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RESEARCH ARTICLE

Diagnostic significance of the synthesis of phenolic compounds and proline in the leaves of *Schisandra chinensis* and *Actinidia arguta* for the indication of the stress levels of plants under conditions of mixed plantings

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Abstract

The peculiarities of the accumulation of polyphenolic compounds and free proline were investigated in the leaves of *Actinidia arguta* and *Schisandra chinensis* during their cultivation in vegetation containers with different ratios of the number of plants, namely 1 : 1, 2 : 1, 1 : 2. Monocultural (single-species) planting was used as a control. The content of free proline in plant leaves was carried out according to the method, which is based on the interaction of proline with a ninhydrin reagent, forming a pink-red color. The amount of polyphenolic compounds was determined by the Folin-Ciocalteu method. It was found that the ratio of plants grown together significantly affects the accumulation of primary and secondary metabolites in their leaves. Under conditions of mixed planting, more proline and phenolic substances are accumulated in plant leaves compared to monoculture. The maximum proline content, 19.44 ± 0.91 mg/g of dry weight (DW), was observed in the leaves of *A. arguta* in the experiment combination with a prevailing number of schisandra plants at a ratio of *S. chinensis* and *A. arguta* plants of 2:1. In the same combination, the leaves of actinidia contained the highest amount of phenolic compounds (36.87 ± 2.22 mg/g DW). The studied root exudates of the experimental plants had an allelopathic inhibitory effect on the test culture. The exudates caused 12.0 % average decrease in root growth of *A. arguta* test objects, and 30.0 % average decrease in root growth of *S. chinensis* test objects compared with the control. This allows us to conclude about the high activity of schisandra's allelochemicals, which negatively affect the development of actinidia plants. The optimal ratio of plants *A. arguta* and *S. chinensis* when grown together is 2 : 1, since a higher concentration of schisandra plants in a container more stress in actinidia plants, expressed as an increased accumulation of phenols and prolines in its leaves. The result of a comparative analysis of the amount of proline and phenolic compounds can be used to assess the mutual influence of plants in mixed plantings to optimize their growing conditions, which confirms the diagnostic significance of these metabolites for indicating the stress state of the studied plants.

Keywords: *Actinidia arguta*, *Schisandra chinensis*, phenolic compounds, proline, monocultures, mixed plantings, allelopathic activity

Authors' contributions: T. Venediktova was engaged in preparing and conducting the biochemical analyses, wrote the methodological part of the research, realized statistical processing of the experimental data, wrote the manuscript, and formulated conclusions. N. Zaimenko developed the research concept, interpreted the results. N. Skrypchenko wrote the manuscript, interpreted the results, and formulated conclusions.

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Introduction

One of the directions for intensifying horticulture is the creation of highly productive plantations with greater technological and economic efficiency (Egorov, 2013). The main goal of intensive horticulture is to increase the productivity and quality of agricultural products and reduce material costs for agricultural production. A characteristic feature of a garden, as an agricultural phytocenosis, is its instability associated with the disruption of the trophic relationships of its components due to the intensive use of natural resources and technogenic factors (Popova, 2005). The garden's biotope is subjected to a long-term unilateral impact due to the use of monoculture and typical agricultural techniques (Kudasov, 1999). Gardens are monocultural (i.e., single-species) plantings, with decreased resistance and productivity of phytocoenoses. Long-term permanent cultivation of agricultural plants leads to a progressive deterioration of soil fertility and soil exhaustion. According to the Food and Agriculture Organization (FAO), soil exhaustion is responsible for the loss of 25% of the world crop yield (Zhuchenko, 2008). Numerous studies indicate that the symptoms of soil exhaustion are more pronounced in intensive gardens than in organic ones (Manici et al., 2003).

Diversifying the species composition of fruit plantations by increasing the biological diversity of garden phytocoenoses is possible due to the introduction of rare fruit plants into the culture, in particular the hardy kiwi (*Actinidia arguta* (Siebold et. Zucc.) Planch. ex Miq.) and Chinese magnolia vine (*Schisandra chinensis* (Turcz.) Baill.) – valuable fruit, medicinal and ornamental plants. Fruits of *A. arguta* accumulate a high content of biologically active substances such as vitamins C, E, K, polysaccharides, polyphenols, triterpenoids, and alkaloids, which have analgesic, antibacterial, antioxidant, antitumor, hypoglycemic, and other pharmacological effects (Sun et al., 2020). Fruits of *S. chinensis* is a source of biologically active compounds – vitamins C, E, and P, saponins, flavonoids, organic acids, lignans, pectins, and aromatic substances, that have adaptogenic, tonic, immunostimulatory, anti-inflammatory,

