

SUPEROXIDE- AND NO-DEPENDENT MECHANISMS OF ANTITUMOR AND ANTIMETASTATIC EFFECT OF L-ARGININE HYDROCHLORIDE AND COENZYME Q₁₀

A.P. Burlaka^{1,*}, I.I. Ganusevich¹, V.V. Golotiuk², A.V. Vovk¹, S.M. Lukin¹

¹R.E. Kavetsky Institute of Experimental Pathology, Oncology and Radiobiology, NAS of Ukraine,
Kyiv 03022, Ukraine

²Ivano-Frankivsk National Medical University, Ivano-Frankivsk 76018, Ukraine

Aim: To study the redox-dependent mechanism of antiradical, antitumor and antimetastatic action of L-arginine hydrochloride (L-Arg) and coenzyme Q₁₀ (CoQ₁₀) *in vivo*. **Materials and Methods:** The study was performed on C₅₇Bl mice with transplanted Lewis lung carcinoma treated by intraperitoneal injections of L-Arg at low or high doses (60 and 360 mg/kg body weight), CoQ₁₀ (0.2 and 1.2 mg/kg body weight) or their combinations. Electron paramagnetic resonance was applied for analysis of mitochondrial electron transport chain, CoQ₁₀ levels, free iron (FI), the level of NO, and the rate of superoxide radical generation. The activity of matrix metalloproteinase (MMP)-2 and -9 in tumor tissue was determined by zymography method in polyacrylamide gel. **Results:** Administration of L-Arg at high doses caused an inhibition of tumor growth by 48 ± 8.0%, increase of superoxide radical generation rate and NO levels to a value of 1.23 ± 0.14 and 2.26 ± 0.31 nm/g tissue · min, and decreased activity of MMP-2 and -9 (3.55 ± 0.8 and 4.8 ± 1.0 r.u., respectively). Treatment with L-Arg at low doses stimulated tumor growth and increased the levels of MMP-2 and -9 activities (8.44 ± 2.7 and 9.8 ± 3.1 r.u., respectively). Administration of CoQ₁₀ at high doses significantly decreased superoxide radical generation rate to the values of 0.44 ± 0.09 nm/g tissue · min, levels of free iron and NO, and caused tumor growth inhibition by 54 ± 11.3%. The combined use of L-Arg and CoQ₁₀ at high doses caused tumor growth inhibition by 51 ± 7.4% compared to Lewis lung carcinoma-bearing untreated animals (p < 0.05). **Conclusions:** Administration of L-Arg and CoQ₁₀ caused the dose-dependent effect on the rate of generation of superoxide radicals, level of ubiquinone, complexes NOFeS-proteins, levels of FI and NO. L-Arg at low doses positively modulated MMP-9 activity that promoted tumor progression. Upon combined use of L-Arg and CoQ₁₀, superoxide radicals and NO form the redox state that causes decrease of MMP-2, -9 activities with consequent inhibition of tumor invasion and metastasis.

Key Words: L-arginine, coenzyme Q₁₀, superoxide radical, nitric oxide, matrix metalloproteinase, Lewis lung carcinoma.

One of the approaches to search for innovative anti-cancer agents with fundamentally different mechanism of action is the use of compounds — donors of nitric oxide (NO), which can modulate the levels of NO and other regulators of transport rate of electrons in the electron transport chain in mitochondria. Normally functioning mitochondrial metabolism is a process of fine-adjustable dynamic balance of thousands of anabolic and catabolic reactions and cell signaling systems. Since NO is the key signaling molecule in tumor-induced angiogenesis, its antitumor activity can be manifested at least due to its ability to regulate angiogenic pathway [1–5].

Coenzyme Q₁₀ (CoQ₁₀) plays the key role in the mechanism of functioning of the power generating cell system and in the formation of redox potential in mitochondria through the regulation of superoxide radical generation to control gene transcription and cellular and extracellular signaling factors. Physiological levels of CoQ₁₀ in plasma are 0.68–1.1 μmol/l and are supported mainly by endogenous synthesis and to a lesser extent by products of exogenous origin. Activity of CoQ₁₀ decreases with aging as well as in various pathologies, including diabetes, chronic heart failure,

myocardial infarction and cancer [6]. There is a growing amount of evidence in favor of the use of food additives with CoQ₁₀ for prevention and treatment of pathological conditions, NO donors and other regulators of electron transport in mitochondria, including CoQ₁₀ are explored as anticancer agents [7, 8]. However, mechanisms of their action have been studied insufficiently. The matrix metalloproteinases (MMPs), which play a role in tumor invasion and metastasis [9, 10], are redox-dependent enzymes [11–13], therefore, they may be involved in the realization of anti-tumor effects of compounds-regulators of electron transport [14, 15].

