

EFFECT OF POLYAMINE METABOLISM INHIBITORS ON LEWIS LUNG CARCINOMA GROWTH AND METASTASIS

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Aim: To study the influence of polyamine metabolism inhibitors on the growth, metastasis and ornithine decarboxylase (ODC) activity of Lewis lung carcinoma. **Materials and Methods:** Experiments were performed on female mice C57Bl/6 with Lewis lung carcinoma. N ω -hydroxy-nor-arginine (nor-NOHA) and α -difluoromethylornithine (DFMO) were used as arginase and ODC inhibitors, correspondently. Inhibition of tumor growth was calculated by comparison of tumor volume in the treated and control groups. The average number of metastases per animal in the group and the average volume of pulmonary metastases per animal in the group have been determined. Determination of ODC — the key enzyme of the polyamine synthesis — in the samples of experimental tumors was performed by method of Luqman S. **Results:** Administration of DFMO or it's combination with nor-NOHA resulted in the decrease of tumor growth rate, number and volume of lung metastases and was accompanied with reduced ODC activity in tumor tissue. **Conclusion:** Modifiers of polyamine metabolism may be considered as promising targeted cancer therapy. **Key Words:** Lewis lung carcinoma, metastases, polyamine metabolism, ornithine decarboxylase, α -difluoromethylornithine, N ω -hydroxy-nor-arginine.

Polyamines (PA) — putrescine, spermidine and spermine — natural polycations, which play a crucial role in many fundamental biological processes: DNA replication, transcription, translation, membrane protein function, protein folding and others [1–6]. The level of PA largely vary during physiological and pathological processes (embryogenesis, ageing [7–9], regeneration, infections, malignant transformation [10, 11]).

The increased level of PA in tumor growth is used as a marker for monitoring in course of a disease and efficiency of therapy [12–14]. Inhibition the level of PA by blocking the activity of the key enzyme for putrescine synthesis ornithine decarboxylase (ODC) — one of the most promising directions in the development of new approaches to the cancer treatment. PA metabolism is regulated at different levels by a complex system of enzymes, which are primarily enzymes of PA synthesis and transformation. The modification of genes regulating PA metabolism by external factors, not studied enough. The aim of this work was to study the effect of inhibitors of PA metabolism on the growth and metastasis of experimental Lewis lung carcinoma in mice.

MATERIALS AND METHODS

Experiments were performed on female C57Bl/6 mice 16.2–23 g. Tumor strain Lewis lung carcinoma was obtained from National Bank of Cell Lines and Transplanted Tumors of R.E. Kavetsky Institute of Experimental Pathology, Oncology and Radiobiology, NAS of Ukraine (Kyiv, Ukraine). All experiments were conducted in accordance to international rules of work with laboratory animals [15]. Lewis lung car-

cinoma transplanted intramuscularly into the shin of the right rear leg $3 \cdot 10^5$ tumor cells in a volume of 0.2 ml of sterile isotonic sodium chloride solution. Animals were kept on a normal diet with free access to water. Each treatment group had 3–6 animals, control group — 4 mice.

Inhibitors applied: arginase inhibitor — N ω -hydroxy-nor-arginine (nor-NOHA) (Cayman Chemical, USA), inhibitor of ODC — α -difluoromethylornithine — DFMO (Centre de Recherche Merrell International, 16 rue d'Ankara, 67084 Strasbourg Cedex, France).

nor-NOHA prepared immediately before use and injected into the abdominal cavity of mice in a dose of 60 mg/kg (1.2 mg per mouse in 0.2 ml of isotonic sodium chloride solution) starting 7 days after tumor transplantation. A comparative analysis was performed *versus* a group of non-treated tumor-bearing mice (control).

In each treatment group, 5 injections of an inhibitor were made. Control animals injected with 0.2 ml of sterile isotonic sodium chloride solution. The volume of tumors registered throughout the experiment. The animals were decapitated 1 day after the last injection of the inhibitors that is 14th day, and on the 26th day after tumor transplantation.