regenerative, antitumor, and other effects (Szopa et al., 2016; Nowak et al., 2019). These plants are widely cultivated in industrial and farm horticulture of many countries. Significant introductory studies of *A. arguta* and *S. chinensis* were carried out at the M.M. Gryshko National Botanical Garden (NBG) of the National Academy of Sciences of Ukraine (Skrypchenko & Latocha, 2017). However, the development of scientific principles for their introduction into horticulture and the perfection of cultivation technology remains crucial.

According to the concept of green horticulture, the main principle for creating garden phytocoenoses should be to optimize their structure by creating multicomponent mixed plantings, that is, by moving from monoculture to polyculture (Moroz, 1990). Interest in “permanent agriculture” reflects growing attention to mixed plantings that are inherently more resilient than monocultures. They can protect and enrich soil ecosystems, allow plants to form mutually beneficial combinations, and create a favorable microclimate (Millner, 2016). With this in mind, studies of *A. arguta* and *S. chinensis* were realized to assess the possibility of co-growth of these fruit vines in mixed plantings, as these plants are characterized by the same life form and similar agronomic cultivation requirements.

Since fruit plants not only absorb the essential mineral elements and organic compounds but also release various metabolites into the environment, the success of the integration of introduced species into the agrocoenoses depends on their allelopathic potential and living compatibility in mixed plantings (Osipova, 1997).

Physiological adaptation of plants to specific conditions is achieved through physiological and biochemical mechanisms. Elucidation of these mechanisms of plant adaptation to changing environmental conditions is of great theoretical and practical importance. Nowadays, it is one of the essential tasks of ecological plant physiology (Fedulov et al., 2015). The presence and accumulation of secondary metabolites reflect the adaptation strategy of plants created by natural selection in the course of evolution.

The effect of any abiotic or biotic stress factor on the plant organism provokes

overproduction of reactive oxygen species (ROS) and upsets the balance between the level of ROS and the activity of the antioxidant defense system (Mittler, 2002; Foyer & Noctor, 2000). An indicator of stress is the amount of proline, the content of which increases tens and hundreds of times under conditions of drought, salinity, high and low temperatures, and other damaging factors. Proline is the most abundant compound accumulated by plants in response to stress (Bassi & Sharma, 1993; Ashraf & Foolad, 2007; Verbruggen & Hermans, 2008). It is an osmoprotector, a stabilizer of macromolecules and membranes, an additional source of energy and nitrogen, an antioxidant (Kuznetsov & Shevyakova, 1999).

The dynamics of accumulation of phenolic compounds, which are important in the adaptation of the organism to environmental conditions, can also be used as a criterion for assessing the adaptive capacity of species (Zaprometov, 1993; Polyakova & Yershova, 2000; Lattanzio, 2013). Changes in the content of phenolic compounds depending on the growing conditions provide the basis of the ecological stability of natural populations in the process of evolution and fit into the general mechanism of plant adaptation to habitat conditions (Polyakova, 1993). Phenolic compounds, or polyphenols, are among the most common secondary metabolites of vascular plants and are formed in all plants' cells and tissues (Alscher & Hess, 2017). The structure of polyphenols is exceptionally diverse, as are the functions they perform. They are known to give color to flowers, fruits, and seeds and participate in plant growth and development regulation. They are chelators of heavy metals, regulate the expression of certain genes, and protect plants from stress (Bidel et al., 2010; Mierziak et al., 2014). Many flavonoids are allelochemicals involved in the formation of rhizobial symbiosis and mycorrhiza. The mechanism of action of allelochemical flavonoids is not fully understood; it is possible that they affect the auxin signaling of recipient plants and inhibit the growth of their cells, disrupt the synthesis of adenosine triphosphoric acid (ATP), induce the accumulation of ROS and, through calcium signaling, systemic root death (Mierziak et al., 2014). The accumulation level of phenolic compounds, to a certain extent, can serve as a

criterion for the potential resistance of plants to stress (Zagoskina, 2005).