The aim of the study was to investigate the redox-dependent mechanism of anti-radical, antitumor and antimetastatic action of L-Arg and CoQ₁₀ *in vivo*.

MATERIALS AND METHODS

The study was performed on 84 C₅₇Bl male mice weighting 22.4 ± 1.12 g bred in animal facility of R.E. Kavetsky Institute of Experimental Pathology, Oncology and Radiobiology of the NAS of Ukraine. The animals were kept on a standard diet. Animal study protocols and operation procedures were carried out in accordance with the main requirements to keeping and working with laboratory animals and to the rules of local Bioethics Committee.

Lewis lung carcinoma (LLC) was used as a tumor model. LLC cells (5 · 10⁵ cells per animal) were transplanted subcutaneously [16]. The compounds (L-Arginine hydrochloride (L-Arg) (Sigma, USA),

Submitted: February 16, 2016.

*Correspondence: E-mail: apburlaka@gmail.com

Abbreviations used: CoQ₁₀ – coenzyme Q₁₀; EPR – electron paramagnetic resonance; FI – free iron; L-Arg – L-arginine hydrochloride; LLC – Lewis lung carcinoma; MMP – matrix metalloproteinase; NO – nitric oxide; O₂⁻ – superoxide radical.

CoQ₁₀ (Sigma, USA) were administered intraperitoneally daily for 10 days at low or high doses: L-Arg — 60 and 360 mg/kg body weight; CoQ₁₀ — 0.2 and 1.2 mg/kg body weight. The animals were distributed into 8 experimental groups; I — intact animals (n = 10); II — tumor control (n = 11); III and IV — animals with tumors treated with high (n = 12) and low (n = 10) doses of L-Arg, respectively; V and VI — animals with tumors treated with high (n = 10) and low (n = 10) doses of CoQ₁₀, respectively; VII and VIII — animals with tumors treated with high (n = 10) and low (n = 10) doses of L-Arg and CoQ₁₀, respectively. Animals of group II received daily intraperitoneal injections of physiological saline at corresponding volume for 10 days and served as a positive control.

The research of the electron transport chain in mitochondria, CoQ₁₀ levels in mitochondria and free iron (FI) in tumor cells was performed by electron paramagnetic resonance (EPR) method with computerized spectrometer RE-1307 using the technology of low-temperature stabilization of biological material (77 K). The rate of superoxide radical generation by mitochondria of cells was determined by EPR method using a spin trap (2,2,6,6-tetramethyl-4-piperidine) at the room temperature in a special paramagnetic pure quartz cuvette. The level of NO in the tumor tissue was investigated by EPR using the Spin Traps technology (spin trap — diethyl dithiocarbamates (Sigma)) at the temperature of 77 K [13].

The activity of MMP-2 and -9 in tumor tissue was determined by zymography method in polyacrylamide gel (with addition of gelatin as a substrate) based on SDS-electrophoresis of proteins [17].

The tumor volume was determined by the formula 1:

$$V = (\pi/6) \cdot D1 \cdot D2 \cdot D3, (1)$$

where V — tumor volume (mm³); D1, D2, D3 — tumor length, width, and height.

The number of metastases was counted, and their volume — by the formula 1.

The antitumor activity of the studied compounds was evaluated by tumor growth inhibition:

$$G_{\%} = (v_c - v_e) / m_k \cdot 100\%, (2)$$

where G_% — percentage of tumor growth inhibition by volume; v_c — average tumor volume in the control; v_e — average tumor volume in the study group.

Statistical analysis of the obtained data was performed with the use of Origin 7.0 program and Student's *t*-criterion. The data were presented as the mean with the standard deviation (M ± SD). The differences were considered significant at *p* < 0.05.

RESULTS AND DISCUSSION

The results of EPR study of LLC are shown in Fig. 1.

