

DIAZOTROPH ACTIVITY REGULATING STRATEGY UNDER THEIR INTRODUCTION IN AGROCENOSSES

S. F. Kozar

Institute of Agricultural Microbiology and Agroindustrial Manufacture, NAAS
97 Shevchenka str., Chernihiv, 14035; Ukraine; e-mail: kozarsf@gmail.com

Objective. Investigate approaches to managing the activity of soil diazotrophs and propose a strategy for its regulation. **Methods.** Theoretical, vegetation and field experiments, microbiological, gas chromatographic, mathematical and statistical. **Results.** The activity of beneficial soil microorganisms can change under the action of temperature, humidity, chemical compounds of various origin, and other microorganisms. It was established that, taking into account a significant variety of factors, it is necessary to develop a set of specific ways to increase the growth and functional activity of nitrogen-fixing bacteria, as well as their viability. It has been proved that the combination of diazotrophs forms an effective symbiotic leguminous-rhizobial system, which provides additional biological nitrogen in agroecosystems. At the same time, there was an increase in plant mass, chlorophyll content in the leaves, protein and oil content in the products. The combined use of diazotrophs increases the yield, in particular, soybeans by 9–16 % compared with inoculation by pure bacterial culture. **Conclusion.** Based on the analysis and generalization of the obtained research results, a strategy for regulating the activity of diazotrophs for their effective introduction into agroecosystems is proposed, which consists in combining bacteria of different species, selecting conditions for their co-cultivation and application upon stabilisation of the number of viable bacterial cells. The proposed strategy involves solving the problem by obtaining an inoculant, which is characterized by a high titre and a stable number of viable cells, which allows to obtain an effective nitrogen-fixing system. The strategy is tried-and-tested on the example of regulating the growth and functional activity of soybean nodule bacteria by combining diazotrophs of different species, substantiating the conditions of their co-cultivation and application to ensure positive interaction in the form of commensalism, as well as by regulating viability of diazotrophs by adding stabilisers to the medium.

Key words: nitrogen-fixing bacteria, growth activity, nitrogen fixation, viability, diazotroph activity regulating strategy.

Introduction. Modern ideas about the interaction of microorganisms and plants suggest that soil nitrogen-fixing bacteria play an important role in agroecosystems. The positive effect of diazotrophs is due to their ability to fix atmospheric nitrogen and supply it to plants, as well as to produce physiologically active compounds that stimulate the growth and development of crops. As specific beneficial bacteria are not always present in the soil in sufficient quantities, appropriate inoculants and methods of their use are being developed in Ukraine and other countries. At the same time, diazotrophs do not fully realize their potential under the influence of negative environmental factors,

which can negatively affect the efficiency of their use. Therefore, it is important to develop and implement a strategy to regulate the activity of nitrogen-fixing bacteria during their introduction into agroecosystems of crops, which would solve this problem

Analysis of the novel studies and publications. The final objective of the use of selected highly effective nitrogen-fixing bacteria is to increase the productivity of crops and obtain quality plant products [1–3]. At the same time, it is necessary to achieve economic and energy efficiency of this agricultural measure, as well as maximum maintenance of soil resources.

Since the main function of introduced dia-

zotrophs is the fixation of atmospheric nitrogen [4], it is necessary to ensure the formation of an effective nitrogen-fixing system, which occurs as a result of the interaction between bacteria and a particular crop. The formation of such a system is impossible without the introduction of a sufficient number of diazotrophs into the appropriate agrocenosis, which can be done by using inoculum with a high titre. However, even having such an inoculant, it is necessary to ensure maximum stability of viable bacterial cells during storage and use [5].

The titre of nitrogen-fixing cells in the inoculant depends on their growth activity during cultivation [6]; the stability of the number of diazotrophs that can be introduced into agrocenoses depends on the viability of these microorganisms [7–9], and the formation of an effective nitrogen-fixing system — also on the implementation of the functional activity of microorganisms.

In ecosystems, microorganisms cannot be considered in isolation, because there is a constant interaction between them [10–14], which can significantly affect their activity. Therefore, it is important to study mixed cultures of microorganisms, including soil nitrogen-fixing bacteria, for maximum implementation of their potential for practical purposes.

In view of the above, it is important to determine the functional activity and viability of diazotrophs under the action of biotic and abiotic factors, as these studies may be the basis of a strategy to regulate the activity of beneficial soil bacteria

Objective. Investigate approaches to managing the activity of soil diazotrophs and propose a strategy for its regulation.

Materials and methods. Bacteria used in the work. *Azospirillum brasilense* sp. 7 (typical strain), *Azospirillum brasilense* 410 (V. V. Volkohon), *A. brasilense* 18-2 (V. I. Likhova, O. V. Nadkernychna), *Azotobacter chroococcum* M-70 (Yu. M. Mochalov, V. I. Kanivets), *Azotobacter vinelandii* M-X (Yu. M. Mochalov, V. I. Kanivets), *Bradyrhizobium japonicum* M-8 (M. Z. Tolkachev), *B. japonicum* 6346 (A. T. Novikova), *Rhizobium leguminosarum* bv. *viceae* 2486, *Rh. leguminosarum* bv. *viceae* 250a (A. F. Antypchuk, A. T. Novikova), *Rh. leguminosarum* 31 (M. Z. Tolkachev), *Rhizobium radiobacter* = *Agrobacterium radiobacter* 204 (M. K. Sherstoboev, A. V. Khotianovych,

V. P. Patyka), *Enterobacter aerogenes* 30-φ (O. O. Berestetskyi, A. V. Yermolina, V. P. Patyka), *Pseudomonas fluorescens* B-17 (typical strain). Microorganisms were obtained from the Collection of Beneficial Soil Microorganisms of the Institute of Agricultural Microbiology and Agroindustrial Manufacture of the NAAS. We sincerely gratitude to the authors for their kindly provided strains

Agricultural crops. Soybean of the varieties Lehenda, Suziria, Poltava. Winter wheat of the variety Poliska 90. Winter rye of the variety Borotba. Winter barley of the variety Honar.

The experiments were performed at the Institute of Agricultural Microbiology and Agroindustrial Manufacture of the NAAS on leached light loam chernozem, which contains from 2.8 % to 3.4 % of humus (according to Tiurin), from 0.27 % to 0.31 % of total nitrogen, about 15 mg/100 g of soil P₂O₅ (according to Kirsanov), from 13 mg/100 g of soil to 16 mg/100 g of K₂O (according to Maslova), pH = 5.9–6.5. Planning and conducting field experiments were performed according to Dospiekhov [15].

The number and growth parameters of microorganisms in the bacterial suspension were determined by microbiological methods [16–19]. Studies of the nitrogen fixation activity of microorganisms were performed by the acetylene method [20; 21] on a Chrom-4 gas chromatograph with a flame ionization detector. Steel sorption columns were filled with sorbent Porapak Q 60–80 mesh. Thermostat temperature was 40 °C. Consumption of gases: hydrogen — 15 cm³/min, nitrogen — 100 cm³/min, air — 500 cm³/min.

The number of representatives of certain physiological and trophic groups of microorganisms in the rhizosphere soil of plants was determined by the plate method via deep seeding on agar media [21; 22]: meat-peptone agar (MPA) to account for microorganisms that use mainly organic forms of nitrogen, starch-ammonia agar (SAA) — for microorganisms that absorb mainly nitrogen of mineral compounds. The number of micromycetes was determined on acidified Czapek medium.