Laboratory bioanalysis is the first step to investigate the possible manifestation of allelopathy in the relationships between plants, plants and microorganisms, or plants and insects (Kondratyev, 2017). It is a necessary tool both for studying the allelopathic potential of plant or soil extracts and assessing the activity of extracts in the purification and identification of allelopathic compounds. The bioanalysis method supposes the germination of recipient plants' seeds in Petri dishes on filter paper, sand, soil, or agar. This is a fast method for a large number of biological repetitions. It can be used to identify potential allelopathic effects under controlled laboratory conditions, using the percentage of germinated seeds of recipient plants to measure the allelopathic activity of compounds (Gawronska et al., 2006).

This work aimed to study the accumulation peculiarities of proline and phenolic substances in the leaves of *A. arguta* and *S. chinensis* in vegetation experiments and model their co-growth to optimize their cultivation.

Material and methods

The research was carried out at the Department of acclimatization of fruit plants of the M.M. Gryshko National Botanical Garden, National Academy of Sciences of Ukraine. The experiments were carried out under controlled conditions. The temperature was maintained within $22 \pm 2^\circ\text{C}$, soil moisture – $60 \pm 5\%$. Plants of *A. arguta* 'Sentyabrskaya' and *S. chinensis* 'Sadovy-1' were used as experimental objects. For each experiment combination, 12 two-year-old vegetatively propagated plants were planted in the last decade of May in 12-liter containers filled with dark gray forest light loamy soil in a ratio of 1:1, 2:1, and 1:2; monocultural plantings of these plants served as control. The experiments were triplicated. Samples for biochemical studies were taken in mid-August. The coefficient of plant resistance was calculated as the ratio of the amount of proline or flavonoids in the leaves of plants of mixed plantings to the amount of the corresponding substances in plants in monoculture.

Determination of proline

Determination of the content of free proline in plant leaves was carried out according to [Bates et al. \(1973\)](#). This method is based on the interaction of proline with a ninhydrin reagent, forming a pink-red color. The content of free proline was determined in a 2 g sample of plant material, which was ground in a mortar with quartz sand and 20 ml of an aqueous solution of sulfosalicylic acid. After that, 2 ml of the filtrate were mixed with 2 ml of acidic ninhydrin and 2 ml of glacial acetic acid in a test tube with a ground glass stopper. The mixture was kept for one hour in a boiling water bath and then cooled. Benzene (4 ml) was added to the test tubes with the cooled mixture and vigorously shaken until the orange color passed into the organic solvent. The upper colored layer was poured into cuvettes (20 mm), and the color density of the solution was measured using a FEK-56M photoelectric colorimeter. Extinction was determined on a blue filter with a wavelength of 520 nm. The amino acid concentration was calculated using a calibration curve built on standard proline solutions and expressed in mg/g of dry weight (DW).

Determination of polyphenolic compounds

The amount of polyphenolic compounds was determined by the Folin-Ciocalteu method ([Sibgatullina et al., 2011](#)), which is widely used to analyze herbal preparations. The method is based on the Folin-Ciocalteu reagent, a mixture of phosphotungstic and phosphomolybdate heteropoly acids, which are reduced by phenolic compounds in an alkaline medium. During the reaction, polyphenolic compounds are oxidized by the action of the reagent, and a blue color appears due to the formation of a mixture of reduced tungstates and molybdates. Namely, 0.05 g of raw material samples were triturated with 1.5 cm³ of 96% ethanol; phenolic compounds were extracted for 45 minutes at 45°C with periodic stirring (every 15 minutes) and subsequent centrifugation for 2 minutes at a rotational speed of 16,000 rpm. After that, 0.075 cm³ of samples were taken from the obtained extract, added 0.075 cm³ of the Folin-Ciocalteu reagent, diluted five times, and stirred. After 3 minutes, 0.15 cm³ of 20% sodium carbonate solution and 1.2 cm³ of distilled water were added, covered with a lid, stirred, and left at room temperature. After

one hour, the optical density of the formed tungsten blue was measured on a Specord-40 spectrophotometer at a wavelength of 725 nm. The total content of phenolic compounds was expressed in mg-equivalents of gallic acid per g of DW, the color intensity of which was proportional to the amount of phenolic compounds.