In tumor cells of group II (EPR spectrum 1) there were registered the low levels of free radical form of CoQ₁₀ — ubisemiquinone (*g* = 2.00) — 0.15 ± 0.02 r.u., Fe-S-protein N-2 (*g* = 1.94) responsible for coupling of oxidation and phosphorylation — 0.14 ± 0.01 r.u. Also, there was noted an accumulation of high levels of FI (*g* = 2.20–2.40) — 0.86 ± 0.08 r.u., the level of NOFeS-protein complexes was 0.18 ± 0.02 r.u.

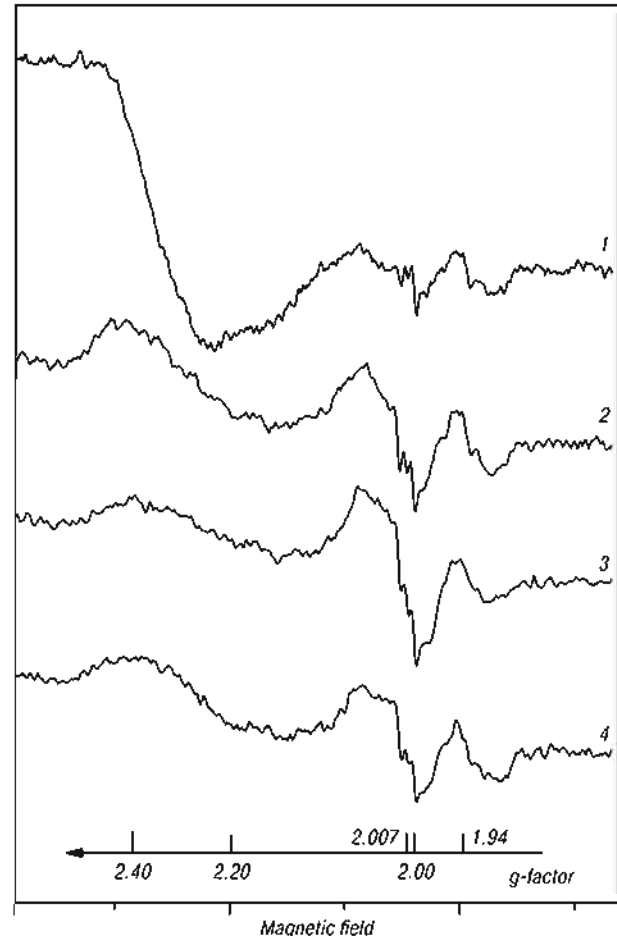


Fig. 1. Effect of L-Arg and CoQ₁₀ at high doses on mitochondrial electron transport chain in LLC cells: 1 — tumor of mice in group II; 2 — tumor of mice in group III; 3 — tumor of mice in group V; 4 — tumor of mice in group VII

In animals injected with L-Arg at high doses, nanomolar levels of NO in the tumor tissue were generated, what caused 2.3-fold decrease of FI level compared to that in group II (*p* < 0.05). Under these conditions, there were recorded significant increase of the levels of ubisemiquinone up to 0.21 ± 0.02 r.u., and NOFeS-proteins complexes in the mitochondrial electron transport chain (up to 0.48 ± 0.02 r.u.), while activity of FeS-protein N-2 remained unaltered (0.14 ± 0.013 r.u.).

Administration of CoQ₁₀ resulted in the decreased FI content in tumor tissue of animals (0.21 ± 0.02 r.u.) compared to groups II and III (*p* < 0.05), and increased formation of NO-FeS-proteins complex (0.58 ± 0.04 r.u.). The level of ubisemiquinone in the I complex of mitochondrial respiratory chain was 0.22 ± 0.01 r.u. what was significantly lower compared with group II; *p* < 0.05; activity of FeS-protein N-2 was 0.11 ± 0.02 r.u. CoQ₁₀, providing mitochondria with ubiquinone (NADH-ubiquinone-oxidoreductase and succinate dehydrogenase complexes), promotes the restoration of oxidative phosphorylation via its elevation in the electron transport chain.

The combined therapy with L-Arg and CoQ₁₀ resulted in the decrease of FI level — 0.28 ± 0.03 r.u. compared to group II (*p* < 0.05), ubisemiquinone (0.15 ± 0.01 r.u.) and NOFeS-proteins complexes (0.38 ± 0.04 r.u.)

compared to groups III and IV ($p < 0.05$), increase of FeS-protein N-2 activity — 0.14 ± 0.02 (compared to group IV; $p < 0.05$).