Statistical processing was performed according to generally accepted methods in mathematical statistics using Microsoft Excel. To assess the significance of the differences between the variants of the experiments, the least signifi-

cant difference was calculated (HIP₀₅).

Results and discussion. The activity of nitrogen-fixing bacteria is not stable and unchangeable, as we have shown after a range of different experiments. In particular, the effect on diazotrophs was studied:

- 1) chemical substances of different nature [23–26];
- 2) soil microorganisms [27–33];
- 3) abiotic environmental factors [34–36].

Based on the analysis of the obtained results, a general strategy for regulating growth and functional activity, as well as the viability of diazotrophs is proposed. This strategy involves combination of diazotrophs of different species, selection of conditions for their co-cultivation and use upon stabilizing the number of viable bacterial cells (Fig. 1).

The strategy is developed on the pattern of regulating the activity of *B. japonicum* to increase productivity of a particular crop, namely soybeans.

Given the extremely large number of different factors that can potentially affect soybean nodule bacteria, it was necessary to identify specific factors that can significantly affect the activity and viability of the studied microorganisms. In our opinion, it is necessary to minimize the number of selected factors, as their increase may reduce the ability to manage them.

When selecting the factors that affect the introduction of nodule bacteria in soybean agrocenoses, it is necessary to take into account the fact that the regulation of growth and functional activity of diazotrophs necessitates activation of the vital processes of microorganisms, and improvement in viability is achieved by reducing bacterial activity, i. e. by bringing them to a state of rest and maintenance in such a state for the required period. Therefore, there is a need to divide the factors into two blocks.

Since in nature, soil diazotrophs function upon constant interaction with other bacteria [37–39], for example, nodule bacteria with associative and free-living nitrogen fixers, one of the effective biotic factors is the use of mixed cultures of microorganisms to increase their growth and functional activity. A feature of our proposed approach to the selection and combination of *B. japonicum* with other soil diazotrophs is the use of microorganisms complementary to nodule bacteria to manage their activity. At the same time, the known combinations of beneficial soil microorganisms in the inoculant aim to expand the spectrum of action of biological preparations: e. g. to improve nitrogen and phosphorus nutrition [40; 41].

Quite often these combinations are mechanistic in nature, without a comprehensive scientific substantiation and without answers to the

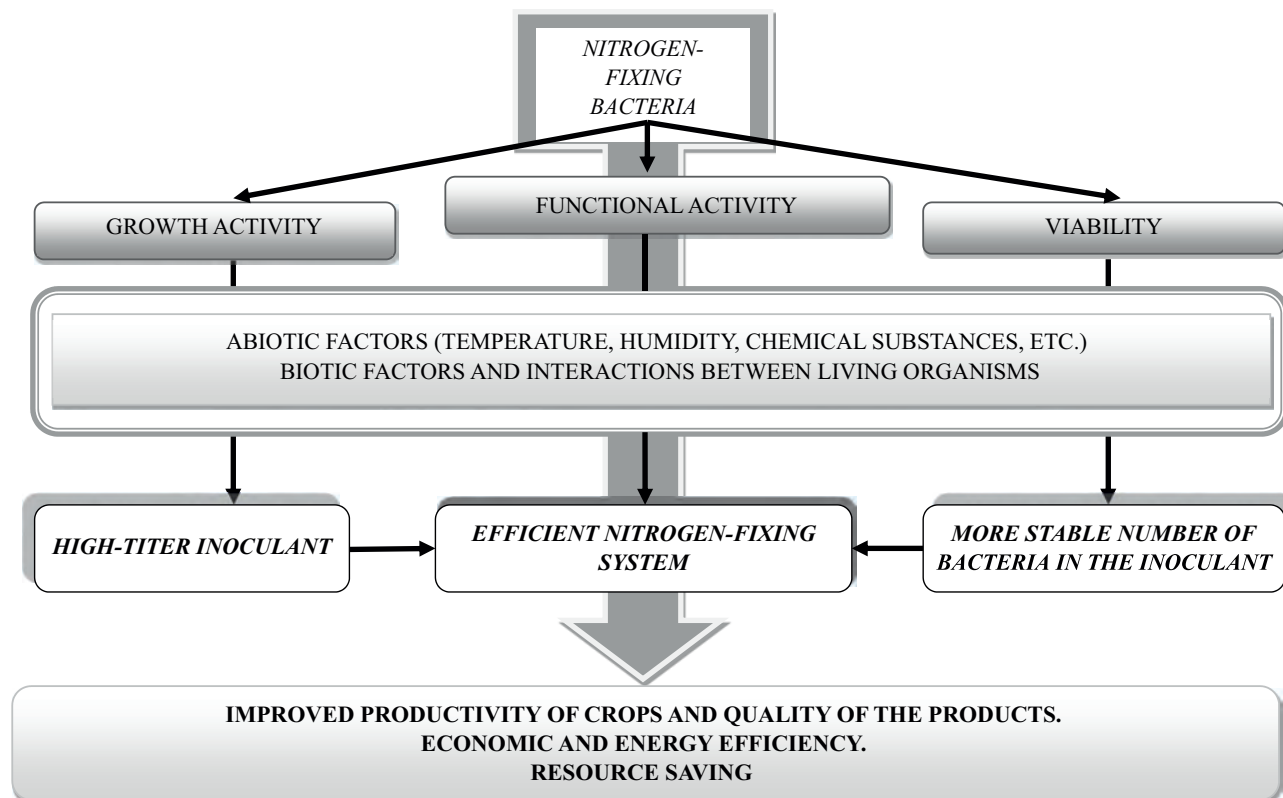


Fig. 1. General strategy for regulating activity of soil diazotrophs.

following questions: how does one microorganism affect the growth activity of another; what is the optimal quantitative ratio of the cells of different types of microorganisms; whether the effect of combining two or more microorganisms is greater than the simple sum of the effects of each individual microorganism, etc.

Furthermore, an important factor influencing diazotrophs during their introduction into agroecosystems is ensuring the viability of these bacteria [42; 43], which is influenced by a number of factors [44–47]: temperature, humidity, chemical substances of different nature, etc. The maintenance of diazotrophs can be achieved by transferring bacteria to a state of rest and protective action of certain substances due to the ability to form a protective cover [48; 49].

It is proposed to manage the activity of *B. japonicum* by regulating their growth and functional activity due to combining diazotrophs with the selection of conditions for their co-cultivation and use to ensure positive interaction in the form of commensalism, as well as by regulating the viability of diazotrophs by adding stabilizers to the medium (Fig. 2).

At the next stage, a number of studies were conducted to confirm the correctness of the selected strategy. Initially, the growth activity of soybean nodule bacteria under the action of sus-

pensions containing metabolites of other diazotrophs was studied. No significant positive effect of suspensions of nitrogen-fixing bacteria belonging to rhizobia, azotobacter, pseudomonads, agrobacteria and enterobacter on the growth of soybean nodule bacteria was found. At the same time, high concentrations of suspensions of these microorganisms inhibited the growth of *B. japonicum* strains. Addition of the bacterial suspension with metabolites of *A. brasilense* in low concentrations to the culture medium for the cultivation of nodule bacteria had a positive effect on the growth activity of soybean nodule bacteria.