Determination of allelopathic activity

The allelopathic activity of the samples was determined according to the standard technique ([Grodzinsky, 1991](#)). To obtain root exudates, donor plants were grown in special funnels filled with quartz sand ([Lobkov & Konoshina, 2004](#)). After 14 days of vegetation, samples of aqueous solutions of root exudates were taken. The resulting aqueous solutions containing root exudates were evaporated in a Heidolph Laborota 4002 rotary vacuum evaporator at 30°C and residual air pressure of 30 mbar. The final solution volume was obtained at the rate of 1 ml from two and ten donor plants. The concentrated solutions were stored at below zero temperatures and used in further studies. An aqueous solution in a volume of 5 ml was placed in Petri dishes, into which 50 pieces of watercress (*Lepidium sativum* L.) seeds were sown. The dishes were kept in the dark at room temperature for three days. After that, the length of *L. sativum* seedling roots was measured. The experiments were carried out in five replicates. Water served as a control.

Statistical analysis

The significance of differences between the combinations of the experiment was established by the dispersion method according to Fisher's test and the significance level of the null hypothesis. The obtained indicators, determined with a 95% confidence interval, are trustworthy due to the high reliability of the arithmetic mean values (the calculated Student's test value significantly exceeds the table values) and the error indicator of experience less than 5%.

Results and discussion

As a result of the studies, it was found that in the leaves of *A. arguta* and *S. chinensis* grown in a mixed planting, a greater amount

Table 1. Accumulation of proline in *Actinidia arguta* and *Shisandra chinensis* plants and their stress resistance coefficient in monocultures and mixed plantings.

Nr	Experiment combination	<i>S. chinensis</i>		<i>A. arguta</i>	
		mg/g DW	k	mg/g DW	k
A	<i>S. chinensis</i> (control)	6.98±0.36			
A	<i>A. arguta</i> (control)			9.64±0.41	
B	<i>S. chinensis</i> (50%) / <i>A. arguta</i> (50%)	19±0.95	2.72	13.57±1.9	1.41
C	<i>S. chinensis</i> (33%) / <i>A. arguta</i> (67%)	21.19±0.81	3.04	9.53±0.35	0.99
D	<i>S. chinensis</i> (67%) / <i>A. arguta</i> (33%)	12.4±0.64	1.78	19.44±0.91	2.02
	LSD 0.95	1.88		1.65	

Note. DW – dry weight; k – stress resistance coefficient; LSD – least significant difference.

of proline accumulates in comparison with the monoculture (Table 1).

The maximum amount of proline (21.19±0.8 mg/g DW) was accumulated in *S. chinensis* leaves in the experiment combination with a prevailing ratio of actinidia plants in mixed planting (with 33% of *S. chinensis* and 67% of *A. arguta*). In this combination of mixed planting, the ratio of proline content in the leaves of schisandra to the corresponding value noted for the monoculture (k) was 3.04, which is the highest observed. The smallest k value (1.78) was noted in the experiment combination with 67% of *S. chinensis* and 33% *A. arguta*. This indicates that *A. arguta* plants have a repressing effect on *S. chinensis* plants in mixed plantings.

The amount of proline in *A. arguta* plants in the combination *S. chinensis* (33%) / *A. arguta* (67%) was 9.53±0.35 mg/g DW. A similar proline amount was registered for the control (monoculture) – 9.64±0.41 mg/g DW. In all other combinations of the vegetation experiments with mixed plantings, both the amount of proline and the ratio of the value of proline in mixed plantings were much higher than the corresponding values in monoculture. The maximum content of proline (19.44±0.91 mg/g DW) was observed in *A. arguta* in the experiment combination with the prevailing numbers of *S. chinensis* plants, with 67% of *S. chinensis* and 33% *A. arguta*, and with k=2.02. The smallest coefficient (k=0.99) was obtained in the combination *S. chinensis* (33%) / *A. arguta* (67%). Thus, with an increase in the number of *S. chinensis* plants in a vegetation container, *A. arguta* plants experience more severe stress (Table 1).

A similar relationship was noted for *S. chinensis* with an increase of *A. arguta* plants.

Many plant species naturally accumulate proline as the main organic osmolytes when subjected to various abiotic stresses. Therefore, this compound, which is effectively involved in the mechanisms of ensuring plant resistance to stress, particularly plays an adaptive role in osmotic adaptation and protects subcellular structures in plants under stress conditions (Verbruggen & Hermans, 2008; Chutia & Borah, 2012). The response of fennel plants (*Foeniculum vulgare* Mill.) to environmental constraints was a marked increase in proline content in its leaves (Zali & Ehsanzadeh, 2018).