Fig. 2 shows the data on the rate of generation of superoxide radicals by LLC cells. In the tumors of group II the value was 0.75 ± 0.13 nm/g tissue · min, while in the lungs of intact animals it was equal to 0.19 ± 0.03 nm/g tissue · min. In the case of administration of high doses of L-Arg, the rate of superoxide radical generation was 1.23 ± 0.14 nm/g tissue · min, that was significantly higher than corresponding values of group II ($p < 0.05$). Therapy with L-Arg at low doses did not influence on this index (0.78 ± 0.22 nm/g tissue · min). The use of high doses of CoQ₁₀ caused nearly 2-fold decrease in levels of superoxide radicals (0.44 ± 0.09 nm/g tissue · min) in mitochondria of tumor cells compared to group II ($p < 0.05$). After administration of low CoQ₁₀ doses this index did not change (0.62 ± 0.08 nm/g tissue · min). Combined administration of L-Arg and CoQ₁₀ at low doses reduced the superoxide radical generation rate in mitochondria of tumor cells (0.32 ± 0.04 nm/g tissue · min) compared with groups II, III, IV, VI ($p < 0.05$), whereas high doses of this compounds reduced this index to 0.65 ± 0.09 nm/g tissue · min what was lower than in group III ($p < 0.05$). Thus, the regulators of the electron transport rate in mitochondrial respiratory chain decreased the rate of superoxide radical generation to corresponding levels in a dose dependent manner.

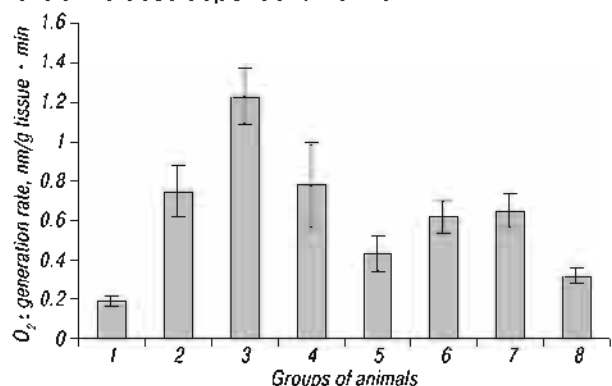


Fig. 2. The rate of superoxide generation (O_2) in the mitochondria of cells: 1 — lungs of intact mice; 2 — tumor control; 3 — tumors of mice that administered high doses of L-Arg; 4 — low doses of L-Arg; 5 — tumors of mice treated with high doses of CoQ₁₀; 6 — low doses of CoQ₁₀; 7 — tumors of mice treated with high doses of L-Arg and CoQ₁₀ and 8 — L-Arg and CoQ₁₀ at low doses. $p < 0.05$ compared to tumor control (group II)

Fig. 3 shows the results of the research of NO content in tumor cells. In the tumor bearing animals this rate was 1.95 ± 0.21 nm/g tissue · min, which exceeded the rate in the group of intact animals — 1.45 ± 0.18 nm/g tissue · min ($p < 0.05$). NO level in the tumor after administration of high doses of L-Arg increased up to 2.26 ± 0.31 nm/g tissue · min versus 1.58 ± 0.19 nm/g tissue · min in the case of low dose L-Arg treatment, and the difference between these indexes is statistically significant. The levels of NO in the cases of administration of high and low doses of CoQ₁₀ were 1.51 ± 0.12 and 1.62 ± 0.23 nm/g tissue · min, respectively. Combined therapy with L-Arg and CoQ₁₀ at high

doses resulted in the increased level of NO up to the values of 2.2 ± 0.3 nm/g tissue · min compared with groups IV, V, VI ($p < 0.05$), whereas in the case of low dose combined treatment this index (1.47 ± 0.09 nm/g tissue · min) was significantly lower than in groups II, III and VII ($p < 0.05$). Therefore, the NO donor and its combined use with CoQ₁₀ at high doses cause the significant increase of NO levels in tumor cells, which is a positive factor in the antitumor and antimetastatic therapy.