Upon mixed cultivation of *B. japonicum* and *A. brasilense*, microorganisms can directly affect the viability of bacterial cells of other species, resulting in an increase in both growth and functional activity. Given the different needs of the studied bacteria in food sources, we have developed a new semi-synthetic nutrient medium for their co-cultivation [50].

The production of physiologically active compounds by diazotrophs in pure and mixed culture has been studied. It has been shown that the amount of cytokinins and gibberellins increases, the content of abscisic acid and the auxin/cytokinin ratio decreases compared to variants with pure cultures of microorganisms in

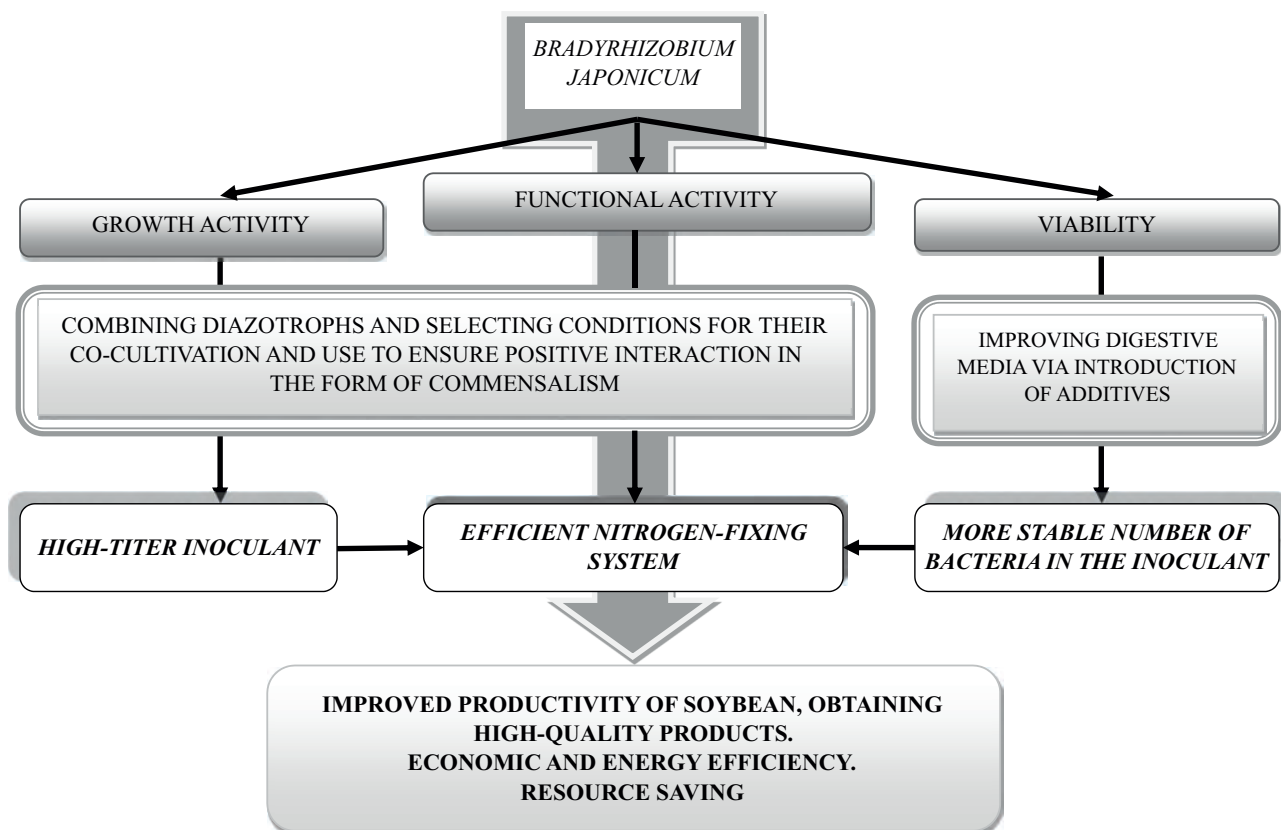


Fig. 2. Strategy for regulating *Bradyrhizobium japonicum* activity.

the culture fluid of nodule bacteria and azospirilla when they are co-cultivated [51]. The use of soybean nodule bacteria and azospirilla in a mixed culture promotes the formation of a more efficient symbiotic apparatus: the number and weight of nodules increases, nitrogen-fixing activity improves [52].

Various chemical compounds have been tested as stabilizers in the medium. During storage of the inoculum, the titre of bacteria grown with stabilizers decreases more slowly, which indicates the feasibility of using these substances to extend the shelf life of *B. japonicum*. The most effective stabilizer, which in our studies was sodium alginate, and its optimal concentration, which helps maintain the viability and functional activity of soybean nodule bacteria in a liquid medium for 4 months were selected [53; 54].

When using the inoculant based on nodule bacteria and azospirilla with sodium alginate, an increase in the mass of soybean plants in all phases of development, an increase in soybean yield by 0.23–0.4 t/ha, or 9–16 %, compared with inoculation with pure culture of nodule bacteria, was reported. At the same time, the protein and oil content in the products increased.

The combined use of *B. japonicum* and

A. brasilense is characterized by a higher level of both economic and energy efficiency compared to the variant without inoculation and the variant with a pure culture of nodule bacteria. The implementation of the strategy for managing the activity of diazotrophs during their introduction into soybean agroecosystems is schematically presented in Fig. 3.

Conclusion. Based on the analysis and generalization of our study results, a strategy for regulating the activity of diazotrophs for their effective introduction into agroecosystems is proposed, which consists in combining bacteria of different species, selecting conditions for their co-cultivation and use upon stabilisation of the number of viable bacterial cells. The proposed strategy involves solving the problem by obtaining an inoculant, which is characterized by a high titre and a more stable number of viable cells, which allows to obtain an efficient nitrogen-fixing system

The strategy is tested on the pattern of regulating the growth and functional activity of soybean nodule bacteria by combining diazotrophs of different species, justifying the conditions of their co-cultivation and use to ensure positive interaction in the form of commensalism, and by regulating the viability of diazotrophs via adding stabilizers to the medium.

