkuni, A., Kitamura, T., Abe, K., Naganuma, T., Ohno, Y., et al., The Sjögren-Larsson syndrome gene encodes a hexadecenal dehydrogenase of the sphingosine 1-phosphate degradation pathway, *Mol. Cell*, 2012, vol. 46, no. 4, pp. 461–471. <http://dx.doi.org/10.1016%2Fj.molcel.2012.04.033>
- Nielsen, R., Hellmann, I., Hubisz, M., Bustamante, C., and Clark, A.G., Recent and ongoing selection in the human genome, *Nat. Rev. Genet.*, 2007, vol. 8, no. 11, pp. 857–868. <https://doi.org/10.1038/nrg2187>
- Nielsen, R., Williamson, S., Kim, Y., Hubisz, M.J., Clark, A.G., and Bustamante, C., Genomic scans for selective sweeps using SNP data, *Genome Res.*, 2005, vol. 15, no. 11, pp. 1566–1575. <https://doi.org/10.1101/gr.4252305>
- Oleksyk, T.K., Smith, M.W., and O'Brien, S.J., Genome-wide scans for footprints of natural selection, *Philosophical Transactions of the Royal Society B: Biological Sciences*, 2010, vol. 365, no. 1537, pp. 185–205. <https://doi.org/10.1098/rstb.2009.0219>
- Palmieri, L., Pardo, B., Lasorsa, F.M., Del Arco, A., Kobayashi, K., Iijima, M., et al., Citrin and aralar1 are Ca²⁺-stimulated aspartate/glutamate transporters in mitochondria, *The EMBO Journal*, 2001, vol. 20, no. 18, pp. 5060–5069. <https://doi.org/10.1093/emboj/20.18.5060>
- Pausch, H., Jung, S., Edel, C., Emmerling, R., Krogmeier, D., Götz, K.U., and Fries, R., Genome-wide association study uncovers four QTL predisposing to supernumerary teats in cattle, *Anim. Genet.*, 2012, vol. 43, no. 6, pp. 689–695. <https://doi.org/10.1111/j.1365-2052.2012.02340.x>
- Pavlidis, P., Živković, D., Stamatakis, A., and Alachiotis, N., SweeD: likelihood-based detection of selective sweeps in thousands of genomes, *Mol. Biol. Evolut.*, 2013, vol. 30, no. 9, pp. 2224–2234. <https://doi.org/10.1093/molbev/mst112>
- Pirooznia, M., Nagarajan, V., and Deng, Y., GeneVenn-A web application for comparing gene lists using Venn diagrams, *Bioinformatics*, 2007, vol. 1, no. 10, pp. 420. <https://doi.org/10.6026/97320630001420>
- Popova, V.V., Brechalov, A.V., Georgieva, S.G., and Kopytova, D.V., Nonreplicative functions of the origin recognition complex, *Nucleus*, 2018, vol. 9, no. 1, pp. 460–473. <https://doi.org/10.1080/19491034.2018.1516484>
- Prakapenka, D., Liang, Z., Jiang, J., Ma, L., and Da, Y., A large-scale genome-wide association study of epistasis effects of production traits and daughter pregnancy rate in uS Holstein cattle, *Genes*, 2021, vol. 12, no. 7, pp. 1089. <https://doi.org/10.3390/genes12071089>
- Prezeworski, M., Coop, G., and Wall, J.D., The signature of positive selection on standing genetic variation, *Evolution*, 2005, vol. 59, no. 11, pp. 2312–2323. <https://doi.org/10.1111/j.0014-3820.2005.tb00941.x>
- Pritchard, J.K., Pickrell, J.K., and Coop, G., The genetics of human adaptation: hard sweeps, soft sweeps, and polygenic adaptation, *Curr. Biol.*, 2010, vol. 20, no. 4, pp. 208–215
- Qanbari, S., and Simianer, H., Mapping signatures of positive selection in the genome of livestock, *Livestock Science*, 2014, vol. 166, pp. 133–143. <https://doi.org/10.1016/j.livsci.2014.05.003>
- Quinlan, A.R., and Hall, I.M., BEDTools: a flexible suite of utilities for comparing genomic features, *Bioinformatics*, 2010, vol. 26, no. 6, pp. 841–842. <https://doi.org/10.1093/bioinformatics/btq033>
- Raudvere, U., Kolberg, L., Kuzmin, I., Arak, T., Adler, P., Peterson, H., and Vilo, J., g: Profiler: a web server for functional enrichment analysis and conversions of gene lists (2019 update), *Nucl. Acids Res.*, 2019, 47, no. 1, pp. 191–198. <https://doi.org/10.1093/nar/gkz369>