The kinetics of tumor growth evaluated by means of measuring tumor diameters. Inhibition of tumor growth was calculated by comparison of tumor volume in the treated and control groups. To assess the intensity of the process of metastasis was used the following criteria: the average number of pulmonary metastases per animal in the group and the average volume of metastases per animal in the group. In these studies, the number and volume of lung metastases in untreated animals and in animals treated as described above for the tumor growth delay studies were scored on day 26 post-tumor implant. The number and volume of metastases were counted in the transmitted light

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Abbreviations used: DFMO — alpha-difluoromethylornithine; nor-NOHA — N ω -hydroxy-nor-arginine; ODC — ornithine decarboxylase; PA — polyamines.

after removal of the lung, fixing it between the glass plates. Tumor volume was calculated by the formula:

$$V = a^2 \cdot b \cdot \pi / 6,$$

where a — short, b — longer diameter of tumor.

The metastasis inhibition index (MII) was calculated by the formula:

$$MII = ((Ac - Ae) / Ac) \cdot 100,$$

where Ac — the number of metastases to the lung in mice of the control group and Ae — the frequency of metastases in a treated group.

Determination of the ODC activity in the samples of experimental tumors was performed by method S. Luqman [16, 17]. The method is based on the fact that the corresponding enzyme converts L-ornithine hydrochloride (substrate) to the yellow colored putrescine adduct soluble in pentanol, which measured by spectrophotometer.

The statistic processing of obtained results was conducted with the help of Student's t -criterion. The data were reported as the $M \pm m$.

RESULTS AND DISCUSSION

Arginine is a precursor for ornithine which may be then converted into putrescine — the first participant in metabolic transformations of PA [18]. It was found that DFMO — specific inhibitor of ODC — prevents progression of cancer cells in model systems [19]. L-arginine is a nonessential amino acid that plays a central role in several biological systems including the immune response. L-arginine is metabolized by arginase I and arginase II with farther creation of ornithine [20]. In our experiments, the effect of PA synthesis inhibitors — DFMO and a competitive inhibitor of ornithine synthesis nor-NOHA — on the Lewis lung carcinoma growth has been performed. The results are presented in Fig. 1, Table 1.

As can be seen, there is a significant inhibition of tumor growth beginning of the eleventh day of the experiment. During the experiment, both inhibitors studied demonstrate inhibiting activity (within 28–43%).

In this regard, it was important to follow the changes in the dynamics of tumor growth that presented in Fig. 1.

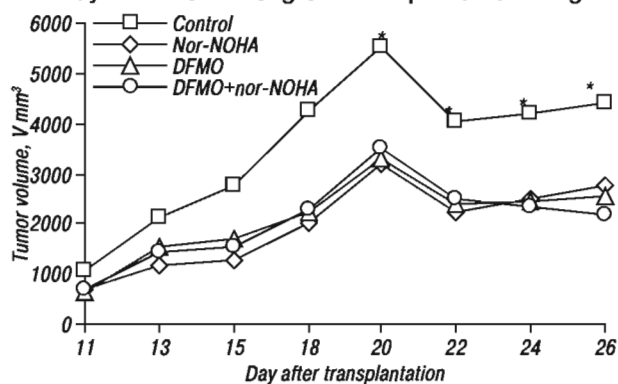


Fig. 1. Lewis lung carcinoma growth in mice under the influence of inhibitors of PA metabolism. *Significant difference ($p < 0.05$) with each experimental group (excluding the DFMO + nor-NOHA group at the 11th day, see Table 1)

Assessing the growth rate of the primary tumor node, it was found that average daily increase of tumor volume in the control group of animals within 9 days

(11–20 days) was $497.2 \pm 56.0 \text{ mm}^3$. The use of both studied inhibitors and their combination led to a significant reduction in the rate of growth, the latter was about 300 mm^3 during the same time period. Significant difference in the impact on growth inhibition when using different inhibitors was not found (Fig. 2).