The study of the features of the accumulation of phenols in the leaves of *A. arguta* and *S. chinensis* showed that their amount in *S. chinensis* leaves increases in mixed plantings with the prevailing number of *A. arguta* plants (Fig. 1).

The maximum accumulation of phenolic compounds (36.87±2.22 mg/g DW) in *S. chinensis* leaves was observed in the combination *S. chinensis* (67%) / *A. arguta* (33%), where the ratio of phenols in the leaves of plants in mixed plantings to the amount of phenols in monoculture plants was the highest (k=2.09). The ratio was much lower (k=1.12) in the combinations of *S. chinensis* (50%) / *A. arguta* (50%) and *S. chinensis* (33%) / *A. arguta* (67%). In leaves of *A. arguta* in monoculture, and in combinations *S. chinensis* (33%) / *A. arguta* (67%), the phenol content differed insignificantly (24.88±1.43 mg/g DW and 23.82±1.47 mg/g DW, respectively). In the experiment combinations *S. chinensis*

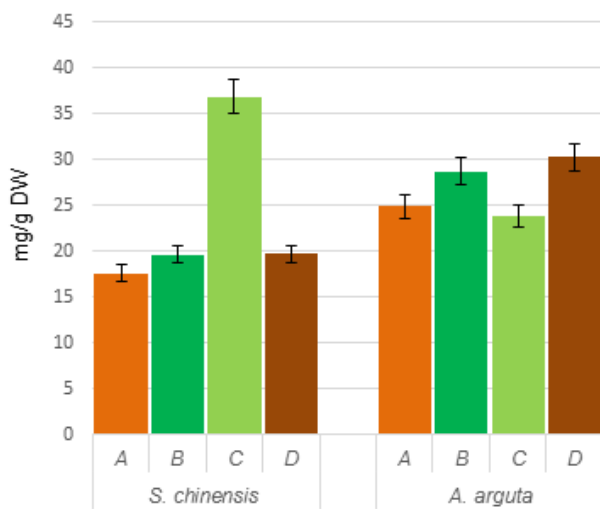


Figure 1. Accumulation of phenolic substances in plants of *Actinidia arguta* and *Schisandra chinensis* in monocultures and mixed plantings: **A** – *A. arguta* (control); **B** – *S. chinensis* (50%) / *A. arguta* (50%); **C** – *S. chinensis* (33%) / *A. arguta* (67%); **D** – *S. chinensis* (67%) / *A. arguta* (33%).

(50%) / *A. arguta* (50%) and *S. chinensis* (33%) / *A. arguta* (67%), an accumulation of a higher content of phenolic compounds in *A. arguta* leaves was observed. The highest coefficient of resistance to stress, $k=1.22$, was recorded in the experiment combination with the prevailing number of *S. chinensis* plants, namely *S. chinensis* (67%) / *A. arguta* (33%).

Thus, it was found that the content of secondary metabolites of phenolic nature in the leaves of plants changes depending on the ratio of experimental plants in the compositions of mixed plantings (Fig. 1). The research results revealed interspecific competition of plants in their co-cultivation, which leads to stress and can be manifested in relation to mineral nutrients, soil moisture, or be caused by root secretions of plants. The concentration and amount of biosynthesized phenolic compounds increase responding to stress factors. A similar reaction of plants is reported in several publications. In particular, the water stress increased the amount of the phenolic compounds in the vegetative organs of *Hypericum brasiliense* Choisy (Abreu & Mazzafera, 2005). Similarly, water-deficit stress increased the productivity of chlorogenic acid, catechin and epicatechin in hawthorn species (Kirakosyan et al., 2004).

To compare the allelopathic potential of the experimental plants, we studied the allelopathic activity of their root exudates by

the biotesting method using the *L. sativum* as a test object (Fig. 2).

The allelopathic activity of the root exudates of *A. arguta* and *S. chinensis* plants was established. Their noticeable phytotoxic effect on the growth of roots of *L. sativum* seedlings was found. Compared with the control, the exudate of *A. arguta* suppressed *L. sativum* roots growth by 12.0% and *S. chinensis* – by 30.0%. A decrease in the growth of *L. sativum* roots under the influence of aqueous solutions of root exudates indicates the presence of dissolved organic substances in the soil, which inhibit the development of test objects. As noted above, *S. chinensis* exhibits a stronger inhibitory effect on the test culture compared to *A. arguta*. These results confirm the previously obtained data that intravital secretions (leaf and root exudates) of *S. chinensis* and *A. arguta* contain allelopathically active compounds – allelochemicals, which have a depressing effect on seed germination and growth of seedlings of various test objects (Osipova, 1997).