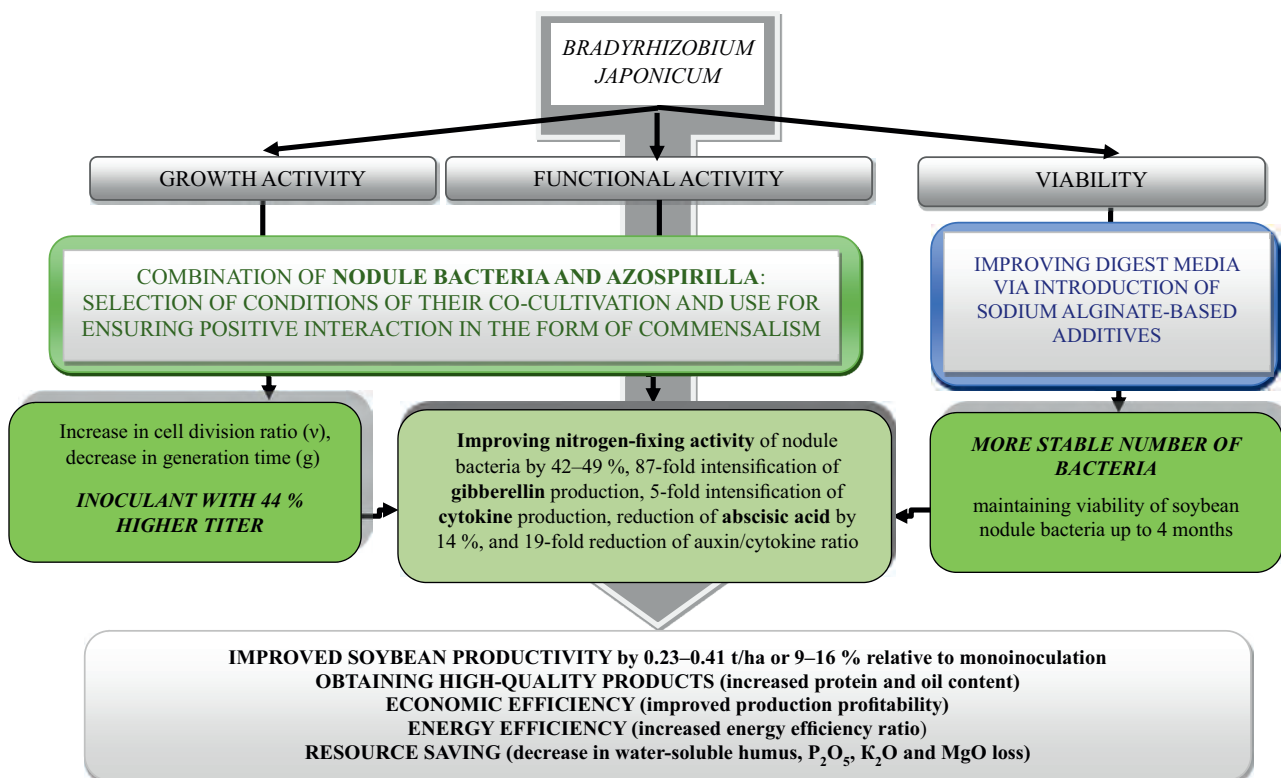


Fig. 3. Implementation of the strategy for regulating *Bradyrhizobium japonicum* activity.

REFERENCES

1. Patyka, V. P. (2003). *Biologichnyi azot* [Biological nitrogen]. Kyiv [in Ukrainian].
2. Volkohon, V. V. (2006). *Mikrobnii Preparaty v Zemlerobstvi. Teoriia i praktyka* [Microbial Preparations in Agriculture. Theory and practice]. Kyiv [in Ukrainian].
3. Volkohon, V. V. (2007). *Mikrobiologichni aspekty optymizatsii azotnoho udobrennia silskohospodarskykh kultur* [Microbiological aspects of nitrogen fertilizer optimization of agricultural crops]. Kyiv [in Ukrainian].
4. Morhun, V. V., Kots, S. Ya. (2018). Rol biologichnoho azotu v azotnomu zhyvlenni roslin. [The role of biological nitrogen in nitrogen nutrition of plants]. *Visnyk Natsionalnoi Akademii Nauk Ukrainy*, 1, 62–74, Kyiv [in Ukrainian].
5. Kurdysh, Y. K., Melnykova, N. N. (2011). Vlyianye hlynystykh myneralov na rost y noduliat-syonnuuiu aktyvnost Bradyrhizobium japonicum. [Influence of clay minerals on growth and nodulation activity of Bradyrhizobium japonicum]. *Mikrobiologichnyi zhurnal*, 73(4), 36–40 [in Ukrainian].
6. Stephens, J. H. G., Rask, H. M. (2000). Inoculant production and formulation. *Field crops research*, 65, 249–258. [https://doi.org/10.1016/S0378-4290\(99\)00090-8](https://doi.org/10.1016/S0378-4290(99)00090-8)
7. Date, R. A. (2001) Advances in inoculant technology: a brief Review. *Australian journal of experimental agriculture*, 41, 3, 321–325. <https://doi.org/10.1071/EA00006>
8. Brockwell, J., Bottomley, P. J. (1995). Recent advances in inoculant technology and prospects for the future. *Soil Biology and Biochemistry*, 27 (4–5), 683–697. [https://doi.org/10.1016/0038-0717\(95\)98649-9](https://doi.org/10.1016/0038-0717(95)98649-9)
9. Crawford, S. L., Berryhill, D. L. (1983). Survival of Rhizobium phaseoli in coal-based Legume inoculants applied to seeds. *Applied and environmental microbiology*, 45(2), 703–705. <https://doi.org/10.1128/aem.45.2.703-705.1983>
10. Argaw, A. (2012). Evaluation of co-inoculation of Bradyrhizobium japonicum and phosphate solubilizing Pseudomonas spp. effect on soybean (Glycine max L. (Merr.)) in Assossa Area. *Journal of agricultural science and technology*, 14, 213–224.
11. Askary, M., Mostajeran, A., Amooaghai, R., & Mostajeran, M. (2009). Influence of the co-inoculation Azospirillum brasilense and Rhizobium meliloti plus 2,4-D on grain yield and N, P, K content of Triticum aestivum (cv. Baccros and Mahdavi). *American-Eurasian Journal of Agricultural & Environmental Science*, 5(3), 296–307.
12. Vorobei, N. A., Patsko, O. V., Kots, S. Ia., & Parshykova, T. V. (2009). Fiziologichni osoblyvosti rozvytku liutserny za inokuliatzii zmishanymy kultu-ramy azotfiksuvalnykh mikroorhanizmiv [Physiological features of alfalfa development by inoculation with mixed cultures of nitrogen-fixing microorganisms]. *Fyzyolohyia y byokhymyia kulturnykh rastenyi — Physiology and Biochemistry of Cultivated Plants*, 41 (1), 344–352. [in Ukrainian].
13. Aung, T. T., Tittabutr, P., Boonkerd, N., Herridge, D., & Teamroong, N. (2013). Co-inoculation effects of Bradyrhizobium japonicum and Azospirillum sp. on competitive nodulation and rhizosphere eubacterial community structures of soybean under rhizobia-established soil conditions. *African Journal of Biotechnology*, 12 (20), 2850–2862.
14. Murodova, S. S., Davranov, K. D. (2014). Kompleksnyie mykrobynye preparaty. Prymenenye v selskokhoziaistvennoi praktyke [Complex microbial preparations. Application in agricultural practice]. *Biotechnologia Acta*, 7 (6). (pp. 92–101) [in Ukrainian].
15. Dospikhov, B. A. (1985). *Metodyka polevoho opyita (s osnovamy statystycheskoi obrabotky rezultatov yssledovanyi)* [Methods of field experience (with the basics of statistical processing of research results)]. Moskva: Ahropromyzdat [in Russian].
16. Shlehel, H. (1987). *Obshchaia mykrobyolohyia* [General microbiology]. Moskva: Myr [in Russian].
17. Shylnykov, V. K. (2005). *Praktykum po mykrobyolohyy* [Workshop on Microbiology]. Moskva: Drofa [in Russian].
18. Bykov, V. A., Vynarov, A. Iu., & Sherstobytov, V. V. (1985). *Raschet protsessov mykrobyolohycheskykh proyzvodstv* [Calculation of microbiological production processes]. Kyiv: Tekhnyka [in Russian].
19. Pert, S. D. (1978). *Osnovy kulturyrovanyia mikroorhanyzmov y kletok* [Fundamentals of Culturing Microorganisms and Cells]. Moskva: Myr [in Russian].
20. Hardy, R. W. F., Hotsren, R. D., Jackson, E. K., & Burns, R. C. (1968). The acetylene-ethylene assay for N₂ fixation: laboratory and field evaluation. *Plant Physiol*, 43 (8), 1185–1207. <https://doi.org/10.1104/pp.43.8.1185>
21. Hardy, R. W. F., Burns, R. C., Holsten, R. D. (1973). Applications of the acetylene-ethylene assay for measurement of nitrogen fixation. *Soil Biology and Biochemistry*, 5 (1), 47–81.
22. Volkohon, V. V. (2010). *Eksperymentalna gruntova mikrobiolohiia* [Experimental soil microbiology]. Kyiv: Ahrarna nauka [in Ukrainian].
23. Kozar, S. F., Symonenko, E. P., Lynnyk, V. O., & Kaplunenka, V. H. (2015). Influence of nanocarboxylates of microelements on Rhizobium radiobacter 204 growth-regulating activity. *Silskohospodarska mikrobiolohiia — Agricultural microbiology*, 22, 3–8. <https://doi.org/10.35868/1997-3004.22.3-8>