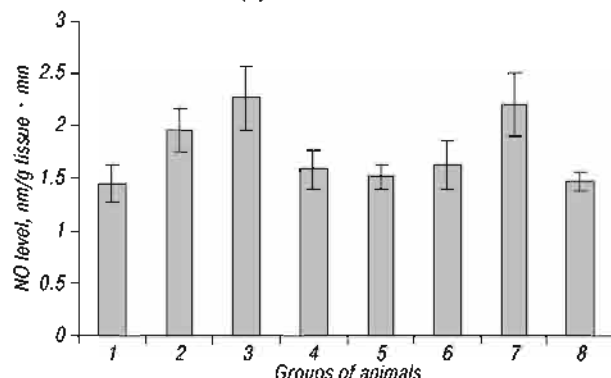


Fig. 3. The level of NO in the mitochondria of cells: 1 — lungs of intact mice; 2 — tumor control; 3 — tumors of mice treated with high doses of L-Arg; 4 — low doses of L-Arg; 5 — tumors of mice treated with high doses of CoQ₁₀; 6 — low doses of CoQ₁₀; 7 — tumors of mice treated with high doses of L-Arg and CoQ₁₀; 8 — L-Arg and CoQ₁₀ at low doses. $p < 0.05$ compared with tumor control (group II)

Administration of L-Arg at high doses causes an increase in NO level, augments mitochondrial damage, intensifies generation of superoxide radicals, increases the rate of oxidation-induced mutations of mtDNA, and induces tumor cell apoptosis [8]. NO at low doses promotes tumor invasiveness, angiogenesis and immune tolerance [1, 7]. Administration of CoQ₁₀ at high doses to animals with LLC significantly reduced NO levels in tumor cells compared to control animals (see Fig. 3). Combined use of L-Arg and CoQ₁₀ at low doses caused an increase of NO levels in tumor cells compared to group II ($p < 0.05$).

Summarizing the obtained information, one may suppose that high doses of regulators of the electron transport rate based on NO and CoQ₁₀ donors cause correlating effects at the level of mitochondria. These compounds influence regulation of superoxide radical generation rate and NO level, and subsequently modulate MMP-2 and -9 activities. Activity of MMP-2 and -9 in tumor tissue of the control animals greatly exceeded these indices in the lungs of the intact animals and correlated with the values of the superoxide radicals generation rate by tumor cells in these groups of animals (Fig. 2, 4, 5). In tumors of animals injected with high doses of L-Arg a significant (almost 2 times; $p < 0.05$) decrease in activity of both gelatinases was observed compared with the control group, which could be a consequence of the ultrahigh rate of superoxide radical generation. On the contrary, low doses of L-Arg caused an insignificant increase in activity of MMP-2 in tumor tissue compared to the control that was due to the regulatory impact of the increase

in rate of superoxide radical generation in tumor cells of this group of animals (see Fig. 2, 4, 5). Treatment with both high and low doses of CoQ₁₀ led to the significant superoxide regulated decrease (1.7–2.3 times; $p < 0.05$) in activity of both gelatinases, compared to group II (see Fig. 2, 4, 5).

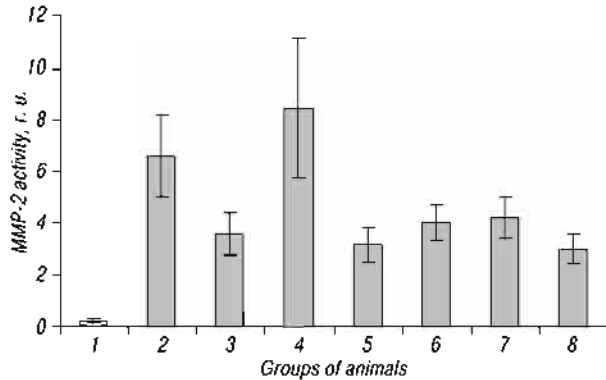


Fig. 4. Activity of MMP-2 (r.u.): 1 — lungs of intact mice; 2 — tumor control; 3 — tumors of mice treated with high doses of L-Arg; 4 — low doses of L-Arg; 5 — tumors of mice treated with high doses of CoQ₁₀; 6 — low doses of CoQ₁₀; 7 — tumors of mice treated with high doses of L-Arg and CoQ₁₀; 8 — L-Arg and CoQ₁₀ at low doses. $p < 0.05$ compared to tumor control (group II)