- Ravi Kumar, D., Nandhini, P.B., Joel Devadasan, M., Sivalingam, J., Mengistu, D.W., Verma, A., et al., Genome-wide association study revealed suggestive QTLs for production and reproduction traits in Indian Murrah buffalo, *3 Biotech*, 2023, vol. 13, no. 3, pp. 100. <https://doi.org/10.1007/s13205-023-03505-2>
- Rocha, R.D.F.B., Garcia, A.O., Otto, P.I., da Silva, M.V.B., Martins, M.F., Machado, M.A., et al.,

- Runs of homozygosity and signatures of selection for number of oocytes and embryos in the Gir Indicine cattle, *Mammalian Genome*, 2023, pp. 1–15. <https://doi.org/10.1007/s00335-023-09989-w>
- Runa, F., Adamian, Y., and Kelber, J.A., Ascending the PEAK1 toward targeting TGF β during cancer progression: recent advances and future perspectives, *Cancer Cell Microenviron.*, 2016, vol. 3, no. 1. <https://doi.org/10.14800%2Fccm.1162>
- Sabeti, P.C., Varilly, P., Fry, B., Lohmueller, J., Hostetter, E., Cotsapas, C., et al., Genome-wide detection and characterization of positive selection in human populations, *Nature*, 2007, vol. 449 no. 7164, pp. 913–918. <https://doi.org/10.1038/nature06250>
- Sánchez-Ramos, R., Trujano-Chavez, M.Z., Gallegos-Sánchez, J., Becerril-Pérez, C.M., Cadena-Villegas, S., and Cortez-Romero, C., Detection of Candidate Genes Associated with Fecundity through Genome-Wide Selection Signatures of Katahdin Ewes, *Animals*, 2023, vol. 13, no. 2, pp. 272. <https://doi.org/10.3390/ani13020272>
- Saravanan, K.A., Panigrahi, M., Kumar, H., Bhushan, B., Dutt, T., and Mishra, B.P., Selection signatures in livestock genome: A review of concepts, approaches and applications, *Livestock Sci.*, 2020, vol. 241, 104257. <https://doi.org/10.1016/j.livsci.2020.104257>
- Saravanan, K.A., Rajawat, D., Kumar, H., Nayak, S.S., Bhushan, B., Dutt, T., and Panigrahi, M., Signatures of selection in riverine buffalo populations revealed by genome-wide SNP data, *Anim. Biotechnol.*, 2022, pp. 1–12. <https://doi.org/10.1080/10495398.2022.2145292>
- Schellhaus, A.K., Moreno-Andrés, D., Chugh, M., Yokoyama, H., Moschopoulou, A., De, S., et al., Developmentally Regulated GTP binding protein 1 (DRG1) controls microtubule dynamics, *Scientific Rep.*, 2017, vol. 7, no. 1, 9996. <https://doi.org/10.1038/s41598-017-10088-5>
- Schmieder, R., and Edwards, R., Quality control and preprocessing of metagenomic datasets, *Bioinformatics*, 2011, vol. 27, no. 6, pp. 863–864. <https://doi.org/10.1093/bioinformatics/btr026>
- Seabury, C.M., Oldeschulte, D.L., Saatchi, M., Beaver, J.E., Decker, J.E., Halley, Y.A., et al., Genome-wide association study for feed efficiency and growth traits in US beef cattle, *BMC Genom.*, 2017, vol. 18, no. 1, pp. 1–25. <https://doi.org/10.1186/s12864-017-3754-y>
- Seifi Moroudi, R., Ansari Mahyari, S., Vaez Torshizi, R., Lanjanian, H., and Masoudi-Nejad, A., Identification of new genes and quantitative trait loci associated with growth curve parameters in F2 chicken population using genome-wide association study, *Anim. Genet.*, 2021, vol. 52, no. 2, pp. 171–184. <https://doi.org/10.1111/age.13038>
- Shaheen, R., Faeih, E., Sunker, A., Morsy, H., Al-Sheddi, T., Shamseldin, H.E., et al., Recessive mutations in DOCK6, encoding the guanidine nucleotide exchange factor DOCK6, lead to abnormal actin cytoskeleton organization and Adams-Oliver syndrome, *Amer. J. Human Genet.*, 2011, vol. 89, no. 2, pp. 328–333. <https://doi.org/10.1016/j.ajhg.2011.07.009>
- Smith, J. M., and Haigh, J., The hitch-hiking effect of a favourable gene, *Genet. Res.*, 1974, vol. 23, no. 1, pp. 23–35. <https://doi.org/10.1017/S0016672300014634>