24. Ievtushenko, T. A., Kozar, S. F., & Pyshchur, I. M. (2018, October). Vplyv nanokarboksylativ mikroelementiv na roztovu i funktsionalnu aktyvnist Bradyrhizobium japonicum [Influence of microelement nanocarboxylates on growth and functional activity of Bradyrhizobium japonicum]. *Mikrobiologhiia v suchasnomu silskohospodarskomu vyrobnytstvi: tezy dopovidi XI naukovoï konferentsii molodykh vchenykh* (pp. 20–22), Chernihiv [in Ukrainian].
25. Kozar, S. F., Yevtushenko, T. A., Usmanova, T. O., & Symonenko, Ye. P. (2017). Mikrobiotsenoz ryzosferneho gruntu ta produktyvnist pshenytsi ozymoi za vykorystannia bakterii Agrobacterium radiobacter 204, aktyvovanykh nanokarboksylatamy metaliv [Microbiocenosis of the rhizosphere soil and productivity of winter wheat for the growth of Agrobacterium radiobacter 204 bacteria, activated by metal nanocarboxylates]. *Silskohospodarska mikrobiologhiia — Agricultural microbiology*, 26, 17–23 [in Ukrainian]. <https://doi.org/10.35868/1997-3004.30.13-19>
26. Kozar, S. F., Symonenko, E. P., Volkohon, V. V., & Volkogon, M. V. (2019). Nanocarboxylates of molybdenum and of iron enhance the functional activity of Rhizobium radiobacter 204. *Applied Nanoscience*, 9(5), 795–800. <https://doi.org/10.1007/s13204-018-00939-6>
27. Kozar, S. F. (2003). Antahonistychni vidnosyny batsyl iz predstavnykamy gruntovykh diazotrofov ta perspektyvy vykorystannia yikhnikh kompleksiv pry vyroshchuvanni silskohospodarskykh kultur [Antagonistic species of bacilli from representatives of primordial diazotrophs and the prospect of victorious complexes in growing silky cultures]. *Visnyk ahrarnoi nauky Prychornomia — Ukrainian Black Sea region agrarian science*, 1, 302–307 [in Ukrainian].
28. Kozar, S. F., Ushakova, M. A. (2004). Antahonizm azotobaktera shchodo gruntovykh diazotrofov ta vplyv kompleksiv bakterii na rist Roslyn [Antagonism of azotobacter from rodent diazotrophs and infusion of bacterium complexes onto the growth of roslin.]. *Ahroekologichnyi zhurnal — Agroecological journal*, 2, 31–34 [in Ukrainian].
29. Kozar, S. F. (2008). Kompleksne zastosuvannia diazotrofov dlia pidvyshchennia urozhainosti ozymoho zhyta [Integrated use of diazotrophs for increasing the yield of winter crops.]. *Zbirnyk naukovykh prats Umanskoho derzhavnoho ahrarnoho universytetu: osnovy formuvannia produktyvnosti silskohospodarskykh kultur za intensyvnykh tekhnolohii vyroshchuvannia — Collected works of Uman state university of horticulture: bases of formation of productivity of agricultural crops at intensive technologies of cultivation* (pp. 312–316) [in Ukrainian].
30. Kozar, S. F., Ushakova, M. A., & Pohorylko, N. V. (2002). Stvorennia shtuchnykh asotsiatsii diazotrofov dlia inokulatsii ozymoho zhyta [The creation of piece associations of diazotrophs for the inoculation of winter crops]. *Stalyi rozvytok ahroekosystem: tezy dopovidi mizhnarodnoi naukovoï konferentsii* (pp. 105–107). Vinnytsia [in Ukrainian].
31. Kozar, S. F. (2005). Biologichna efektyvnist kompleksnoho zastosuvannia mikrobnnykh preparativ [Biological efficiency of complex storage of microbial preparations]. *Silskohospodarska mikrobiologhiia — Agricultural microbiology*, 1–2, 86–94 [in Ukrainian].
32. Kozar, S. F. (2006). Vplyv mikrobnnykh preparativ na aktyvnist mikroflory ryzosferneho gruntu yaroho yachmeniu [Infusion of microbial preparations on the activity of microflora in rhizosphere soil of bright barley]. *Visnyk KhNAU im. V. V. Dokuchaieva — The Bulletin of the kharkiv national agricultural university of V. V. Dokuchaev*, 4, 126–133 [in Ukrainian].
33. Kozar, S. F. (2005). Byolohycheskye elementy tekhnolohii vyirashchuvannia yarovoho yachmenia na osnove kompleksnoho yspolzovannia mykrobnnykh preparativ [Biological elements of the technology of growing spring barley based on the integrated use of microbial preparations]. *Chernihivskiy derzhavnyi tsentr naukovo-tekhnichnoi i ekonomichnoi informatsii*, 4 [in Ukrainian].
34. Kozar, S. F., Nesterenko, V. M., Yevtushenko, T. A., Firsovskiy, O. V., & Usmanova, T. O. (2013). Efektyvnist zastosuvannia mikrobnnoho preparatu ABT v tekhnolohii vyroshchuvannia tsybuli ripchastoi [The effectiveness of the intake of the microbial drug ABT in the technology of viral cybulization]. *Naukovyi visnyk Natsionalnoho universytetu bioresursiv i pryrodokorystuvannia Ukrainy (Seriiia "Ahronomiia") — Scientific Bulletin of the National University of Life and Environmental Sciences of Ukraine (Series "Agronomy")*. 183 (1), 207–214 [in Ukrainian].
35. Bilokonska, O., Kozar, S. (2020). Viability of Azotobacter chroococcum IMB B-7836 and their influence on cucumber productivity. *Anale Universitații din Oradea, Fascicula Biologie.; XXVII, 2*, 111–115.
36. Bilokonska, O. M., Kozar, S. F., Yevtushenko, T. A., & Usmanova, T. O. (2018). Zberezhenist bakterii rodu Azotobacter na nasinni Cucumis sativus L. za dii riznykh temperature [Preservation of bacteria of the genus Azotobacter on plants Cucumis sativus L. for different temperatures. *Silskohospodarska Mikrobiologhiia — Agricultural microbiology*, 27, 11–17. [in Ukrainian]. <https://doi.org/10.35868/1997-3004.27.11-17>
37. Goldenfeld, N., Woese, C. (2007). *Biology's Next Revolution. Nature*. 445. <https://doi.org/10.1038/445369a>