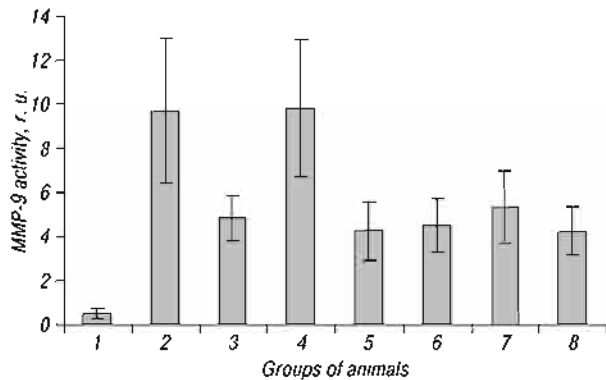


Fig. 5. Activity of MMP-9 (r.u.): 1 — lungs of intact mice; 2 — tumor control; 3 — tumors of mice treated with high doses of L-Arg; 4 — low doses of L-Arg; 5 — tumors of mice treated with high doses of CoQ₁₀; 6 — low doses of CoQ₁₀; 7 — tumors of mice treated with high doses of L-Arg and CoQ₁₀; 8 — L-Arg and CoQ₁₀ at low doses. $p < 0.05$ compared to tumor control (group II)

Activity of MMP-2 and -9 in tumors of animals injected by both substances at high or low doses was significantly below the control values ($p < 0.05$). In this case (Table 1, 2) low gelatinase activity levels corresponded to slower primary tumor growth and decreased metastasis in experimental animals. In the control group and group of animals injected with low doses of L-Arg, high gelatinase activity levels were observed in tumor tissue along with accelerated LLC growth and metastasis.

Tables 1 and 2 shows the data on antitumor and antimetastatic activity of L-Arg and CoQ₁₀ administered at low and high doses. As one may see, the antitumor and antimetastatic activity of L-Arg differed significantly depending on the level of NO in tumor cells ($p < 0.05$). Thus, high doses of the compound caused a decrease in the tumor volume by $48.0 \pm 8.0\%$, while low doses stimulated its growth, number of metastases and their volume ($p < 0.05$). Treatment with CoQ₁₀ at high and low

doses resulted in inhibition of the tumor growth by 54.0 ± 11.3 and $39.0 \pm 5.4\%$, respectively, the difference between these indexes was insignificant. Combined effect of both compounds at high doses caused tumor growth inhibition by $51.0 \pm 7.4\%$. Thus, a NO donor and CoQ₁₀ showed the antitumor and antimetastatic activity, and the pro- and anti-tumor effect of NO depended on its dose. These data were in accordance with those of other authors who demonstrated the dual effect of NO donors [1, 7, 8]. Superoxide regulated MMP-2 and -9 mediate anti-tumor effects of L-Arg and CoQ₁₀ at the level of intercellular matrix proteolysis, because reduced activity of gelatinases contributes to inhibition of migration, dissemination of tumor cells and formation of metastatic centers.

Table 1. Antitumor activity of L-Arg and CoQ₁₀

Groups of animals	Doses			
	High		Low	
	Tumor volume, mm ³	Inhibition of tumor growth, %	Tumor volume, mm ³	Inhibition of tumor growth, %
Tumor control	1600 ± 95	—	1600 ± 95	—
L-Arg	832 ± 115 ¹	48 ± 8.0	1840 ± 125 ¹	15 ± 2.6
CoQ ₁₀	736 ± 137 ¹	54 ± 11.3	970 ± 112 ^{1,2}	39 ± 5.4
L-Arg and CoQ ₁₀	784 ± 104 ¹	51 ± 7.4	1410 ± 99 ^{2,3}	12 ± 3.1

Note: ¹ $p < 0.05$ compared to tumor control (group II); ² $p < 0.05$ compared to animals treated with L-Arg; ³ $p < 0.05$ compared to animals treated with CoQ₁₀.

Table 2. Antimetastatic activity of L-Arg and CoQ₁₀

Groups of animals	Doses			
	High		Low	
	Number of metastases	Volume of metastases, mm ³	Number of metastases	Volume of metastases, mm ³
Tumor control	6.8 ± 3.1	139 ± 54	6.8 ± 3.1	139 ± 54
L-Arg	3.6 ± 1.1	79 ± 21	10.8 ± 1.1	212 ± 33
CoQ ₁₀	3.1 ± 2.3	76 ± 13	4.7 ± 2.3 ¹	89 ± 13 ¹
L-Arg and CoQ ₁₀	3.2 ± 1.5	78 ± 18	4.0 ± 2.5 ^{1,2}	99 ± 18 ¹

Note: ¹ $p < 0.05$ compared to animals treated with L-Arg; ² $p < 0.05$ compared to animals treated with CoQ₁₀.