- Stegemiller, M.R., Ellison, M.J., Hall, J.B., Sprinkle, J.E., and Murdoch, B.M., Identifying genetic variants affecting cattle grazing behavior experiencing mild heat load, *Translat. Anim. Sci.*, 2021, vol. 5, pp. 61–66. <https://doi.org/10.1093/tas/txab151>
- Stephan, W., Selective sweeps, *Genetics*, 2019, vol. 211, no. 1, pp. 5–13. <https://doi.org/10.1534/genetics.118.301319>
- Su, T.C., Lin, S.H., Lee, P.T., Yeh, S.H., Hsieh, T.H., Chou, S.Y., et al., The sigma-1 receptor-zinc finger protein 179 pathway protects against hydrogen peroxide-induced cell injury, *Neuropharmacology*, 2016, vol. 105, pp. 1–9. <https://doi.org/10.1016/j.neuropharm.2016.01.015>
- Sukhija, N., Malik, A.A., Devadasan, J.M., Dash, A., Bidyalaxmi, K., Ravi Kumar, D., et al., Genome-wide selection signatures address trait specific candidate genes in cattle indigenous to arid regions of India, *Anim. Biotechnol.*, 2023, pp. 1–15. <https://doi.org/10.1080/10495398.2023.2290521>
- Surya, T., Vineeth, M.R., Sivalingam, J., Tantia, M.S., Dixit, S.P., Niranjana, S.K., and Gupta, I.D., Genomewide identification and annotation of SNPs in *Bubalus bubalis*, *Genomics*, 2019, vol. 111, no. 6, pp. 1695–1698. <https://doi.org/10.1016/j.ygeno.2018.11.021>
- Tajima, F., Statistical method for testing the neutral mutation hypothesis by DNA polymorphism, *Genetics*, 1989, vol. 123, no. 3, pp. 585–595. <https://doi.org/10.1093/genetics/123.3.585>
- Tang, H., Zhang, H., Liu, D., Wang, Z., Yu, D., and Fan, W., Genome-wide association study reveals the genetic determinism of serum biochemical indicators in ducks, *BMC genomics*, 2022, vol. 23, no. 1, pp. 1–13. <https://doi.org/10.1186/s12864-022-09080-9>
- Toro-Ospina, A.M., Herrera Rios, A.C., Bizarria Santos, W., Pimenta Schettini, G., Vallejo Aristizabal, V.H., Tovar Claros, G., and Morea, E.G.O., Genetic architecture and signatures of selection in the Caqueteco creole (Colombian native cattle), *Diversity*, 2022, vol. 14, no. 10, pp. 828. <https://doi.org/10.3390/d14100828>
- Turner, S.D., qqman: an R package for visualizing GWAS results using QQ and manhattan plots, *J. Open Source Softw.*, 2018, vol. 3, pp. 731

- Tyagi, S.K., Mehrotra, A., Singh, A., Kumar, A., Dutt, T., Mishra, B.P., and Pandey, A.K., Comparative signatures of selection analyses identify loci under positive selection in the Murrah Buffalo of India, *Front. Genet.*, 2021, 1915. <https://doi.org/10.3389/fgene.2021.673697>
- Uddin, M., Pellecchia, G., Thiruvahindrapuram, B., D'Abate, L., Merico, D., Chan, A., et al., Indexing effects of copy number variation on genes involved in developmental delay, *Scientific Rep.*, 2016, vol. 6, no. 1, 28663. <https://doi.org/10.1038/srep28663>
- Uemura, M., Tamura, K., Chung, S., Honma, S., Okuyama, A., Nakamura, Y., and Nakagawa, H., Novel 5 α -steroid reductase (SRD5A3, type-3) is overexpressed in hormone-refractory prostate cancer, *Cancer Sci.*, 2008, vol. 99, no. 1, pp. 81–86. <https://doi.org/10.1111/j.1349-7006.2007.00656.x>
- Vargas, G., Schenkel, F.S., Brito, L.F., de Rezende Neves, H.H., Munari, D.P., de Albuquerque, L.G., and Carvalheiro, R., Genomic regions associated with principal components for growth, visual score and reproductive traits in Nellore cattle, *Livestock Science*, 2020, vol. 233, 103936. <https://doi.org/10.1016/j.livsci.2020.103936>
- Vohra, V., Chhotaray, S., Gowane, G., Alex, R., Mukherjee, A., Verma, A., and Deb, S.M., Genome-wide association studies in Indian Buffalo revealed genomic regions for lactation and fertility, *Front. Genet.*, 2021, vol. 12, 696109. <https://doi.org/10.3389/fgene.2021.696109>