Table 1. Inhibition of growth of Lewis lung carcinoma in mice under influence of inhibitors of PA metabolism

| Groups | Number of animals | The volume of tumors, mm^3 | | | Inhibition, % | | |
|-----------------|-------------------|-------------------------------------|----------------------|----------------------|----------------------|----------------------|----------------------|
| | | 11 th day | 13 th day | 20 th day | 11 th day | 13 th day | 20 th day |
| Control | 4 | 1067 ± 133 | 2123 ± 165 | 5542 ± 439 | – | – | – |
| nor-NOHA | 5 | 690 ± 129* | 1194 ± 163* | 3163 ± 662* | 35.3 | 43.0 | 43.0 |
| DFMO | 4 | 633 ± 114* | 1534 ± 232* | 3305 ± 461* | 40.7 | 28.0 | 40.0 |
| DFMO + nor-NOHA | 3 | 754 ± 138 | 1439 ± 149* | 3504 ± 281* | 0 | 32.0 | 37.0 |

Note: * $p < 0.05$ as compared to control.

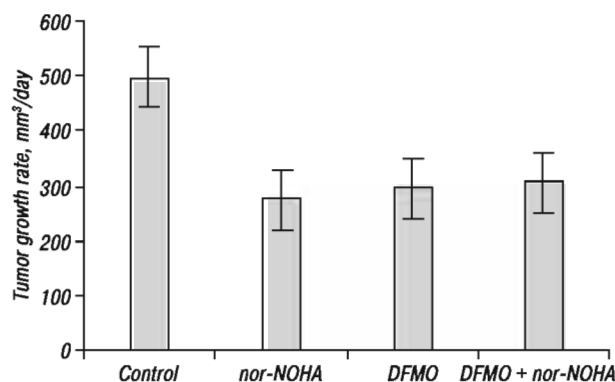


Fig. 2. The rate of tumor growth of Lewis lung carcinoma in mice during 11–20 days compared to control

It is known that transplanted Lewis lung carcinoma refers to metastatic tumors, so anti-metastatic effect, as to this tumor strain, is an important indicator of antitumor effect. The results of the next section of experiments devoted to examining the influence of inhibitors of PA metabolism on the growth of lung metastases (Table 2).

Table 2. Effect of inhibitors of PA metabolism on the number and volume of metastases of Lewis lung carcinoma in mice (26th day)

| Groups | Number of animals | The average number of metastases per 1 animal | | The average volume of metastases per 1 animal | |
|-----------------|-------------------|---|-------|---|-------|
| | | n | MI, % | mm^3 | MI, % |
| Control | 4 | 11.0 ± 2.2 | – | 160.1 ± 44.1 | – |
| nor-NOHA | 5 | 16.6 ± 3.2 | 0 | 138.8 ± 44.2 | 0 |
| DFMO | 4 | 12.0 ± 0.7 | 0 | 9.8 ± 2.3* | 94 |
| DFMO + nor-NOHA | 3 | 11.0 ± 2.3 | 0 | 11.9 ± 5.3* | 93 |

Note: * $p < 0.01$ as compared to control.

As it is seen, the number of metastases in the lungs of all treated mice did not differ versus control. At the same time, volume of the metastatic lesions in the DFMO and DFMO + nor-NOHA groups was near 90% less versus control.

Taking in mind the importance of the ODC enzyme and PA synthesis for cell proliferation, the next section of research was devoted to study of ODC activity in the cells of Lewis lung cancer after the action of inhibitors of PA metabolism. The experimental results are presented in Table 3.

The data obtained show that ODC activity in the tumor increases during the tumor development. On 26th day of experiment, ODC activity in tumors in all treated groups

was decreased *versus* control (see Table 3). It may be expected that inhibition of ODC activity is the main cause of inhibition of tumor growth. Our preclinical data indicate that DFMO in combination with inhibitors of arginase has potential for chemoprevention of cancer and should be evaluated in other models and in combination with other drugs in anticipation of future clinical trials.

Table 3. Effect of inhibitors of PA metabolism on ODC activity in cells of experimental Lewis lung carcinoma

| Groups | Number of animals | 13 th day | | 26 th day | |
|-----------------|-------------------|-------------------------------|---------------|-------------------------------|---------------|
| | | ODC activity, A.U./mg protein | Inhibition, % | ODC activity, A.U./mg protein | Inhibition, % |
| Control | 4 | 0.034 ± 0.002 | – | 0.578 ± 0.070 | – |
| nor-NOHA | 5 | 0.070 ± 0.012* | 0 | 0.346 ± 0.046* | 40 |
| DFMO | 4 | 0.010 ± 0.001* | 0 | 0.355 ± 0.060** | 39 |
| DFMO + nor-NOHA | 3 | 0.019 ± 0.001* | 0 | 0.348 ± 0.050* | 40 |

Note: *0.1 < p < 0.25; *0.025 < p < 0.01; **0.01 < p < 0.005 as compared to control.