Reprogramming of mitochondria metabolism, disorders of redox homeostasis in mitochondria electron transport chain are characteristic for malignant tumor cells. That is accompanied by the replacement of four- to single-electron restoration of oxygen molecule with subsequent formation of superoxide radicals. Increased production of superoxide radicals in tumor cells promotes cancer progression through the activation of signaling pathways that regulate metabolic changes, proliferation, angiogenesis and metastasis [18, 19].

CoQ₁₀ plays the role of electron transporter in complexes I, II and III electron transport chain of mitochondria. Reduced form of CoQ₁₀ protects the membrane, proteins and mitochondrial DNA from oxidative agents, including superoxide radicals. We have found that CoQ₁₀ realizes its protective function restoring the electron transport process and reducing the level of generation of superoxide radicals by mitochondria in LLC cells. These data are in agreement with the data on protective effect of CoQ₁₀ toward UV irradiated astrocytes [20] and fibroblasts [21].

Recent studies have shown that NO takes part in the regulation of tumor cell proliferation, in particular, in its inhibition and induction of apoptosis, which is possible due

to the ability of NO to inactivate iron-containing enzyme involved in the synthesis of ATP and DNA replication. Activation of NO synthase (NOS) and increased levels of NO generation can have both antitumor effect, and promote the initiation and progression of tumor [2]. The use of different doses of NO may allow to regulate phenotypic responses through molecular mechanisms that form pro- and antitumor effects.

Our results showed a dose-dependent action of L-Arg on redox-dependent processes in LLC. Thus, the use of L-Arg at high doses leads to increased levels of NO and superoxide radicals in mitochondria of LLC cells, increasing their cytotoxic effect by formation of peroxynitrite, which can cause DNA damage and initiation of apoptosis. Similar results were obtained in the study of dose-depending action of NO donors on proliferation and apoptosis in Ehrlich ascites carcinoma [22]. It should be noted that the use of NO donors at low doses enhances the functional activity of mitochondria, stimulates the formation of superoxide radicals in physiological range and, conversely, at high doses — depresses mitochondrial respiration, inhibits the incorporation of arachidonic acid and activates its output.

In modern studies of suppression, stabilization or enhancement of the degradation of extracellular matrix are considered critical characteristics of malignant progression [9] as well as expression level of gelatinases is thought to be associated with the metastasis and overall survival of cancer patients [10]. Known synthetic inhibitors inhibit MMP activity in *in vitro* experiments and in animal models, but failed in clinical trials [23]. On the other hand, some studies have shown that antioxidants [15, 24], including CoQ₁₀ [25, 26], suppress tumor growth by inhibition of MMP activity via intracellular regulation of reactive oxygen species. Our results showed that MMP-2 and -9 mediate antitumor effects of L-Arg and CoQ₁₀ at the level of intercellular matrix proteolysis. Redox-regulated decrease of gelatinases activity promotes inhibition of tumor cell migration, dissemination and formation of metastatic niches.