- Wang, Q., Moyret-Lalle, C., Couzon, F., Surbiguet-Clippe, C., Saurin, J.C., Lorca, T., et al., Alterations of anaphase-promoting complex genes in human colon cancer cells, *Oncogene*, 2003, vol. 22, no. 10, pp. 1486–1490. <https://doi.org/10.1038/sj.onc.1206224>
- Weigand, H., and Leese, F., Detecting signatures of positive selection in non-model species using genomic data, *Zoologic. J. Linnean Society*, 2018, vol. 184, no. 2, pp. 528–583. <https://doi.org/10.1093/zoolinlean/zly007>
- Wright, S., The genetical structure of populations, *Ann. Eugenics*, 1949, vol. 15, no. 1, pp. 323–354. <https://doi.org/10.1111/j.1469-1809.1949.tb02451.x>
- Wu, X., Fang, M., Liu, L., Wang, S., Liu, J., Ding, X., et al., Genome wide association studies for body conformation traits in the Chinese Holstein cattle population, *BMC genomics*, 2013, vol. 14, pp. 1–10. <https://doi.org/10.1186/1471-2164-14-897>
- Xu, J., Cai, Y., Ma, Z., Jiang, B., Liu, W., Cheng, J., et al., The RNA helicase DDX5 promotes viral infection via regulating N6-methyladenosine levels on the DHX58 and NF κ B transcripts to dampen antiviral innate immunity, *PLoS Pathogens*, 2021, vol. 17, no. 4, e1009530. <https://doi.org/10.1371/journal.ppat.1009530>
- Xu, Z., Wang, X., Song, X., An, Q., Wang, D., Zhang, Z., et al., Association between the copy number variation of CCSE1 gene and growth traits in Chinese Capra hircus (goat) populations, *Anim. Biotechnol.*, 2023, vol. 34, no. 4, pp. 1377–1383. <https://doi.org/10.1080/10495398.2022.2025818>
- Yang, J.I., Li, W.R., Lv, F.H., He, S.G., Tian, S.L., Peng, W.F., et al., Whole-genome sequencing of native sheep provides insights into rapid adaptations to extreme environments, *Mol. Biol. Evolut.*, 2016, vol. 33, no. 10, pp. 2576–2592. <https://doi.org/10.1093/molbev/msw129>
- Yuan, T., Wang, Y., Zhao, Z.J., and Gu, H., Protein-tyrosine phosphatase PTPN9 negatively regulates ErbB2 and epidermal growth factor receptor signaling in breast cancer cells, *J. Biolog. Chem.*, 2010, vol. 285, no. 20, pp. 14861–14870. <https://doi.org/10.1074/jbc.M109.099879>
- Zhang, R., Große-Brinkhaus, C., Heidt, H., Uddin, M.J., Cinar, M.U., Tesfaye, D., et al., Polymorphisms of NELL1 and RNCK1 in relation to porcine growth, carcass and meat quality traits, *J. Anim. Sci.*, 2016, vol. 94, pp. 55–56.
- Zhang, S., Liu, X., Qiu, P., Chen, B., Xu, C., Dong, W., and Liu, T., Microplastics can selectively enrich intracellular and extracellular antibiotic resistant genes and shape different microbial communities in aquatic systems, *Sci. Total Environ.*, 2022, vol. 822, 153488. <https://doi.org/10.1016/j.scitotenv.2022.153488>
- Zhang, X., Lin, Q., Liao, W., Zhang, W., Li, T., Li, J., et al., Identification of New Candidate Genes Related to Semen Traits in Duroc Pigs through Weighted Single-Step GWAS, *Animals*, 2023, vol. 13 no. 3, pp. 365. <https://doi.org/10.3390/ani13030365>
- Zhao, F.Q., Okine, E.K., Cheeseman, C.I., Shirazi-Beechey, S.P., and Kennelly, J.J., Glucose transporter gene expression in lactating bovine gastrointestinal tract, *J. Anim. Sci.*, 1998, vol. 76, no. 11, pp. 2921–2929. <https://doi.org/10.2527/1998.76112921x>
- Zhao, F., McParland, S., Kearney, F., Du, L., and Berry, D.P., Detection of selection signatures in dairy and beef cattle using high-density genomic information, *Genet. Select. Evolut.*, 2015, vol. 47, no. 1, pp. 1–12. <https://doi.org/10.1186/s12711-015-0127-3>
- Zheng, S., Chen, Y., Yu, S., Weng, C., Lin, N., Luo, Z., et al., DISC1 as a prognostic biomarker correlated with immune infiltrates in gastric cancer, *Heliyon*, 2023, vol. 9, no. 4, e15058. <https://doi.org/10.1016/j.heliyon.2023.e15058>

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