REFERENCES

1. Igarashi K, Kashiwagi K. Protein-conjugated acrolein as a biochemical marker of brain infarction. *Mol Nutr Food Res* 2011; **55**: 1332–41.
2. Lopatin AN, Makhina EN, Nichols CG. Potassium channel block by cytoplasmic polyamines as the mechanism of intrinsic rectification. *Nature* 1994; **372**: 366–9.
3. Williams K. Modulation and block of ion channels: a new biology of polyamines. *Cell Signal* 1997; **9**: 1–13.
4. Ha HC, Sirisoma NS, Kuppusamy P, *et al.* The natural polyamine spermine functions directly as a free radical scavenger. *Proc Natl Acad Sci USA* 1998; **95**: 11140–5.
5. Kurata HT, Marton LJ, Nichols CG. The polyamine binding site in inward rectifier K⁺ channels. *J Gen Physiol* 2006; **127**: 467–80.
6. Agostinelli E, Marques MP, Calheiros R, *et al.* Polyamines: fundamental characters in chemistry and biology. *Amino Acids* 2010; **38**: 393–403.
7. Liu P, Gupta N, Jing Y, *et al.* Age-related changes in polyamines in memory-associated brain structures in rats. *Mol Neurosci* 2008; **55**: 789–96.
8. Gupta VK, Scheunemann L, Eisenberg T, *et al.* Restoring polyamines protects from age-induced memory impairment in an autophagy-dependent manner. *Nat Neurosci* 2013; **16**: 1453–60.
9. Minois N, Carmona-Gutierrez D, Madeo F. Polyamines in aging and disease. *Aging* 2011; **3**: 716–32.
10. Casero RA Jr, Marton LJ. Targeting polyamine metabolism and function in cancer and other hyperproliferative diseases. *Nat Rev Drug Discov* 2007; **6**: 373–90.
11. Pegg AE, Casero RA Jr. Current status of the polyamine research field. *Meth Mol Biol* 2011; **720**: 3–35.
12. Park MH, Igarashi K. Polyamines and their metabolites as diagnostic markers of human diseases. *Biomol Ther* 2013; **21**: 1–9.
13. Kawakita M, Hiramatsu K. Diacetylated derivatives of spermine and spermidine as novel promising tumor markers. *J Biochem* 2006; **139**: 315–22.
14. Hiramatsu K, Takahashi K, Yamaguchi T, *et al.* N(1),N(12)-Diacetylspermine as a sensitive and specific novel marker for early- and late-stage colorectal and breast cancers. *Clin Cancer Res* 2005; **11**: 2986–90.
15. Council Directive 2010/63/EU of 22 September 2010 on the protection of animals used for scientific purposes. *Official J Eur Commun* 2010; **L 276**: 33–79.
16. Luqman S, Masood N, Srivastava S, *et al.* Modified spectrophotometric and methodical approach to find novel inhibitors of ornithine decarboxylase enzyme: a path through the maze. *Protocol Exchange* 2013; doi:10.1038/protex.2013.045.
17. Shlyakhovenko VA, Milinevska OA. Determination of ornithine decarboxylase activity. *Lab Diagnostics* 2014; **1**: 36–38 (in Ukrainian).
18. Morris SM Jr. Recent advances in arginine metabolism: roles and regulation of the arginases. *Br J Pharmacol* 2009; **157**: 922–30.
19. Mohammed A, Janakiram NB, Madka V, *et al.* Eflornithine (DFMO) prevents progression of pancreatic cancer by modulating ornithine decarboxylase signaling. *Cancer Prev Res* 2014; **7**: 1198–209.
20. Albina JE, Caldwell MD, Henry WL, *et al.* Regulation of macrophage functions by L-arginine. *J Exp Med* 1989; **169**: 1021–9.