38. Patyka, M. V., Tanchyk, S. P., Kolodiazhy, O. Yu., Ivaniuk, M. F., Kruhlov, Yu. V., Melnychuk, M. D., & Patyka, T. I. (2012). Formuvannya bioriznomanittia ta filotypovoi struktury eubakterialnoho kompleksu chornozemu typovoho pry vyroshchuvanni pshenytsi ozymoi [Formation of bioriznomanittya and phylotype structure of eubacterial complex black soil typical for growing winter wheat]. *Dopovidi Natsionalnoi akademii nauk Ukrainy — Reports of the National Academy of Sciences of Ukraine*, 11, 163–171 [in Ukrainian].
39. Lenheler, Y., Drevs, H., & Shlehel, H. (2005). *Sovremennaia mykrobiologiya. Prokaryoty: v 2-kh tomakh* [Modern microbiology. Prokaryotes: in 2 volumes] Moskov: Myr. [in Russian].
40. De-Bashan, L. E., Hernandez, J., Morey, T., & Bashan, Y. (2004). Microalgae growth-promoting bacteria as “helpers” for microalgae: a novel approach for removing ammonium and phosphorus from municipal wastewater. *Water Research*, 38 (2), 466–474. <https://doi.org/10.1016/j.watres.2003.09.022>
41. Patil, G. B., Lakshman, H. C., Mirdhe, R. M., & Agadi, B. S. (2013) Effect of co-inoculation of AM fungi and two beneficial microorganisms on growth and nutrient uptake of Eleusine coracana Gaertn. (Finger millet). *Asian Journal of Plant Science and Research*, 3 (1). 26–30.
42. Date, R. A. (2001). Advances in inoculant technology: a brief review. *Australian Journal of Experimental Agriculture*, 41 (3), 321-325.
43. Brockwell, J., Bottomley, P. J. (1995). Recent advances in inoculant technology and prospects for the future. *Soil Biology and Biochemistry*, 27 (4–5), 683-697. [https://doi.org/10.1016/0038-0717\(95\)98649-9](https://doi.org/10.1016/0038-0717(95)98649-9)
44. Kremer, R. J., Peterson, H. L. (1983). Effects of Carrier and Temperature on Survival of Rhizobium spp. in Legume Inocula: Development of an Improved Type of Inoculant. *Applied and Environmental Microbiology*, 45 (6), 1790–1794. <https://doi.org/10.1128/aem.45.6.1790-1794.1983>
45. Kovalevska, T. M., Kozar, S. F., Krutylo, D. V., Horban, V. P., Romanova, I. M., & Usmanova, T. O. (2015). Metody kultyvuvannya ta tryvaloho zberihannya bulbochkovykh bakterii v kolektsiakh: metodychni rekomendatsii [The method of cultivation of the trivial harvesting of bulbous bacteria in collections: methodical recommendations]. Chernihiv: ISMAV NAAN [in Ukrainian].
46. Hartley, E. J., Gemell, L. G., & Deaker, R. (2012). Some factors that contribute to poor survival of rhizobia on preinoculated legume seed. *Crop and Pasture Science*, 63 (8–9), 858–865. <https://doi.org/10.1071/CP12132>
47. Stout, D. G., Hall, J. W., Brooke, B. M., Baalim, G., Thompson, D. J. (1993). Effect of storage temperature and time on viability of rhizobia on lime-coated alsike clover (*Trifolium hybridum*) seed. *The Journal of Agricultural Science*, 120 (2), 205–211. <https://doi.org/10.1017/S0021859600074244>
48. Bashan, Y., Hernandez, J. P., Leyva, L. A., & Bacilio, M. (2002). Alginate microbeads as inoculant carriers for plant growth-promoting bacteria. *Biology and Fertility of Soils*, 35, 359–368. <https://doi.org/10.1007/s00374-002-0481-5>
49. Bashan, Y., Gonzalez, L. E. (1999). Long-term survival of the plant-growth-promoting bacteria *Azospirillum brasilense* and *Pseudomonas fluorescens* in dry alginate inoculant. *Applied Microbiology and Biotechnology*, 51 (2), 262–266. <https://doi.org/10.1007/s002530051391>
50. Kozar, S. F., Yevtushenko, T. A., & Nesterenko, V. M. (2017). Vplyv rehovyn riznoho khimichnoho skladu na zhyttiezdatnist diazotrofov na nasinni silskohospodarskykh kultur [The inflow of rehovins from a small chemistry warehouse to the life of diazotrophs on the national agricultural cultures]. *Silskohospodarska mikrobiologiya — Agricultural microbiology*, 25, 10–17 [in Ukrainian]. <https://doi.org/10.35868/1997-3004.25.10-17>
51. Mulder, L., Hogg, B., Bersoult, A., & Cullimore, J. V. (2005). Integration of signalling pathways in the establishment of the legume-rhizobia symbiosis. *Physiologia Plantarum*, 123 (2), 207–218. <https://doi.org/10.1111/j.1399-3054.2005.00448.x>

Received 05.04.2021

СТРАТЕГІЯ РЕГУЛЮВАННЯ АКТИВНОСТІ ДІАЗОТРОФІВ ЗА ЇХ ІНТРОДУКЦІЇ В АГРОЦЕНОЗИ СІЛЬСЬКОГОСПОДАРСЬКИХ КУЛЬТУР

С. Ф. Козар

Інститут сільськогосподарської мікробіології та агропромислового виробництва НААН, м. Чернігів
e-mail: kozarsf@gmail.com

Мета. Дослідити підходи щодо управління активністю ґрунтових діазотрофів та запропонувати стратегію її регулювання. **Методи.** Теоретичні, вегетаційного і польового дослідження, мікробіологічні, газохроматографічні, математично-статистичні. **Результати.** Показано, що активність корисних ґрунтових мікроорганізмів може змінюватися за дії температури, вологості, хімічних сполук різної природи, інших мікроорганізмів. Встановлено, що зважаючи на значну різноманітність чинників, необхідна розробка комплексу конкретних способів підвищення ростової й функціональної активності азотфіксувальних бактерій, а також їхньої життєздатності. Доведено, що за поєднання діазотрофів формується ефективна симбіотична бобово-ризобіальна система, що забезпечує надходження додаткового біологічного азоту в агроценози. Водночас відзначено збільшення маси рослин, вмісту хлорофілів у листках, вмісту протеїну та олії в продукції. За сумісного застосування діазотрофів збільшується урожайність, зокрема сої — на 9–16 %, якщо порівняти з інокуляцією чистою культурою бактерій. **Висновки.** На основі аналізу й узагальнення отриманих результатів досліджень пропонується стратегія регулювання активності діазотрофів для їх ефективної інтродукції в агроценози, яка полягає у поєднанні бактерій різних видів, підборі умов їх сумісного культивування й застосування за стабілізації чисельності життєздатних клітин бактерій. Запропонована стратегія передбачає розв'язання поставленого завдання шляхом отримання інокулянту, який характеризується високим титром і більш стабільною кількістю життєздатних клітин, що дозволяє отримати ефективну азотфіксувальну систему. Стратегія опрацьована на прикладі регулювання ростової й функціональної активності бульбочкових бактерій сої за рахунок поєднання діазотрофів різних видів, обґрунтування умов їх сумісного культивування і застосування для забезпечення позитивної взаємодії за формою коменсалізму, а також шляхом регулювання життєздатності діазотрофів за рахунок внесення добавок-стабілізаторів у середовище.