REFERENCES

- Mocellin S, Bronte V, Nitti D. Nitric oxide, a double edged sword in cancer biology: searching for therapeutic opportunities. *Med Res Rev* 2007; 27: 317–52.
- Ridnour L, Thomas D, Switzer Ch, *et al.* Molecular mechanisms for discrete nitric oxide levels in cancer. *Nitric Oxide* 2008; 19: 73–6.
- Burlaka AP, Sidorik EP. Radical forms of oxygen and nitrogen oxide in the tumor process. Kyiv: Naukova dumka, 2006. 227 p. (in Ukrainian).
- Burlaka AP, Sidorik EP. Redox-dependent signal molecules in mechanisms of tumor process. Kyiv: Naukova dumka, 2014. 255 p. (in Ukrainian).
- Jadeski LC, Lala PK. Nitric oxide synthase inhibition by N(G)-nitro-L-arginine methyl ester inhibits tumor-induced angiogenesis in mammary tumors. *Am J Pathol* 1999; 155: 1381–90.
- Garrido-Maraver Ju, Cordero M, Oropesa-Avila M, *et al.* Coenzyme Q₁₀ Therapy. *Mol Syndromol* 2014; 5: 187–97.
- Cheng H, Wang L, Mollica M, *et al.* Nitric oxide in cancer metastasis. *Cancer Lett* 2014; 353: 1–7.
- Rapozzi V, Pietra E, Zorzet S, *et al.* Nitric oxide-mediated activity in anti-cancer photodynamic therapy. *Nitric Oxide* 2013; 30: 26–35.
- Fingleton B. Matrix metalloproteinases: roles in cancer and metastasis. *Front Biosci* 2006; 11: 479–91.
- Ganusevich II. Role of matrix metalloproteinases (MMP) in malignancies. I. MMP characteristics, regulation, and prognostic value. *Oncology* 2010; 12: 10–6 (in Ukrainian).
- McCormick ML, Gavrilu D, Weintraub NL. Role of oxidative stress in the pathogenesis of abdominal aortic aneurysms. *Arterioscler Thromb Vasc Biol* 2007; 27: 461–9.
- Castro MM, Rizzi E, Rodrigues GJ, *et al.* Antioxidant treatment reduces matrix metalloproteinase-2-induced vascular changes in renovascular hypertension. *Free Radic Biol Med* 2009; 46: 1298–307.
- Burlaka AP, Sidorik EP, Ganusevich II, *et al.* Effects of radical oxygen species and NO: formation of intracellular hypoxia and activation of matrix metalloproteinases in tumor tissues. *Exp Oncol* 2006; 28: 49–53.
- Panyathap A, Chewonarin T, Taneyhill K, *et al.* Antioxidant and anti-matrix metalloproteinases activities of dried longan (*Euphoria longana*) seed extract. *Science Asia* 2013; 39: 12–8.
- Lee SJ, Kim MM. Resveratrol with antioxidant activity inhibits matrix metalloproteinase via modulation of SIRT1 in human fibrosarcoma cells. *Life Sci* 2011; 88: 465–72.
- Sophina Z, Goldin A. Experimental evaluation of anticancer drugs in USSR and USA. Moscow: Medicina, 1980. 295 p. (in Russian).
- DeClerk YA, Perez N, Shimada H, *et al.* Inhibition of invasion and metastasis in cells transfected with an inhibitor of metalloproteinases. *Cancer Res* 1992; 52: 701–8.
- Sabharwal SS, Schumacker PT. Mitochondrial ROS in cancer: initiators, amplifiers or an Achilles' heel? *Nat Rev Cancer* 2014; 14: 709–21.
- LB, Chandel NS. Mitochondrial reactive oxygen species and cancer. *Cancer Metab* 2014; 2: 17.
- Jing L, He MT, Chang Y, *et al.* Coenzyme Q₁₀ protects astrocytes from ROS-induced damage through inhibition of mitochondria-mediated cell death pathway. *Int J Biol Sci* 2015; 11: 59–66.
- Cornelius N, Byron C, Hargreaves I, *et al.* Secondary coenzyme Q₁₀ deficiency and oxidative stress in cultured fibroblasts from patients with riboflavin responsive multiple Acyl-CoA dehydrogenation deficiency. *Hum Mol Genet* 2013; 22: 3819–27.
- Deriagina VP, Ryzhova NI, Krivosheeva LV, *et al.* Production of nitric oxide metabolites during transplanted tumors growth with different metastatic potential. *Biomed Khim* 2014; 60: 661–7 (in Russian).
- Ganusevich II. Role of matrix metalloproteinases (MMPs) in malignancies. II. Participation of MMPs in angiogenesis, invasion and metastasis of tumors. *Oncology* 2010; 12: 108–17 (in Ukrainian).
- Park I-H, Kwon E-J, Lee S-J, *et al.* Quercetin with Antioxidant Activity Inhibits Matrix Metalloproteinase-2 and -9 in HT1080 Cell Line. *Cancer Prev Res* 2011; 16: 223–30.
- Babar M, Khaghani S, Pasalar P, *et al.* Exogenous coenzyme Q₁₀ modulates MMP-2 activity in MCF-7 cell line as a breast cancer cellular model. *Nutr J* 2010; 9: 62.
- Ioui M, Ooe M, Fujii K, *et al.* Mechanisms of inhibitory effects of CoQ₁₀ on UVB-induced wrinkle formation *in vitro* and *in vivo*. *BioFactors* 2008; 32: 237–43.