Conclusions

The results of the vegetation experiment indicate a pronounced interaction between *A. arguta* and *S. chinensis* crops during joint growth. The percentage of plants in model experiments significantly affects the accumulation of primary and secondary metabolites in leaves.

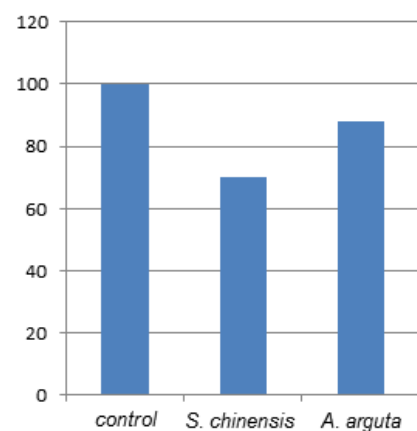


Figure 2. Allelopathic activity of root secretions of *Actinidia arguta* and *Schisandra chinensis* (bioassay – root growth of *Lepidum sativum*), percentage to control.

The result of a comparative analysis of the amount of proline and phenolic compounds can be used to assess the mutual influence of plants in mixed plantings to optimize their growing conditions, which confirms the diagnostic significance of these metabolites for indicating the stress state of the studied plants.

The inhibitory effect of *S. chinensis* and *A. arguta* root secretions on *L. sativum* indicates their allelopathic activity, more pronounced in *S. chinensis*.

Based on our results, it is concluded that monoculture cultivation is the most optimal for both studied species. However, for the co-growth of *A. arguta* and *S. chinensis*, planting in the ratio of 2:1 is the best.

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Діагностичне значення синтезу фенольних сполук і проліну в листках *Schisandra chinensis* та *Actinidia arguta* для індикації рівня стресу рослин в умовах змішаних насаджень

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Досліджено особливості накопичення поліфенольних сполук та вільного проліну в листках *Actinidia arguta* та *Shisandra chinensis* при їх вирощуванні в вегетаційних дослідах з різним співвідношенням кількості рослин в контейнерах, а саме 1 : 1, 2 : 1, 1 : 2. В якості контролю використовували одновидові посадки рослин. Вміст вільного проліну в листках рослин здійснювали за методикою, яка заснована на взаємодії проліну з реактивом нінгідрину, утворюючи рожево-червоне забарвлення. Кількість поліфенольних сполук визначали методом Фоліна-Чіокальтеу. Встановлено, що співвідношення рослин в контейнерах істотно впливає на накопичення первинних і вторинних метаболітів у їх

листках. За умов змішаної посадки в листках рослин накопичувалося більше проліну та фенольних речовин порівняно з монокультурою. Максимальний вміст проліну $19,44 \pm 0,91$ мг/г сухої маси (СМ) спостерігався в листках *A. arguta* у варіанті з переважаючою кількістю рослин лимонника при співвідношенні рослин *S. chinensis* і *A. arguta* 2 : 1. У цьому ж варіанті найбільша кількість фенольних сполук ($36,87 \pm 2,22$ мг/г СМ) була у листках актинідії. Досліджувані кореневі екsudати *S. chinensis* і *A. arguta* мали інгібуючу алелопатичну дію на тест-культуру. Ексудати *A. arguta* викликали середнє зниження росту коренів тест-об'єктів на 12,0 %, а *S. chinensis* – на 30,0 % порівняно з контролем. Це дозволяє зробити висновок про високу активність алелохімікатів лимонника, які негативно впливають на ріст рослин актинідії. Оптимальне співвідношення рослин *A. arguta* і *S. chinensis* при спільному вирощуванні становить 2 : 1, оскільки вища концентрація рослин лимонника в контейнері посилює стрес у рослин актинідії, що виражається в підвищеному накопиченні фенолів і проліну в її листках. Результат порівняльного аналізу кількості проліну та фенольних сполук може бути використаний для оцінки взаємного впливу рослин у змішаних насадженнях з метою оптимізації умов їх вирощування, що підтверджує діагностичну значущість цих метаболітів для індикації стресового стану рослин.

Ключові слова: *Actinidia arguta*, *Shisandra chinensis*, фенольні сполуки, пролін, монокультура, змішані насадження, алелопатична активність