Ключові слова: азотфіксувальні бактерії, ростова активність, азотфіксація, життєздатність, стратегія регулювання активності діазотрофів.

ЦИТОВАНА ЛІТЕРАТУРА

1. Біологічний азот / За ред. В. П. Патики. Київ. 2003. 424 с.
2. Мікробні препарати в землеробстві. Теорія і практика / За ред. В. В. Волкогона. Київ. 2006. 312 с.
3. Волкогон В. В. Мікробіологічні аспекти оптимізації азотного удобрення сільськогосподарських культур. К. : Аграрна наука, 2007. 144 с.
4. Моргун В. В., Коць С. Я. Роль біологічного азоту в азотному живленні рослин. *Вісник Національної Академії Наук України*. 2018. № 1. С. 62–74.
5. Курдиш И. К., Мельникова Н. Н. Влияние глинистых минералов на рост и нодуляционную активность *Bradyrhizobium japonicum*. *Мікробіологічний журнал*. 2011. Т. 73 (4). С. 36–40.
6. Stephens J. H. G., Rask H. M. Inoculant production and formulation. *Field crops research*. 2000. Vol. 65. С. 249–258. [https://doi.org/10.1016/S0378-4290\(99\)00090-8](https://doi.org/10.1016/S0378-4290(99)00090-8)
7. Date R. A. Advances in inoculant technology: a brief Review. *Australian journal of experimental agriculture*. 2001. Vol. 41 (3). С. 321–325. <https://doi.org/10.1071/EA00006>
8. Brockwell J., Bottomley P. J. Recent advances in inoculant technology and prospects for the future. *Soil Biology and Biochemistry*. 1995. Vol. 27 (4–5). С. 683–697. [https://doi.org/10.1016/0038-0717\(95\)98649-9](https://doi.org/10.1016/0038-0717(95)98649-9)
9. Crawford S. L., Berryhill D. L. Survival of

- Rhizobium phaseoli in coal-based Legume inoculants applied to seeds. *Applied and environmental microbiology*. 1983. Vol. 45 (2). С. 703–705. <https://doi.org/10.1128/aem.45.2.703-705.1983>
10. Argaw A. Evaluation of co-inoculation of Bradyrhizobium japonicum and phosphate solubilizing Pseudomonas spp. Effect on soybean (Glycine max L. (Merr.)) in Assossa Area. *Journal of agricultural science and technology*. 2012. Vol. 14. С. 213–224.
11. Askary M., Mostajeran A., Amooaghaei R., Mostajeran M. Influence of the co-inoculation Azospirillum brasilense and Rhizobium meliloti plus 2,4-D on grain yield and N, P, K content of Triticum aestivum (cv. Baccros and Mahdavi). *American-Eurasian Journal of Agricultural & Environmental Sciences*. 2009. Vol. 5 (3). С. 296–307.
12. Воробей Н. А., Пацко О. В., Коць С. Я., Паршикова Т. В. Фізіологічні особливості розвитку люцерни за інокуляції змішаними культурами азотфіксувальних мікроорганізмів. *Фізіологія і біохімія культурних рослин*. 2009. Т. 41 (4). С. 344–352.
13. Aung T. T., Tittabutr P., Boonkerd N., Heridge D., Teaumroong N. Co-inoculation effects of Bradyrhizobium japonicum and Azospirillum sp. on competitive nodulation and rhizosphere eubacterial community structures of soybean under rhizobia-established soil conditions. *African Journal of Biotechnology*. 2013. Vol. 12 (20). С. 2850–2862.
14. Муродова С. С., Давранов К. Д. Комплексные микробные препараты. Применение в сельскохозяйственной практике. *Biotechnologia Acta*. 2014. Vol. 7 (6). С. 92–101.
15. Доспехов Б. А. Методика полевого опыта (с основами статистической обработки результатов исследований). М. : Агропромиздат. 1985. 351 с.
16. Шлегель Г. Общая микробиология. М. : Мир, 1987. 567 с.
17. Практикум по микробиологии / под ред. В. К. Шильникова. М. Дрофа, 2005. 256 с.
18. Быков В. А., Винаров А. Ю., Шерстобитов В. В. Расчет процессов микробиологических производств. К. : Техника. 1985. 246 с.
19. Перт С. Д. Основы культивирования микроорганизмов и клеток. М. : Мир, 1978. 330 с.
20. Hardy R. W. F., Hotsren R. D., Jackson E. K., Burns R. C. The acetylene-ethylene assay for N₂ fixation: laboratory and field evaluation. *Plant Physiol*. 1968. Vol. 43 (8). С. 1185–1207. <https://doi.org/10.1104/pp.43.8.1185>
21. Hardy R. W. F., Burns R. C., Holsten, R. D. Applications of the acetylene-ethylene assay for measurement of nitrogen fixation. *Soil Biology and Biochemistry*. 1973. Vol. 5 (1). С. 47–81.
22. Експериментальна ґрунтова мікробіологія / За ред. В. В. Волкогона. К. : Аграрна наука, 2010. 464 с.
23. Kozar S. F., Symonenko E. P., Lynnyk V. O., Kaplunenko V. H. Influence of nanocarboxylates of microelements on Rhizobium radiobacter 204 growth-regulating activity. *Сільськогосподарська мікробіологія*. 2015. Вип. 22. С. 3–8. <https://doi.org/10.35868/1997-3004.22.3-8>
24. Євтушенко Т. А., Козар С. Ф., Пищур І. М. Вплив нанокарбоксилатів мікроелементів на ріст і функціональну активність Bradyrhizobium japonicum. Мікробіологія в сучасному сільськогосподарському виробництві: тези доповіді ХІ наукової конференції молодих вчених (м. Чернігів, 5–6 жовтня). Чернігів, 2018. С. 20–22.
25. Козар С. Ф., Євтушенко Т. А., Усманова Т. О. Вплив полісахаридно-білкового комплексу на ефективність бактеризації сої Ризогуміном. *Сільськогосподарська мікробіологія*. 2019. Вип. 30. С. 13–19. <https://doi.org/10.35868/1997-3004.30.13-19>
26. Kozar S. F., Symonenko E. P., Volkohon V. V., Volkogon M. V. Nanocarboxylates of molybdenum and of iron enhance the functional activity of Rhizobium radiobacter 204. *Applied Nanoscience*. 2019. Vol. 9. Num. 5. С. 795–800. <https://doi.org/10.1007/s13204-018-00939-6>
27. Козар С. Ф. Антагоністичні відносини бацил із представниками ґрунтових діазотрофів та перспективи використання їхніх комплексів при вирощуванні сільськогосподарських культур. *Вісник аграрної науки Причорномор'я*. 2003. № 3 (1). С. 302–307.
28. Козар С. Ф., Ушакова М. А. Антагонізм азотобактера щодо ґрунтових діазотрофів та вплив комплексів бактерій на ріст рослин. *Агро-екологічний журнал*. 2004. № 2. С. 31–34.
29. Козар С. Ф. Комплексне застосування діазотрофів для підвищення урожайності озимого жита. *Збірник наукових праць Уманського державного аграрного університету: основи формування продуктивності сільськогосподарських культур за інтенсивних технологій вирощування*. 2008. С. 312–316.
30. Козар С. Ф., Ушакова М. А., Погорилько Н. В. Створення штучних асоціацій діазотрофів для інокуляції озимого жита. Сталий розвиток агроєкосистем: тези доповіді міжнародної наукової конференції (м. Вінниця, 17–20 вересня). Вінниця, 2002. С. 105–107.
31. Козар С. Ф. Біологічна ефективність комплексного застосування мікробних препаратів. *Сільськогосподарська мікробіологія*. 2005. Вип. 1–2. С. 86–94.
32. Козар С. Ф. Вплив мікробних препаратів на активність мікрофлори ризосферного ґрунту ярого ячменю. *Вісник ХНАУ ім. В. В. Докучаєва*. 2006. 4. С. 126–133.
33. Козар С. Ф. Биологические элементы

технології вирощування ярового ячменя на основі комплексного використання мікробних препаратів. *Чернігівський державний центр науково-технічної і економічної інформації*. 2005. № 4. 4 с.

34. Козар С. Ф., Нестеренко В. М., Євтушенко Т. А., Фіровський О. В., Усманова Т. О. Ефективність застосування мікробного препарату АБТ в технології вирощування цибулі ріпчастої. *Науковий вісник Національного університету біоресурсів і природокористування України (Серія «Агрономія»)*. 2013. Вип. 183 (1). С. 207–214.

35. Bilokonska O., Kozar S. Viability of *Azotobacter chroococcum* IMB B-7836 and their influence on cucumber productivity. *Analele Universității din Oradea, Fascicula Biologie*. 2020; XXVII, Issue 2. С. 111–115.

36. Білоконська О. М., Козар С. Ф., Євтушенко Т. А., Усманова Т. О. Збереженість бактерій роду *Azotobacter* на насінні *Cucumis sativus* L. за дії різних температур. *Сільськогосподарська мікробіологія*. 2018. Вип. 27. С. 11–17. <https://doi.org/10.35868/1997-3004.27.11-17>

37. Goldenfeld N., Woese C. Biology's Next Revolution. *Nature*. 2007. Vol. 445. С. 369. <https://doi.org/10.1038/445369a>

38. Патица М. В., Танчик С. П., Колодяжний О. Ю., Іванюк М. Ф., Круглов Ю. В., Мельничук М. Д., Патица Т. І. Формування біорізноманіття та філотипової структури еубактеріального комплексу чорнозему типового при вирощуванні пшениці озимої. *Доповіді Національної академії наук України*. 2012. № 11. С. 163–171.

39. Современная микробиология. Прокариоты: в 2-х томах / Под ред. Й. Ленгелера, Г. Дрекса, Г. Шлегеля. М.: Мир. 2005.

40. De-Bashan L. E., Hernandez J., Morey T., Bashan Y. Microalgae growth-promoting bacteria as “helpers” for microalgae: a novel approach for removing ammonium and phosphorus from municipal wastewater. *Water Research*. 2004. № 38 (2). С. 466–474. <https://doi.org/10.1016/j.watres.2003.09.022>

41. Patil G. B., Lakshman H. C., Mirdhe R. M., Agadi B. S. Effect of co-inoculation of AM fungi and two beneficial microorganisms on growth and nutrient uptake of *Eleusine coracana* Gaertn. (Finger millet). *Asian Journal of Plant Science and Research*. 2013. Vol. 3 (1). С. 26–30.

42. Date R. A. Advances in inoculant technology: a brief review. *Australian Journal of Experi-*

mental Agriculture. 2001. Vol. 41 (3). С. 321–325.

43. Brockwell J., Bottomley P. J. Recent advances in inoculant technology and prospects for the future. *Soil Biology and Biochemistry*. 1995. Vol. 27 (4–5). С. 683–697. [https://doi.org/10.1016/0038-0717\(95\)98649-9](https://doi.org/10.1016/0038-0717(95)98649-9)

44. Kremer R. J., Peterson H. L. Effects of Carrier and Temperature on Survival of *Rhizobium* spp. in Legume Inocula: Development of an Improved Type of Inoculant. *Applied and Environmental Microbiology*. 1983. Vol. 45 (6). С. 1790–1794. <https://doi.org/10.1128/aem.45.6.1790-1794.1983>

45. Ковалевська Т. М., Козар С. Ф., Крутило Д. В., Горбань В. П., Романова І. М., Усманова Т. О. Методи культивування та тривалого зберігання бульбочкових бактерій в колекціях: методичні рекомендації. Чернігів: ІСМАВ НААН. 2015. 36 с.

46. Hartley E. J., Gemell L. G., Deaker R. Some factors that contribute to poor survival of rhizobia on preinoculated legume seed. *Crop and Pasture Science*. 2012. Vol. 63 (8–9). С. 858–865. <https://doi.org/10.1071/CP12132>

47. Stout D. G., Hall J. W., Brooke B. M., Baalim G., Thompson D. J. Effect of storage temperature and time on viability of rhizobia on lime-coated alsike clover (*Trifolium hybridum*) seed. *The Journal of Agricultural Science*. 1993. Vol. 120 (2). С. 205–211. <https://doi.org/10.1017/S0021859600074244>

48. Bashan Y., Hernandez J. P., Leyva L. A., Bacilio M. Alginate microbeads as inoculant carriers for plant growth-promoting bacteria. *Biology and Fertility of Soils*. 2002. Vol. 35. С. 359–368. <https://doi.org/10.1007/s00374-002-0481-5>

49. Bashan Y., Gonzalez L. E. Long-term survival of the plant-growth-promoting bacteria *Azospirillum brasilense* and *Pseudomonas fluorescens* in dry alginate inoculant. *Applied Microbiology and Biotechnology*. 1999. Vol. 51 (2). С. 262–266. <https://doi.org/10.1007/s002530051391>

50. Козар С. Ф., Євтушенко Т. А., Нестеренко В. М. Вплив речовин різного хімічного складу на життєздатність діазотрофів на насінні сільськогосподарських культур. *Сільськогосподарська мікробіологія*. 2017. Вип. 25. С. 10–17. <https://doi.org/10.35868/1997-3004.25.10-17>

51. Mulder, L., Hogg, B., Bersoult, A., Cullimore, J. V. Integration of signalling pathways in the establishment of the legume-rhizobia symbiosis. *Physiologia Plantarum*. 2005. Vol. 123 (2). P. 207–218. <https://doi.org/10.1111/j.1399-3054.2005.00448.x>

Отримано 05.04.2021